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Methane Flux in Cropland and Adjacent Riparian Buffers with Different Vegetation Covers

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While water quality functions of conservation buffers established adjacent to cropped fields have been widely documented, the relative contribution of these re-established perennial plant systems to greenhouse gases has not been completely documented. In the case of methane (CH$_4$), these systems have the potential to serve as sinks of CH$_4$ or may provide favorable conditions for CH$_4$ production. This study quantifies CH$_4$ flux from soils of riparian buffer systems comprised of three vegetation types and compares these fluxes with those of adjacent crop fields. We measured soil properties and diel and seasonal variations of CH$_4$ flux in 7 to 17 yr-old re-established riparian forest buffers, warm-season and cool-season grass filters, and an adjacent crop field located in the Bear Creek watershed in central Iowa. Forest buffer and grass filter soils had significantly lower bulk density ($P < 0.01$); and higher pH ($P < 0.01$), total carbon (TC) ($P < 0.01$), and total nitrogen (TN) ($P < 0.01$) than crop field soils. There was no significant relationship between CH$_4$ flux and soil moisture or soil temperature among sites within the range of conditions observed. Cumulative CH$_4$ flux was $–0.80$ kg CH$_4$–C ha$^{-1}$ yr$^{-1}$ in the cropland, $–0.46$ kg CH$_4$–C ha$^{-1}$ yr$^{-1}$ within the forest buffers, and $0.04$ kg CH$_4$–C ha$^{-1}$ yr$^{-1}$ within grass filters, but difference among vegetation covers was not significant. Results suggest that CH$_4$ flux was not changed after establishment of perennial vegetation on cropped soils, despite significant changes in soil properties.

The global atmospheric concentration of CH$_4$ has increased from a preindustrial value of about 715 to 1774 μL L$^{-1}$ in 2005, likely a result of anthropogenic activities such as agricultural production and fossil fuel use (IPCC, 2007). Soils have been shown to both produce and consume CH$_4$ (Topp and Pattey, 1997; Le Mer and Roger, 2001). In a recent review, Dutaur and Verchot (2007) summarized net CH$_4$ flux as the result of the balance between the two offsetting processes of methanogenesis (microbial production under anaerobic conditions) and methanotrophy (microbial consumption). These authors identified methanotrophy as the dominant process in upland soils, where oxidation generally exceeds production with a resulting net uptake of atmospheric CH$_4$ by soil. It is well known that forest soils are the most active sink of CH$_4$, followed by grass lands and cultivated soils, and that the CH$_4$ uptake potential of many upland soils is reduced by cultivation and application of ammonium N fertilizer (Topp and Pattey, 1997; Le Mer and Roger, 2001; Dutaur and Verchot, 2007). It has been reported that land-use change can also influence CH$_4$ uptake rates. For instance, higher rates of CH$_4$ oxidation have been observed in soils afforested from croplands or pastures (e.g., Ball et al., 2002; Merino et al., 2004; Tate et al., 2007). Observed increases in CH$_4$ uptake resulting from land-use change are attributed to changes in soil porosity, moisture content, and methanotroph population (Priemé et al., 1997).

Nonpoint source (NPS) pollutants such as sediment, N, P, and pesticides are major causes of water quality problems around the world (Duda, 1993; Tonderski, 1996; Sabater et al., 2003). Riparian buffers have been recommended as one of the most effective tools for mitigating NPS pollution (Hubbard et al., 2004; Mayer et al., 2007). Some of the important functions of riparian buffers related to NPS pollution control are filtering and retaining sediment and immobilizing, storing, and transforming chemical inputs from uplands (Schultz et al., 2000). Generally, riparian buffers re-established on cultivated crop fields consist of combinations of grasses, forbs, shrubs, and trees (Schultz et al., 2004). While these systems have been well documented for their water quality functions, little is known about other ecosystem processes such as their relative greenhouse gas flux. If these systems perform similar to perennial plant systems in upland positions, it would suggest that

**Abbreviations:** IPCC, Intergovernmental Panel on Climate Change; NPS, nonpoint source.
riparian buffers re-established on cropped soils may produce less and consume more CH₄ than crop fields. However, riparian buffers are often flooded and also sustain relatively high soil moisture conditions caused by high water tables, long residence time and slow discharge (Schultz et al., 2000). These conditions may be favorable for CH₄ production. For example, Ambus and Christensen (1995) reported that CH₄ was produced in temporarily flooded riparian areas at rates of 78.8 kg CH₄-C ha⁻¹ yr⁻¹. Methane was also produced from riparian areas of ponded depressions in northern Germany at rates of 0.33 to 330.3 kg CH₄-C ha⁻¹ yr⁻¹ (Merbach et al., 1996). In contrast, rates of CH₄ consumption in temperate regions have been estimated as 1.29 ± 0.16 kg CH₄ ha⁻¹ yr⁻¹ in crop fields (n = 48), 5.75 ± 0.59 kg CH₄ ha⁻¹ yr⁻¹ in grasslands (n = 24), and 2.40 ± 0.40 kg CH₄ ha⁻¹ yr⁻¹ in forests (n = 91) (data extracted from Dutaur and Verchot (2007)). These results suggest that reestablished riparian buffers may produce more CH₄ than crop fields and natural lands, at least when they are flooded, and the benefits of reduced nonpoint-source pollution from riparian buffers may offset by increased greenhouse gas emissions.

Numerous studies have emphasized the role of vegetation in soil biogeochemical processes within natural or re-established riparian buffers, with many studies demonstrating an improvement in soil quality indicators (e.g., Tufekcioglu et al., 1999; Bharati et al., 2002; Marquez et al., 2004). However, most studies have often found conflicting results regarding the effect of vegetation type on biogeochemical process regulation. For example, there are uncertainties about the effect of vegetation type on groundwater 
NO₃⁻ removal or denitrification in riparian buffers (e.g., Groffman et al., 1991; Schnabel et al., 1996; Hefting et al., 2003). With respect to CH₄ flux, several studies have compared rates among vegetation types (Topp and Pattey, 1997; Le Mer and Roger, 2001; Chan and Parkin, 2001a, 2001b; Dutaur and Verchot, 2007). However, few studies of CH₄ flux have focused on riparian soils, particularly those re-established to perennial vegetation, or on the relationship between observed changes in soil quality on conditions regulating methane flux. Specific objectives of this study were to compare CH₄ flux from riparian buffer systems comprised of forest, warm-season grasses, and cool-season grasses and an adjacent crop field, and to relate these fluxes to changes in soil properties after re-establishment of perennial plants.

**Materials and Methods**

**Study Site**

The study area consisted of three forest buffers, three warm-season grass filters, one cool-season grass filter, and one adjacent crop field located in the Bear Creek watershed, Story County and Hamilton County, Iowa (42° 11’ N, 93° 30’ W). The Bear Creek watershed (6810 ha) is a predominantly agricultural watershed typical of north central Iowa, with a mean annual air temperature of 8.7°C and mean annual precipitation of 810 mm over the period of record (United States Department of Commerce—National Oceanic and Atmospheric Administration, 2009). Most of the area was originally covered with prairie and wetland vegetation except for riparian forests along higher order streams. Most of the area is now rain-fed agriculture cultivated with soybean [Glycine max (L.) Merr.] and corn (Zea mays L.), which are usually grown in rotation. Re-established forest buffers, and warm-season and cool-season grass filters were previously under row-crop cultivation or livestock grazing. The forest buffers and grass filters ranged in age from 7 to 17 yr since re-establishment.

Forest buffers included the following tree and shrub species: silver maple (Acer saccharinum L.), green ash (Fraxinus pennsylvanica Marsh.), black walnut (Juglans nigra L.), willow (Salix spp.), cottonwood hybrids (Populus spp.), red oak (Quercus rubra L.), and bur oak (Q. bicolor Willd.). Shrub species included chokecherry (Prunus virginiana L.), Nanking cherry (P. tomentosa Thunb), wild plum (P. americana Marsh), red osier dogwood (Cornus stolonifera Michx.), and ninebark (Physocarpus opulifolius Maxim.). Warm-season grass filters included native species such as switchgrass (Panicum virgatum L.), Indian grass (Sorghastrum nutans Nash), and big bluestem (Andropogon gerardii Vitman). Numerous forb species were present, including purple prairie clover (Petalostemum purpureum Vent.), black-eyed susan (Rudbeckia hirta L.), yellow coneflower (Ratibida pinnata Vent.), stiff goldenrod (Solidago rigida L.), prairie blazing star (Listaria cynrnostachya Michx.), and others. The cool-season grass buffers were dominated by nonnative forage grasses (Bromus inermis Leysser, Phleum pratense L., and Poa pratensis L.). Details of the riparian buffer design, placement, and plant species are given in Schultz et al. (1995). The crop fields adjacent to the riparian buffers served as a control, representing conditions before buffer establishment. The crop fields were planted to soybean in 2007. Pelletized urea (134 kg N ha⁻¹) was applied during corn rotation years and cultivation consists of fall chisel plowing (15–20 cm depth). All areas used in this study were located on Coland soil (fine-loamy, mixed, superactive, mesic Cumulic Endoaquoll) which is well- to poorly drained and formed from till or local alluvium and colluvium derived from till (DeWitt, 1984).

**Soil Sampling and Analysis**

Six intact soil cores (5.3-cm diam.) were collected to a depth of 15 cm around each of three gas sampling points in a forest buffer, a warm-season grass filter, a cool-season grass filter, and an adjacent crop field in October 2006 and September 2007. A plastic sleeve liner was placed inside the metal core tube and the liner and intact soil core pulled from the tube and capped for transport to the laboratory. Soil samples were transported back to lab in a cooler and stored at 4°C until analysis. Soil pH was determined by using 1:1 diluted soil solution. Gravimetric moisture content was determined by oven drying a subsample at 105°C for 24 h. Bulk density was estimated using the core method (Grossman and Reinsch, 2002). For C and N analysis, soils were air dried at room temperature, and sieved (2 mm). Total C and TN were measured using a Flash EA 2000 (ThermoFinnigan, Italy) elemental analyzer. Soil inorganic N was extracted with 2 mol L⁻¹ potassium chloride (KCl) within 4 h of sampling and stored at 4°C until filtration (Van Miegroet, 1995). Filtrates were frozen and stored until analysis. Nitrate (NO₃⁻) and ammonium (NH₄⁺) contents were analyzed by colorimetric method (Mulyaneya, 1996) with an auto analyzer (Quikchem 8000 FIA+, Lachat Instruments, Milwaukee, WI).
Field Gas Sampling, Methane Gas Analysis, and Flux Calculation

Soil CH₄ flux from riparian forest buffers, warm- and cool-season grass filters and one crop field was measured from January through December 2007. To assess the temporal variation of flux, five locations were randomly selected in each of three forest buffers, three warm-season grass filters, one cool-season grass filter, and one crop field with the distance between gas sampling points ranging from 5 to 10 m. A polyvinyl chloride (PVC) ring (30 cm diam. by 15 cm height) served as base for gas chambers and was installed to a depth of approximately 10 cm. In the crop field, rings were placed either between plants within the row or between rows. These rings were left in place between sampling periods, but were removed for fertilization, planting, and tillage events in the crop field. Vegetable inside the rings was cut before gas sampling in the forest buffers and grass filters during growing seasons. Gas samples were collected within static vented chambers (PVC, 30 cm diam. by 15 cm height with a vent) weekly or biweekly during the mid-morning. Chambers were equipped with a thermometer to measure air temperature within the chambers at the time of sampling. Ten milliliters of air was sampled from the chamber with a polypropylene syringe at 15-min intervals for 45 min (four samples 0, 15, 30, and 45 min) and the gas stored in pre-evacuated glass vials (6 mL, fitted with butyl rubber stoppers) until analysis. Glass vials were prepared by alternately evacuating the vial headspace and flushing with helium to remove air. In addition to the regular measurements, diel variation of CH₄ flux was measured on 16–17 July 2007. For this assessment, three gas sampling points were randomly selected in a forest buffer, a warm-season grass filter, a cool-season grass filter, and an adjacent crop field and gas samples were collected every 3 h for 24 h. Methane concentrations were determined with a gas chromatograph (GC) (Model GC17A; Shimadzu, Kyoto, Japan) equipped with a flame ionization detector (FID) and a stainless steel column (0.3175 cm diam. × 74.54 cm long) with Porapak Q (80–100 mesh). Samples were introduced into the gas chromatograph using an autosampler described by Arnold et al. (2001). Three different CH₄ standards (0, 2.0, and 10 μL L⁻¹) were used to perform calibration curves and field ambient samples and CH₄ standards were analyzed every 20 gas samples to verify accuracy in GC results. Methane fluxes were obtained by applying linear regression to the CH₄ concentration vs. time data (Holland et al., 1999). Linearity with R² > 0.8 was accepted as a valid flux rate, which resulted in the inclusion of 90% of the flux rates in this study. Where removing a sample corrected a poor linearity (R² < 0.8) to R² > 0.9, the sample was eliminated from the calculation of flux rate (Altor and Mitsch, 2006). Therefore, flux rate was determined using a minimum of three gas samples.

The minimum detectable CH₄ flux was calculated using an average of standard deviations of CH₄ concentrations of lab ambient air samples and CH₄ standards (n = 500) analyzed with collected gas samples, chamber volume, chamber footprint, and chamber enclosure time in the fields as following:

\[
\text{Minimum detectable } \text{CH}_4 \text{ flux } (\mu \text{L CH}_4 \text{C m}^{-2} \text{ h}^{-1}) = \frac{2 \times \text{average of standard deviation(μL L}^{-1}) \times \text{chamber volume (L)}}{\text{chamber footprint (m}^2) \times \text{chamber enclosure time (h)}} \quad [1]
\]

The calculated minimum detectable CH₄ flux (μL CH₄C m⁻² h⁻¹) converted to mass unit (μg CH₄C m⁻² h⁻¹) through application of the universal gas law (Holland et al., 1999). Our estimated minimum detectable flux was 33.2 μg CH₄C m⁻² h⁻¹. Some of the fluxes measured from the individual chambers were smaller than our detection limit. In these situations, we followed the recommendation of Gilbert (1987) and Chan and Parkin (2001a) and included the measured values of these “nondetects” in computing mean fluxes. Cumulative CH₄ fluxes from each site over the 1-d study period (16–17 July 2007) and the 1-yr study period (January–December 2007) were calculated by linear interpolation and numerical integration between sampling times.

Soil temperature (ST) and soil water content (SWC) were measured simultaneously with CH₄ gas collection around the chamber at a 5 cm depth using a digital thermocouple (ThermoWorks, Orem, UT) and a digital soil moisture meter (HydroSense, Campbell Scientific, Inc., Logan, UT). Air temperature was measured simultaneously with CH₄ gas collection inside and outside the gas chamber. A soil temperature and soil moisture data logger (HOBO Micro station data logger with sensors, Onset Computer Corporation, Bourne, MA) was installed at 5 cm soil depths around a chamber at each site to measure hourly ST and SWC at each site. Daily rainfall and snow data were provided by a nearby meteorology station (Colo, IA, 42° 1’N, 93° 19’W).

Soil Incubation with Control and 10 Pa Acetylene (C₂H₂)

Aerobic CH₄ production and net CH₄ flux were estimated using the intact soil cores (0–15 cm depth) collected in September 2007. Soil samples were transported in a cooler and stored at 4°C until experiments, and incubation experiments with the intact soil cores were conducted within 6 h of sampling. All aboveground vegetation in the soil cores was cut off before the experiments. Six intact soil cores (5.3 cm diam. by 8 cm long) collected at each site were placed into 350-mL glass jars with gas-tight lids containing a gas-sampling port and all jars were sealed. Gravimetric moisture content of each soil was determined by oven drying a subsample at 105°C for 24 h. Three soil cores from each site were treated with 10 Pa C₂H₂ and three were retained as controls (no C₂H₂). Soil cores were incubated at 22°C, the on-site soil temperature. Ten milliliters of air was sampled from the jars with a polypropylene syringe at 3, 9, and 16 h, and stored until analysis. Storage, gas analysis, and flux calculations were as described above. Aerobic CH₄ production was estimated from soil incubations in which CH₄ oxidation was inhibited by 10 Pa C₂H₂ (Chan and Parkin, 2000, 2001b). Net CH₄ flux was determined from CH₄ flux in soil incubations without C₂H₂.

Aerobic CH₄ production = CH₄ flux under 10 Pa C₂H₂ [2]

Net CH₄ flux = CH₄ flux under no C₂H₂ [3]

Statistical Analyses

The Shapiro-Wilk normality test was used to assess normality of data. A two-sample t test was used to evaluate differences in soil C measured in 1998–1999 and 2006–2007 in the same sites. One-way ANOVA was used to evaluate the differences in soil properties, and diel and seasonal CH₄ flux by site. When the
standard assumptions of normality were violated, nonparametric Kruskal–Wallis one-way ANOVA on ranks was used. Differences were considered significant at the $P < 0.05$ level. To determine the relationship between soil properties and CH$_4$ flux, correlation analysis using the GLM procedure was applied. Statistical analyses were conducted using SAS ver 8.1 (SAS Institute, 1999).

**Results**

**Soil Properties**

The texture of all treatment site soils (Coland) was loam (Marquez et al., 2004) (Table 1). Soils within all riparian buffer vegetation types had significantly lower bulk density (one-way ANOVA $P < 0.01$); higher pH ($P < 0.01$), and NH$_4^+$ ($P < 0.01$) than crop field soils. TC ($P < 0.01$) and TN ($P < 0.01$) within the pooled riparian buffer vegetation soils were significantly higher that crop field soils. However, this difference is apparently driven by the forest buffer soils, as indicated in pairwise comparisons. Soil NO$_3^-$ was not significantly different among sites (Table 1). Within the same sites, soil carbon content (0–15 cm soil depth) was 30.4 ± 1.6 g C kg soil$^{-1}$ (n = 6) in the forest buffer, 24.4 ± 1.0 g kg$^{-1}$ (n = 6) in the warm-season grass filter, and 31.0 ± 1.8 g kg$^{-1}$ (n = 6) in the cool-season grass filter in 1998 and 1999 (J. Raich, unpublished data, 1999). Comparing these data with those of this study (Table 1), soil C in the forest buffer (42.9 ± 3.2 g kg$^{-1}$, n = 6) in this study was significantly higher than those in 1998 and 1999 (two sample t test $P = 0.006$, 95% CI for difference of means: 4.5 – 20.5 g kg$^{-1}$).

From 15 June to 15 August 2007 (93 d), average daily soil moisture in the crop field (8.7 ± 0.2%, n = 93), was significantly lower (one-way ANOVA $P < 0.01$) than soils within either forest buffer (16.9 ± 0.2%, n = 93), or grass filter (19.0 ± 0.2%, n = 93). During the same period, average daily soil temperature in the crop fields (22.8 ± 0.3°C, n = 93) was significantly higher than those in forest buffers and grass filters (21.8°C, n = 93) ($P < 0.01$).

**Soil Incubation Experiments**

Soil gravimetric moisture at the time of soil incubations was 14.8 ± 0.3% in the crop field soils (n = 18), 22.8 ± 0.8% within forest buffers soils (n = 18), 19.0 ± 0.7% within warm-season grass filters soils (n = 18), and 26.1 ± 0.3% within cool-season grass filters soils (n = 18) with the observed differences significant (one-way ANOVA $P < 0.01$). Aerobic CH$_4$ production in the incubated forest buffer soils (3.5 × 10$^{-2}$ ± 2.3 × 10$^{-2}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 3), warm-season grass filter soils (1.6 × 10$^{-3}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 1), and cool-season grass filter soils (5.4 × 10$^{-3}$ ± 1.9 × 10$^{-3}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 3) and crop field soils (1.8 × 10$^{-2}$ ± 6.9 × 10$^{-3}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 3) were not significantly different (Fig. 1). Similarly, net CH$_4$ fluxes in the incubated forest buffer soils (–4.9 × 10$^{-2}$ ± 2.3 × 10$^{-2}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 3), warm-season grass filter soils (–1.2 × 10$^{-2}$ ± 5.3 × 10$^{-3}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 3), and cool-season grass filter soils (–1.5 × 10$^{-2}$ ± 1.2 × 10$^{-2}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 3) and crop field soils (5.4 × 10$^{-3}$ ± 6.5 × 10$^{-3}$ ng CH$_4$–C kg soil$^{-1}$ h$^{-1}$, n = 3) were not significantly different (Fig. 1).

**Diel Variation of Methane Flux and Controlling Factors**

The diel variation of CH$_4$ flux for sampling conducted during 16–17 July 2007 did not differ among any of the vegetation types. Similarly, there was no significant difference in CH$_4$ flux among vegetation types (Fig. 2A). The average CH$_4$ flux (n = 8) was 6.9 ± 12.2 μg CH$_4$– C m$^{-2}$ h$^{-1}$ in crop field, -23.6 ± 8.1 μg CH$_4$– C m$^{-2}$ h$^{-1}$ within the forest buffer, –19.1 ± 10.6 μg CH$_4$– C m$^{-2}$ h$^{-1}$ within the warm-season grass filter and –25.5 ± 10.2 μg CH$_4$– C m$^{-2}$ h$^{-1}$ within the cool-season grass filter. The variation of CH$_4$ flux in the crop field, forest buffer, warm-season grass filter and cool-season grass filter was not correlated with soil temperature. Cumulative CH$_4$ flux over 24 h was 165.7 μg CH$_4$– C m$^{-2}$ d$^{-1}$ in the crop field, –567.3 μg CH$_4$– C m$^{-2}$ d$^{-1}$ in the forest buffer, –459.1 μg CH$_4$– C m$^{-2}$ d$^{-1}$ in the warm-season
grass filter, and -612.7 μg CH₄–C m⁻² d⁻¹ in the cool-season grass filter (Fig. 2B).

**Seasonal Variation of Methane Flux and Annual Methane Emission**

Since there was no significant variation in CH₄ flux through time within a day (Fig. 2A), daily fluxes were calculated by multiplying measured hourly fluxes (mid-morning) by 24 h. Observed maximum positive daily CH₄ flux was 2.4 mg CH₄–C m⁻² d⁻¹ in crop fields (7 August), 2.4 mg CH₄–C m⁻² d⁻¹ in forest buffers (1 May), and 1.6 mg CH₄–C m⁻² d⁻¹ and grass filters (11 September) (Fig. 3). Observed maximum negative daily CH₄ flux was -2.2 mg CH₄–C m⁻² d⁻¹ in crop fields (16 January), -2.6 mg CH₄–C m⁻² d⁻¹ in forest buffers (20 September), and -3.7 mg CH₄–C m⁻² d⁻¹ (20 September) grass filters (Fig. 3). Mean daily CH₄ flux was -0.2 ± 0.1 mg CH₄–C m⁻² d⁻¹ (n = 40) in the crop field, -0.5 to 0.9 mg CH₄–C m⁻² d⁻¹ (n = 45–50) in forest buffers, and -0.2 to 0.1 mg CH₄–C m⁻² d⁻¹ (n = 41–49) in grass filters, with no significant differences (Fig. 4). There were no significant relationships between CH₄ flux and soil moisture (P > 0.05) or soil temperature (P > 0.05) in any of the sites. Cumulative CH₄ flux from January to December 2007 was -0.80 kg CH₄–C ha⁻¹ yr⁻¹ (n = 1) in the crop field, -0.46 ± 0.48 kg CH₄–C ha⁻¹ yr⁻¹ (n = 3) in forest buffers, and 0.04 ± 0.2 kg CH₄–C ha⁻¹ yr⁻¹ (n = 4) in grass filters (Fig. 5). Cumulative CH₄ flux in forest buffers (95% confidence interval (CI): -2.54 to 1.61 kg CH₄–C ha⁻¹ yr⁻¹) and grass filters (95% CI: -0.51 to 0.61 kg CH₄–C ha⁻¹ yr⁻¹) were not significantly different from zero. The cumulative CH₄ flux in the crop field, forest buffers and grass filters were not significantly different from one another (one-way ANOVA P = 0.40) (Fig. 5).

**Discussion**

**Change of Soil Properties after Re-establishment of Riparian Buffers**

Soils within forest buffers and grass filters had significantly lower bulk density, higher pH, TC, TN, and NH₄⁺ than those in adjacent crop fields. This suggests that the re-establishment of the perennially vegetated buffers changed these properties in soils that were previously under row-crop cultivation. This conclusion is corroborated when comparing data collected from the same sites in 1998 and 1999 and indicate a 29% increase in soil C in the forest buffer over the last 9 yr. Decomposition of above and below-ground litter, root exudates, and microbial C accumulation may contribute to the observed C increase. Increased soil C resulting from conservation practices such as conversion from crop lands to grasslands or forest has been reported in other studies (Gebhart et al., 1994; Knops and Tilman, 2000; Uri, 2000; Post and Kwon, 2000; Guo and Giff ord, 2002; McLauchlan et al., 2006). Johnson et al. (2005) reported that conversion of previous cropland to grass increased soil organic C by 4.2 ± 4.5 Mg C ha⁻¹ yr⁻¹ after 6–8 yr since establishment in the central United States.

We observed significantly higher soil moisture and lower soil temperature in the soils of riparian buffers compared to those of the crop field. This may be the result of the vegetation within riparian buffers providing more shade to prevent high temperatures in the summer and the lower soil bulk density and high organic matter of riparian buffers holding more soil moisture. In contrast, soils in conventionally cultivated crop fields are more exposed to direct sunlight, have higher bulk density and lower soil organic matter and tend to hold less soil moisture compared with riparian buffers soils.

**Methane Flux in Riparian Buffers**

Methane flux observed within the forest buffers and grass filter soils (-0.5 to 0.9 mg CH₄–C m⁻² d⁻¹, n = 45–50) is similar to results of studies conducted in other riparian systems with infrequent saturation. McLain and Martens (2006) found the CH₄ sink averaged 26.1 ± 6.3 μg CH₄ m⁻² h⁻¹ in the semiarid riparian soils of southeastern Arizona. In a riparian alder stand in southeastern Estonia, Teiter and Mander (2005) observed an average CH₄ flux of 0.1 to 265 μg CH₄–C m⁻² h⁻¹. However, the CH₄ flux in the forest buffers and grass filters soils was lower than those reported in other studies conducted in temporarily submerged areas such as rice (Oryza sativa L.) fields, wetlands, or riparian areas with frequent saturation. For example, Ambus and Christensen...
(1995) found CH$_4$ produced at rates of 7877 mg CH$_4$–C m$^{-2}$ yr$^{-1}$ (78.8 kg CH$_4$–C ha$^{-1}$ yr$^{-1}$) in a temporarily flooded riparian area in Denmark. Methane was produced from riparian areas in northern Germany at rates of 33 to 33,030 mg CH$_4$–C m$^{-2}$ yr$^{-1}$ (0.33–330.3 kg CH$_4$–C ha$^{-1}$ yr$^{-1}$) (Merbach et al., 1996). Altor and Mitsch (2006) reported that annual CH$_4$ flux from intermittently flooded zones was 13 g CH$_4$–C m$^{-2}$ yr$^{-1}$ (130 kg CH$_4$–C ha$^{-1}$ yr$^{-1}$) in the midwestern United States. Le Mer and Roger’s (2001) review of the literature found that the median of CH$_4$ emissions were 0.72 kg CH$_4$ ha$^{-1}$ d$^{-1}$ (3 mg CH$_4$ m$^{-2}$ h$^{-1}$) in swamps, 0.43 kg CH$_4$ ha$^{-1}$ d$^{-1}$ (1.8 mg CH$_4$ m$^{-2}$ h$^{-1}$) in peat lands and 1.0 kg CH$_4$ ha$^{-1}$ d$^{-1}$ (4.2 mg CH$_4$ m$^{-2}$ h$^{-1}$) in rice fields. These results suggest that riparian zones soils under certain conditions are not major sources of CH$_4$ compared to wetlands, rice fields, or riparian zones with more frequent saturation. In the case of riparian zones in many areas of the midwestern United States, changes in landscape hydrology resulting from the conversion to agriculture have resulted in incised stream channels and lowered riparian water tables, likely altering conditions favorable to CH$_4$ production. At our sites, some riparian buffers were easily affected by flooding caused by snow melting (14 March) and heavy rainfall (26 April) and we conducted gas sampling when we were able to access the sites after flooding subsided. However, it is likely that because these conditions were so ephemeral, that observed CH$_4$ mass flux reflect the hydrologic characteristics of riparian buffers within this landform region.

Fig. 3. (A,B) Methane flux, (C) daily precipitation, (D) daily soil moisture, and (E) soil temperature in crop fields (n = 1), forest buffers (n = 3), and grass filters (n = 4) in 2007. Each mean represents observations and bars are the standard error of the mean. Gaps in the soil moisture data (D) were caused by either data loggers malfunction or removing a data logger during planting and harvesting periods in the crop field.
Because this study was conducted for only 1 yr (January 2007–December 2007), the interannual variation of CH₄ flux cannot be assessed for these sites. However, a main objective of this study was to compare CH₄ flux among crop fields and adjacent riparian buffers riparian buffers re-established for water quality. Since all sites were in close proximity and experienced similar conditions, it can be assumed that annual (or interannual) climate variability did not affect study conclusions. Since climatic conditions in 2007 (mean air temperature 9.4°C; annual precipitation 1097 mm) were within the standard deviation of conditions over the last 37 yr of record (mean air temperature 8.7 ± 0.8°C; mean annual precipitation 914 ± 210 mm) results from this study could be considered representative of flux rates measured over multiple years.

Results of soil incubation experiments, and did and seasonal CH₄ flux measurements indicate that CH₄ flux in the crop field, forest buffers, and grass filters were not significantly different from one another. In contrast, in the same region (central Iowa), Chan and Parkin (2001a) found that forest and prairie soils were net CH₄ consumers, with cumulative CH₄ fluxes ranging from –0.27 to –0.07 g CH₄ m⁻² (–2.7 to –0.7 kg CH₄ ha⁻¹) over the 258-d sampling season, while agricultural sites were net CH₄ producers, with cumulative CH₄ fluxes ranging from –0.02 to 3.19 g CH₄ m⁻² (–0.2 to 31.9 kg CH₄ ha⁻¹) over the same season. The prairie and forest soils were found to have the greatest potential to oxidize atmospheric concentrations of CH₄ (Chan and Parkin, 2001b). Within temperate regions globally, reported CH₄ consumption rates include 1.29 ± 0.16 kg CH₄ ha⁻¹ yr⁻¹ in crop fields (n = 48), 5.75 ± 0.59 kg CH₄ ha⁻¹ yr⁻¹ within grasslands (n = 24), and 2.40 ± 0.40 kg CH₄ ha⁻¹ yr⁻¹ within forests (n = 91) [data extracted from Dutaur and Verchot (2007)]. These reports indicate that CH₄ consumption within re-established riparian forest buffer and grass filter soils examined in this study were much lower than other reported CH₄ consumption rates within grasslands and forests in Iowa and the temperate regions. Such a contrast suggests that CH₄ soil oxidation capacity has not been improved during the 7 to 17 yr following re-establishment of perennial vegetation (forest buffers and grass filters) on conventional crop fields, even when soil properties such as soil bulk density pH, TC, and soil moisture have changed significantly. It is well known that CH₄ oxidation potential of upland soils is reduced by cultivation and ammonium N-fertilizer application (e.g., Topp and Pattey, 1997; Le Mer and Roger, 2001; Dutaur and Verchot, 2007). Le Mer and Roger (2001) summarized the effects of cultural practices on CH₄ oxidation as following: (i) an increase in NH₄⁺ content of soil by fertilizer application inhibits CH₄ oxidation because NH₄⁺ produces competition at the level of methane-mono-oxygenase, a transfer of the CH₄ oxidizing activity toward nitrification (Castro et al., 1994; Nesbit and Breitenbeck,1992), and (ii) cultural practices that destroy micro-aerophilic niches suitable for CH₄ oxidizers reduce CH₄ oxidation (Hünsch et al., 1994; Sitaula et al., 2000). Slow recovery of CH₄ oxidation after land use change has been reported. In a range of successional sites on former arable land in Denmark and Scotland, CH₄ oxidation rates took more than 100 yr to reach precultivation levels (Priemé et al., 1997). Similarly, Suwanwaree and Robertson (2005) observed that rates of CH₄ oxidation in soils of 40 to 60 yr-old successional fields were between those of the no-till and deciduous forest sites in southwest United States. Singh et al. (2007) reported that afforestation and reforestation of pastures (30–50 yr later) resulted in changes in methane oxidation by altering the community structure of methanotrophic bacteria in these soils. In the case of the re-established riparian buffers investigated in this study, it appears that, while soil properties have been altered, additional time is needed for changes in CH₄ flux to be manifested.

Conclusions

Soil properties such as soil bulk density, pH, TC, and soil moisture in riparian forest buffer and grass filter soils were significantly different from those in adjacent crop fields, suggesting that soil properties have changed since re-establishment of perennial vegetation on previously cultivated field soils. Soil incubation experiments provide some indication that CH₄ consumption was higher than CH₄ production in forest buffers...
and grass filters soils, while crop field soils showed the opposite response. However, none of the CH$_4$ fluxes from incubation experiments were significantly different, nor were diel and seasonal variation of CH$_4$ fluxes in forest buffers, grass filters, and adjacent crop fields. The cumulative CH$_4$ flux $-0.80$ kg CH$_4$-C ha$^{-1}$ yr$^{-1}$ in the crop field, $-0.46$ kg CH$_4$-C ha$^{-1}$ yr$^{-1}$ in forest buffers, and 0.04 kg CH$_4$-C ha$^{-1}$ yr$^{-1}$ in grass were also not significantly different. The CH$_4$ flux in forest buffers and grass filter soils was less than that reported for wetlands, rice fields, or riparian areas with more frequent saturation, which are known to be sinks of CH$_4$. These results suggest that these re-established riparian forest buffers and grass filters, possibly due to altered hydrology, cannot be considered as major sources of CH$_4$ as has been found in other riparian areas or systems with more frequent saturation. However, any potential benefit as increased sinks of CH$_4$ has not yet been achieved after 7 to 17 yr since re-establishment. These results have important management implications given the significant effort to promote such systems for water quality improvement and other ecosystem services.

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References


Priemé, A., S. Christensen, K.E. Dobbie, and K.A. Smith. 1997. Slow increase in rate of methane oxidation in soils with time following land use change
Sabater, S., A. Butturini, J.C. Clement, T. Burt, D. Dowrick, M. Hefting,
removal by riparian buffers along a European climatic gradient: Patterns
in a grassed and a wooded, valley and ridge, riparian ecotone. J. Environ.
Qual. 25:1230–1235.
Schultz, R.C., J.P. Colletti, T.M. Isenhart, C.O. Marquez, W.W. Simpkins,
Garrett et al. (ed.) North American agroforestry: An integrated science
and practice. ASA, Madison, WI.
Schultz, R.C., J.P. Colletti, T.M. Isenhart, W.W. Simpkins, C.W. Mize, and
forest buffers in agroecosystems—lessons learned from the Bear Creek
Watershed, central Iowa, USA. Agrofor. Syst. 61:35–50.
Singh, B.K., K.R. Tate, G. Kolipaka, C.B. Hedley, C.A. Macdonald, P.
Millard, and J.C. Murrell. 2007. Effect of afforestation and reforestation
of pastures on the activity and population dynamics of methanotrophic
oxidation potentials and fluxes in agricultural soil: Effects of fertilisation
Suwanwaree, P., and G. Robertson. 2005. Methane oxidation in forest,
successional, and no-till agricultural ecosystems: Effects of nitrogen and
Tate, K.R., D.J. Ross, S. Saggar, C.B. Hedley, J. Dando, B.K. Singh, and S.M.
Lambie. 2007. Methane uptake in soils from Pinus radiata plantations, a
reverting shrubland and adjacent pastures: Effects of land-use change, and
constructed wetlands for wastewater treatment and from riparian buffer
Tonderski, A. 1996. Landuse-based nonpoint source pollution. A threat to
Topp, E., and E. Pattey. 1997. Soils as sources and sinks for atmospheric
biomass, soil respiration, and root distribution in crop fields and riparian
United States Department of Commerce–National Oceanic and Atmospheric
Van Miegroet, H. 1995. Inorganic nitrogen determined by laboratory and field