The Characterization of Salmonella isolated from Pig Meat in Northern Ireland by PFGE and Antibiotic Resistance Profiles

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Abstract

The emergence of antibiotic resistance and especially multiple antibiotic resistance Salmonella has become a concern for the pig industry throughout the EU. Pig herds and pork are considered as principal reservoirs for the multi-resistant Salmonella type Typhimurium DT104, which has acquired resistance to ampicillin, chloramphenicol, streptomycin, sulphonamides and tetracycline. This resistance pattern is also known as ACSSuT. In this study Porcine Salmonella strains were isolated between December 2005 and December 2006. The strains originated from an abattoir study sampling the 'oyster' cut. The Antimicrobial Resistance Profiles of all Salmonella isolates in this study were determined by disk diffusion tests. Twelve antibiotics were utilized throughout the profiling procedure. Pulsed Field Gel Electrophoresis (PFGE), which is regarded as the "Gold Standard" for the typing and strain identification of Salmonella isolates, was used to determine DNA fingerprints of the Salmonella isolates using the restriction enzyme XbaI (Invitrogen). The fragments were then separated by PFGE in a Chef DR II system (Bio-Rad). This enabled comparison of Salmonella isolated in this study.

Introduction

There are over 2,300 serotypes of Salmonella worldwide (Foley et al., 2001, 2000). In 2005 there were 176 reported cases of human salmonellosis in Northern Ireland. This has been the lowest level reported in the past 12 years, but in 2004 there were three major outbreaks of Salmonella in the Northern Ireland involving S. typhimurium DT104, S. virchow and S. newport (CDSC, 2006). These outbreaks are a reminder of how important it is to monitor and control Salmonella in our food animals. Over the past five years Salmonella typhimurium has consistently been the second most common cause of human salmonellosis. S. typhimurium type DT104 has also been the most commonly isolated Salmonella on the island of Ireland and in Great Britain (CDSC, 2006)(Randall et al., 2003) (Foley et al., 2000). There were 33 reported cases of Salmonella typhimurium in Northern Ireland alone, of which the most frequently reported phage type was Salmonella typhimurium DT104 (CDSC, 2006). It has been established that multi-drug resistant S. typhimurium DT104 has acquired antibiotic resistance to ampicillin, chloramphenicol, streptomycin, sulphonamides and tetracycline's. This resistance profile has been abbreviated to ACSSuT resistance (Alban et al., 2001) (Threlfall, 2000). An abattoir study undertaken in the UK in 2003 to determine the carriage of food borne pathogens in food animals reported the prevalence of Salmonella carriage at slaughter was 23.4% for pigs (DEFRA, 2004). Studies in The Netherlands into the distribution of sero- and phage types of Salmonella strains found that the serovar Typhimurium was the predominant serovar in pigs between the years 1984 and 2001 (Duijkeren et al., 2002). This information suggests that pigs may be one of the main reservoirs of Salmonella and more importantly a reservoir of the multi-drug resistant S. typhimurium definitive phage type DT104. Wide spread antibiotic resistance has become a serious public health risk over the last decade and multi antibiotic resistance has been reported in many food borne pathogens including Salmonella. Antibiotic resistance profiles for strains of Salmonella spp. isolated from the pork samples in Northern Ireland were determined using an agar disk diffusion technique.
The aim of this research is to:

1. Assess antibiotic resistance among *Salmonella* spp. isolated from pig meat in Northern Ireland.
2. Generate PFGE macro-restriction fingerprints of the *Salmonella* spp. isolated from pig meat in Northern Ireland. PFGE is regarded as the “Gold Standard” for the typing and strain identification of *Salmonella* isolates (Brown et al., 2006) (Tamada et al., 2001).

**Material and methods**

All *Salmonella* isolates were examined by the “Modified Stoke’s Technique” a disc diffusion method testing isolates for sensitivity to 12 antibiotics: Amikacin (AK 30μg), Ampicillin (AMP 10μg), Apramycin (APR 15μg), Cefotaxime (CTX 30μg), Ceftazidime (CAZ 30μg), Chloramphenicol C 10μg, Ciprofloxacin (CIP 1μg), Compound Sulphonamides (S3 300μg), Furazolidone (FR 15μg), Streptomycin (S 25μg), Sulphamethoxazole/trimethoprim (SXT 25μg) and Tetracycline (TE 10μg). PFGE macro-restriction fingerprint images are created using the PulseNet protocols (CDC, 2006). This molecular technique is used to create an individual DNA fingerprints for all *Salmonella* isolates.

**Results**

Antibiotic resistance was demonstrated in 29 isolates. The antibiotics with least effect were Tetracycline (74%), Streptomycin (29%), Sulphamethoxazole/trimethoprim (26%), Compound Sulphonamides (26%) and Ampicillin (11%). Two of the *S. typhimurium* isolates displayed the ACSSuT phenotype associated with multidrug resistant *S. typhimurium* DT 104. These two *Salmonella* isolates also displayed resistance to Sulphamethoxazole/trimethoprim.

**Discussion**

In this study the antibiotic resistance profiles for strains of *Salmonella* spp. isolated from the pork samples in Northern Ireland were determined using the Modified Stokes technique. Comparison of the antibiotic resistance of the salmonellae tested in this project suggested that antibiotic resistance was observed in 76% of all serovars tested. A high resistance to tetracycline (74%) was observed, this result may be attributed to tetracycline being the most common therapeutic drug used by the pork industry (Burch, 2005). The second most common therapeutic antibiotic used in the pig industry a sulphonamethoxazole and trimethoprim (Burch, 2005) which also exhibited a high antibiotic resistance profile, with over a quarter (26%) of all the *Salmonella* isolated from pork in Northern Ireland displaying resistance. Ampicilllin and apramycin are also commonly used therapeutically throughout the pig industry (Burch, 2005) (DEFRA, 2007). In all 18 of the 19 S. *rissen* isolates were resistant to tetracycline. This result was supported by a Spanish study that concluded that a S. *rissen* isolated from pork harboured the tet(A) resistant gene (Ioana et al., 2006). A high level of intermediate resistance was observed in 6 out of the 12 antibiotics analysed in this study. This may suggest an increase in the number of antibiotic resistant *Salmonella* spp. in years to come. PFGE has become a valuable tool for the epidemiological typing of all *Salmonella* including *S. typhimurium*. The majority of multi-resistant DT104 has the distinctive XbaI generated macrorestriction fingerprint that can be detected by PFGE (Threlfall, 2000) (Doran et al., 2005). PFGE images were generated for all the *Salmonella* isolates in this project.

**Conclusion**

76% of *Salmonella* spp. isolated in this study had resistance to one or more of the antibiotics tested. These results highlight the need for continual monitoring and control of *Salmonella* in food animals.
References

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