

12-2012

Effect of immune system stimulation and divergent selection for residual feed intake on digestive capacity of the small intestine in growing pigs

A. Rakhshandeh
Iowa State University

Jack C. M. Dekkers
Iowa State University, jdekke@iastate.edu

Brian J. Kerr
United States Department of Agriculture

Thomas E. Weber
Elanco Animal Health, tom.weberte@ars.usda.gov

J. English
Iowa State University
Follow this and additional works at: http://lib.dr.iastate.edu/ans_pubs

 *next page for additional authors*
Part of the [Agriculture Commons](#), and the [Meat Science Commons](#)

The complete bibliographic information for this item can be found at http://lib.dr.iastate.edu/ans_pubs/33. For information on how to cite this item, please visit <http://lib.dr.iastate.edu/howtocite.html>.

This Article is brought to you for free and open access by the Animal Science at Iowa State University Digital Repository. It has been accepted for inclusion in Animal Science Publications by an authorized administrator of Iowa State University Digital Repository. For more information, please contact digirep@iastate.edu.

Effect of immune system stimulation and divergent selection for residual feed intake on digestive capacity of the small intestine in growing pigs

Abstract

Little is known of the consequences of divergent selection for residual feed intake (RFI) on intestinal digestion capacity, particularly during immune system stimulation (ISS). Our objective was to evaluate the impact of ISS and divergent selection for RFI on apparent ileal digestibility (AID) and apparent fecal digestibility (AFD) of nutrients and intestinal nutrient active transport and barrier function. Twenty-eight gilts (63 ± 4 kg BW) from low RFI (LRFI; $n = 14$) and high RFI (HRFI; $n = 14$) Yorkshire lines were randomly selected from the Iowa State University RFI herd. Following adaptation, 8 pigs in each line were injected intramuscularly and every 48 h for 7 d with increasing amounts of *Escherichia coli* lipopolysaccharide (ISS+). Remaining pigs were injected with saline (ISS-). Pigs were then euthanized and ileal digesta was collected for measuring AID of nutrients. Fecal samples were collected on a daily basis and pooled for measuring AFD of nutrients. A segment of ileum was used to measure nutrient transport and transepithelial resistance (TER) and/or barrier integrity by Ussing chambers. No effects of line or its interaction with ISS on AID of CP ($N \times 6.25$) and OM, TER, and active nutrients transport were observed. However, ISS decreased ($P < 0.05$) and tended to ($P < 0.1$) decrease AID of CP and OM, respectively. Decrease in AFD of CP as result of ISS was greater in the LRFI line compared to the HRFI line ($P < 0.05$). Relative to ISS-, active glucose and P transport was greater in ISS+ pigs ($P < 0.05$). Genetic selection for LRFI increases the AFD but has no effect on AID of nutrients. It also reduces the total tract digestive capacity of growing pigs during ISS. Immune system stimulation affects both AID and AFD of dietary CP.

Keywords

Swine Feed Efficiency, ileal digestibility, immune system stimulation, nutrient transport, residual feed intake

Disciplines

Agriculture | Animal Sciences | Meat Science

Comments

This article is from *Journal of Animal Science* 90 (2012): 233–235, doi:[10.2527/jas.53976](https://doi.org/10.2527/jas.53976).

Rights

Works produced by employees of the U.S. Government as part of their official duties are not copyrighted within the U.S. The content of this document is not copyrighted.

Authors

A. Rakhshandeh, Jack C. M. Dekkers, Brian J. Kerr, Thomas E. Weber, J. English, and Nicholas K. Gabler

JOURNAL OF ANIMAL SCIENCE

The Premier Journal and Leading Source of New Knowledge and Perspective in Animal Science

Effect of immune system stimulation and divergent selection for residual feed intake on digestive capacity of the small intestine in growing pigs

A. Rakhshandeh, J. C. M. Dekkers, B. J. Kerr, T. E. Weber, J. English and N. K. Gabler

J ANIM SCI 2012, 90:233-235.

doi: 10.2527/jas.53976

The online version of this article, along with updated information and services, is located on the World Wide Web at:

http://www.journalofanimalscience.org/content/90/Supplement_4/233



American Society of Animal Science

www.asas.org

Effect of immune system stimulation and divergent selection for residual feed intake on digestive capacity of the small intestine in growing pigs¹

A. Rakhshandeh,* J. C. M. Dekkers,* B. J. Kerr,† T. E. Weber,‡ J. English,* and N. K. Gabler*²

*Department of Animal Science, Iowa State University, Ames, IA50011; †USDA-ARS, Ames, IA 5011; and

‡Elanco Animal Health, Greenfield, IN 46140

ABSTRACT: Little is known of the consequences of divergent selection for residual feed intake (RFI) on intestinal digestion capacity, particularly during immune system stimulation (ISS). Our objective was to evaluate the impact of ISS and divergent selection for RFI on apparent ileal digestibility (AID) and apparent fecal digestibility (AFD) of nutrients and intestinal nutrient active transport and barrier function. Twenty-eight gilts (63 ± 4 kg BW) from low RFI (LRFI; $n = 14$) and high RFI (HRFI; $n = 14$) Yorkshire lines were randomly selected from the Iowa State University RFI herd. Following adaptation, 8 pigs in each line were injected intramuscularly and every 48 h for 7 d with increasing amounts of *Escherichia coli* lipopolysaccharide (ISS+). Remaining pigs were injected with saline (ISS-). Pigs were then euthanized and ileal digesta was collected for measuring AID of nutrients. Fecal samples were

collected on a daily basis and pooled for measuring AFD of nutrients. A segment of ileum was used to measure nutrient transport and transepithelial resistance (TER) and/or barrier integrity by Ussing chambers. No effects of line or its interaction with ISS on AID of CP ($N \times 6.25$) and OM, TER, and active nutrients transport were observed. However, ISS decreased ($P < 0.05$) and tended to ($P < 0.1$) decrease AID of CP and OM, respectively. Decrease in AFD of CP as result of ISS was greater in the LRFI line compared to the HRFI line ($P < 0.05$). Relative to ISS-, active glucose and P transport was greater in ISS+ pigs ($P < 0.05$). Genetic selection for LRFI increases the AFD but has no effect on AID of nutrients. It also reduces the total tract digestive capacity of growing pigs during ISS. Immune system stimulation affects both AID and AFD of dietary CP.

Key words: ileal digestibility, immune system stimulation, nutrient transport, residual feed intake.

© 2012 American Society of Animal Science. All rights reserved.

J. Anim. Sci. 2012.90:233–235

doi:10.2527/jas53976

INTRODUCTION

The gastrointestinal tract is involved in defense activities during systemic immune system stimulation (ISS). This involvement results in physiological changes in intestinal tissues including edema, change in gut motility, permeability, microflora (Barbara et al., 2005; Dharmani et al., 2009), expression of digestive enzymes (Jurjus et al., 2006), increased mucin production (Faure et al., 2007), and disordering of epithelial transport systems (Hang et al., 2003). However, little is known about the impact of ISS on measures of nutrient digestibility as it relates to feed efficiency in pigs. Residual feed intake (RFI) is a measure of feed

efficiency that reflects differences in the efficiency of the use of feed for maintenance and growth, and genetic selection for low RFI (LRFI) substantially reduced the amount of feed required for growth and backfat without affecting meat quality (Cai et al., 2008). One of the major physiological processes that contributes to variation in RFI is the digestion of feed (Herd and Arthur, 2009). However, the consequences of genetic selection for RFI on intestinal nutrient digestion capacity, particularly during ISS, are poorly documented for growing pigs. The objective of this study was to evaluate the impact of ISS in pigs divergently selected for RFI on apparent ileal digestibility (AID) and apparent fecal digestibility (AFD) of nutrients, intestinal nutrient transport, and barrier function.

¹This research was supported by the Agriculture and Food Research Initiative competitive grant number 2011-68004-30336 from the USDA National Institute of Food and Agriculture.

²Corresponding author: ngabler@iastate.edu

MATERIAL AND METHODS

Twenty-eight Yorkshire gilts (63 ± 4 kg initial BW) from pigs selected for LRFI ($n = 14$) and high RFI (HRFI; $n = 14$) were randomly selected from the Iowa State University RFI herd and used in this study. Pigs were fed a corn (*Zea mays*)–soybean (*Glycine max*)-based diet containing 15.9 MJ/kg DE and 5.2 g/kg standardized ileal digestible Lys twice daily and feed restricted (1.5 kg/d) (Rakhshandeh and de Lange, 2012). Titanium dioxide was used as an indigestible marker. Immune system stimulation was induced according to Rakhshandeh and de Lange (2012). Following adaptation, 8 pigs in each line were injected intramuscularly and every 48 h for 7 d with increasing amounts of *Escherichia coli* O5:B55 lipopolysaccharide (ISS+; initial dose of 30 μ g/kg BW). Remaining pigs were injected with sterile saline (ISS–). Eye temperatures were measured on a daily basis using infrared imaging technique (Rakhshandeh and de Lange, 2012). Blood samples were taken at 0, 24, 72, 120, and 168 h after ISS. At the end of the 7-d period, pigs were euthanized and digesta was collected from the distal 1.5 m of the small intestine for measuring AID of dietary nutrients. The electrophysiological parameters, including transepithelial resistance (TER) and short circuit current changes of ileal segment, were determined using Ussing chambers according to Gabler et al. (2007). Feces were collected at least twice daily and pooled for a 7-d collection period for measuring AFD of dietary nutrients. Data were analyzed using a factorial randomized complete block design [PROC MIXED of the SAS system (SAS Institute Inc., Cary, NC)]. Data collected over time were analyzed as repeated measurements.

RESULTS AND DISCUSSION

Repeated injection with lipopolysaccharide (ISS– vs. ISS+) increased ($P < 0.03$) eye temperature (37.7 vs. 38.4°C; SE = 0.03), plasma levels of IL-1 β (0 vs. 152 ng/L; SE = 35) and haptoglobin (2.2 vs. 3.5 g/L; SE = 0.27), and decreased white blood cell count (21.7 vs. 19.6 $\times 10^3/\mu$ L; SE = 0.89) confirming successful ISS (Gabler et al., 2008; Rakhshandeh and de Lange, 2012). No line or line \times ISS interaction effects on AID of nutrients were observed ($P > 0.90$). Immune system stimulation decreased ($P < 0.05$) AID and AFD of dietary CP and tended to decrease ($P < 0.07$) AID of OM (Table 1). It has been suggested that systemic ISS increases gut endogenous nutrient losses and reduces intestinal absorptive capacity by compromising intestinal integrity and interfering with nutrient transport system (Hang et al., 2003; Faure et al., 2007). However, in the current study, systemic ISS did not affect intestinal integrity (TER) but increased ileal glucose and P active transport as measured by modified Ussing chambers ($P < 0.05$; Table 1). Therefore, in the present study, the decrease in AID of nutrients could largely be attributed to increased intestinal endogenous nutrient, especially protein losses. Increase in glucose use and reabsorption of anions have also been reported by other investigators during ISS (Klasing and Leshchinsky, 2000; Yajima et al., 2009). Immune system stimulation decreased ($P < 0.05$) and tended to decrease ($P < 0.06$) AFD of CP and OM, respectively, in the LRFI line. This probably is associated with compromised microbial activity and increased large intestinal mucin secretion (Barbara et al., 2005; Dharmani et al., 2009). We have observed more exaggerated immune response to a controlled ISS in the

Table 1. Effect of immune system stimulation (ISS) and divergent selection for residual feed intake (RFI) on apparent ileal digestibility (AID) and apparent fecal digestibility (AFD) of dietary nutrients and electrophysiological parameters of ileum¹

	HRFI ²			LRFI ²			<i>P</i>		
	ISS–	ISS+	SE	ISS–	ISS+	SE	Line	ISS	Line \times ISS
AID, %									
CP (N \times 6.25)	82	76	5.6	81	72	5.8	0.39	0.05	0.97
OM	89	80	6.3	87	79	5.2	0.70	0.07	0.90
AFD, %									
CP (N \times 6.25)	80	78	1.1	82	79	0.6	0.01	0.04	0.02
OM	90	90	0.3	91	90	0.2	0.01	0.21	0.06
Nutrient transport, μ A ³ /cm ²									
Glucose	7.0	9.6	2.01	6.3	11.6	2.32	0.74	0.05	0.48
Glutamine	1.0	1.6	0.94	0.5	0.6	0.2	0.17	0.56	0.70
Phosphorous	7.0	8.4	2.01	5.3	12.7	2.93	0.53	0.04	0.15
TER, Ω^4 cm ²	110	126	15	135	139	29	0.34	0.60	0.75

¹Data are least squares means and represent data obtained on day 7 after the start of ISS. The AID and AFD was determined using indicator method and slaughter technique.

²HRFI = high RFI; LRFI = low RFI; ISS– = repeatedly injected with sterile saline; ISS+ = repeatedly injected with increasing amounts of *Escherichia coli* lipopolysaccharide.

³A = ampere.

⁴TER = transepithelial resistance; Ω = ohm.

LRFI line compared to the HRFI line (unpublished data). Apparent fecal digestibility of CP and OM were lower ($P < 0.01$) in the HRFI line compared to the LRFI line (Table 1). Higher AFD of nutrients in animals selected for LRFI largely has been associated with level of feed intake (Herd and Arthur, 2009). However, in the current study feed intake was controlled and identical for both lines. Therefore, the higher AFD of nutrients in the LRFI line can most likely be attributed to lower endogenous (i.e., mucins) losses or higher microbial activity and lower colonic motility or both (McDonald et al., 2010). These results are in general agreement with findings in lines of sheep, cattle, and chicken selected for LRFI (Richardson et al., 2004; Herd and Arthur, 2009). Genetic selection for LRFI increases AFD, but it has no effect on AID of nutrients. The latter suggests that differences in AFD might not be important sources of variation in RFI. Immune system stimulation affects both AID and AFD of dietary nutrients in pigs and may be a major source of feed efficiency variation. Genetic selection for LRFI reduces the total tract digestive capacity of growing pigs during ISS.

LITERATURE CITED

- Barbara, G., V. Stanghellini, G. Brandi, C. Cremon, G. Di Nardo, R. De Giorgio, and R. Corinaldesi. 2005. Interactions between commensal bacteria and gut sensorimotor function in health and disease. *Am. J. Gastroenterol.* 100:2560–2568.
- Cai, W., D. S. Casey, and J. C. Dekkers. 2008. Selection response and genetic parameters for residual feed intake in Yorkshire swine. *J. Anim. Sci.* 86:287–298.
- Dharmani, P., V. Srivastava, V. Kisson-Singh, and K. Chadee. 2009. Role of intestinal mucins in innate host defense mechanisms against pathogens. *J. Innate Immun.* 1:123–135.
- Faure, M., F. Choné, C. Mettraux, J. P. Godin, F. Béchereau, J. Vuichoud, I. Papet, D. Breuillé, and C. Obléd. 2007. Threonine utilization for synthesis of acute phase proteins, intestinal proteins, and mucins is increased during sepsis in rats. *J. Nutr.* 137:1802–1807.
- Gabler, N. K., J. D. Spencer, D. M. Webel, and M. E. Spurlock. 2007. In utero and postnatal exposure to long chain (n-3) PUFA enhances intestinal glucose absorption and energy stores in weanling pigs. *J. Nutr.* 137:2351–2358.
- Gabler, N. K., J. D. Spencer, D. M. Webel, and M. E. Spurlock. 2008. n-3 PUFA attenuate lipopolysaccharide-induced down-regulation of toll-like receptor 4 expression in porcine adipose tissue but does not alter the expression of other immune modulators. *J. Nutr. Biochem.* 19:8–15.
- Hang, C. H., J. X. Shi, J. S. Li, W. Wu, and H. X. Yin. 2003. Alterations of intestinal mucosa structure and barrier function following traumatic brain injury in rats. *World J. Gastroenterol.* 9:2776–2781.
- Herd, R. M., and P. F. Arthur. 2009. Physiological basis for residual feed intake. *J. Anim. Sci.* 87:E64–E71.
- Jurjus, A., K. Barada, N. Khoury, M. D. Assef, C. J. Foltzer, J. M. Reimund, and M. Keding. 2006. Morphological and biochemical alterations in the jejunum following iodoacetamide-induced colitis in rats. *Can. J. Physiol. Pharmacol.* 84:1191–1203.
- Klasing, K. C., and V. Leshchinsky. 2000. Interaction between nutrition and immunity. Pages 363–373 in *Nutrition and Immunology*. M.E. Gershwin, J. B. German, and C. L. Keen, eds. 1st ed. Humana Press, Burbank, CA.
- McDonald, P., R. A. Edwards, J. F. D. Greenhalgh, C. A. Morgan, L. A. Sinclair, and R. G. Wilkinson. 2010. Digestion. Pages 130–137 in *Animal Nutrition*. 7th ed. Prentice Hall.
- Rakhshandeh, A., and C. F. de Lange. 2012. Evaluation of chronic immune system stimulation models in growing pigs. *Animal* 6:305–310.
- Richardson, E. C., R. M. Herd, J. A. Archer, and P. F. Arthur. 2004. Metabolic differences in angus steers divergently selected for residual feed intake. *Aust. J. Exp. Agric.* 44:441–452.
- Yajima, S., H. Morisaki, R. Serita, T. Suzuki, N. Katori, T. Asahara, K. Nomoto, F. Kobayashi, A. Ishizaka, and J. Takeda. 2009. Tumor necrosis factor-alpha mediates hyperglycemia-augmented gut barrier dysfunction in endotoxemia. *Crit. Care Med.* 37(3):1024–1030.

References

This article cites 12 articles, 4 of which you can access for free at:
http://www.journalofanimalscience.org/content/90/Supplement_4/233#BIBL

Citations

This article has been cited by 1 HighWire-hosted articles:
http://www.journalofanimalscience.org/content/90/Supplement_4/233#otherarticles