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Abstract

Background

The object of this study was to describe and contrast the kinetics of the humoral response in serum and oral fluid specimens during acute porcine reproductive and respiratory syndrome virus (PRRSV) infection. The study involved three trials of 24 boars each. Boars were intramuscularly inoculated with a commercial modified live virus (MLV) vaccine (Trial 1), a Type 1 PRRSV field isolated (Trial 2), or a Type 2 PRRSV field isolate (Trial 3). Oral fluid samples were collected from individual boars on day post inoculation (DPI) -7 and 0 to 21. Serum samples were collected from all boars on DPI -7, 0, 7, 14, 21 and from 4 randomly selected boars on DPI 3, 5, 10, and 17. Thereafter, serum and oral fluid were assayed for PRRSV antibody using antibody isotype-specific ELISAs (IgM, IgA, IgG) adapted to serum or oral fluid.

Results

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Conclusions

This study enhanced our knowledge of the PRRSV humoral immune response and provided a broader foundation for the development and application of oral fluid antibody-based diagnostics.

Keywords

PRRSV, Oral fluid, ELISA, IgM, IgA, IgG, Antibody kinetics

Disciplines

Large or Food Animal and Equine Medicine | Other Veterinary Medicine | Statistical Methodology | Veterinary Physiology

Comments

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Apisit Kittawornrat, Mark Engle, Yaowalak Panyasing, Chris Olsen, Kent J. Schwartz, Anna Rice, Sergio Lizano, Chong Wang, and Jeffrey J. Zimmerman

RESEARCH ARTICLE

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Kinetics of the porcine reproductive and respiratory syndrome virus (PRRSV) humoral immune response in swine serum and oral fluids collected from individual boars

Apsit Kittawornrat¹, Mark Engle³, Yaowalak Panyasing¹, Chris Olsen¹, Kent Schwartz¹, Anna Rice⁴, Sergio Lizano⁴, Chong Wang^{1,2} and Jeffrey Zimmerman^{1*}

Abstract

Background: The object of this study was to describe and contrast the kinetics of the humoral response in serum and oral fluid specimens during acute porcine reproductive and respiratory syndrome virus (PRRSV) infection. The study involved three trials of 24 boars each. Boars were intramuscularly inoculated with a commercial modified live virus (MLV) vaccine (Trial 1), a Type 1 PRRSV field isolated (Trial 2), or a Type 2 PRRSV field isolate (Trial 3). Oral fluid samples were collected from individual boars on day post inoculation (DPI) -7 and 0 to 21. Serum samples were collected from all boars on DPI -7, 0, 7, 14, 21 and from 4 randomly selected boars on DPI 3, 5, 10, and 17. Thereafter, serum and oral fluid were assayed for PRRSV antibody using antibody isotype-specific ELISAs (IgM, IgA, IgG) adapted to serum or oral fluid.

Results: Statistically significant differences in viral replication and antibody responses were observed among the three trials in both serum and oral fluid specimens. PRRSV serum IgM, IgA, and IgG were first detected in samples collected on DPI 7, 10, and 10, respectively. Oral fluid IgM, IgA, and IgG were detected in samples collected between DPI 3 to 10, 7 to 10, and 8 to 14, respectively.

Conclusions: This study enhanced our knowledge of the PRRSV humoral immune response and provided a broader foundation for the development and application of oral fluid antibody-based diagnostics.

Keywords: PRRSV, Oral fluid, ELISA, IgM, IgA, IgG, Antibody kinetics

Background

The presence of systemic and locally-produced antibodies in oral fluid has led to its use as a diagnostic specimen for a variety of infectious diseases. In humans, oral fluids have been used in the diagnosis of human immunodeficiency virus (HIV), Hepatitis A, B, and C viruses, measles, mumps and other infectious diseases [1]. In swine, antibodies against a variety of economically significant pathogens have been reported in oral fluids, including classical swine fever virus [2,3], porcine circovirus type 2 [4], porcine reproductive and respiratory syndrome virus [5,6],

swine influenza virus [7], transmissible gastroenteritis virus [8], *Actinobacillus pleuropneumoniae* [9], and *E. coli* [10]. In large measure, the kinetics of the antibody response against individual agents has not been described. Therefore, the purpose of present study was to describe and contrast the ontogeny of PRRSV IgM, IgA, and IgG in oral fluids and serum specimens collected from individually housed boars during acute PRRSV infection.

Results

PRRSV antibody isotypes in serum

The PRRSV antibody isotype (IgM, IgA, and IgG) responses in serum samples are shown in Figure 1 and Table 1. Estimates for DPI -7, 0, 7, 14, 21 were based on data from 72 boars, whereas estimates for DPIs 3, 5, 10,

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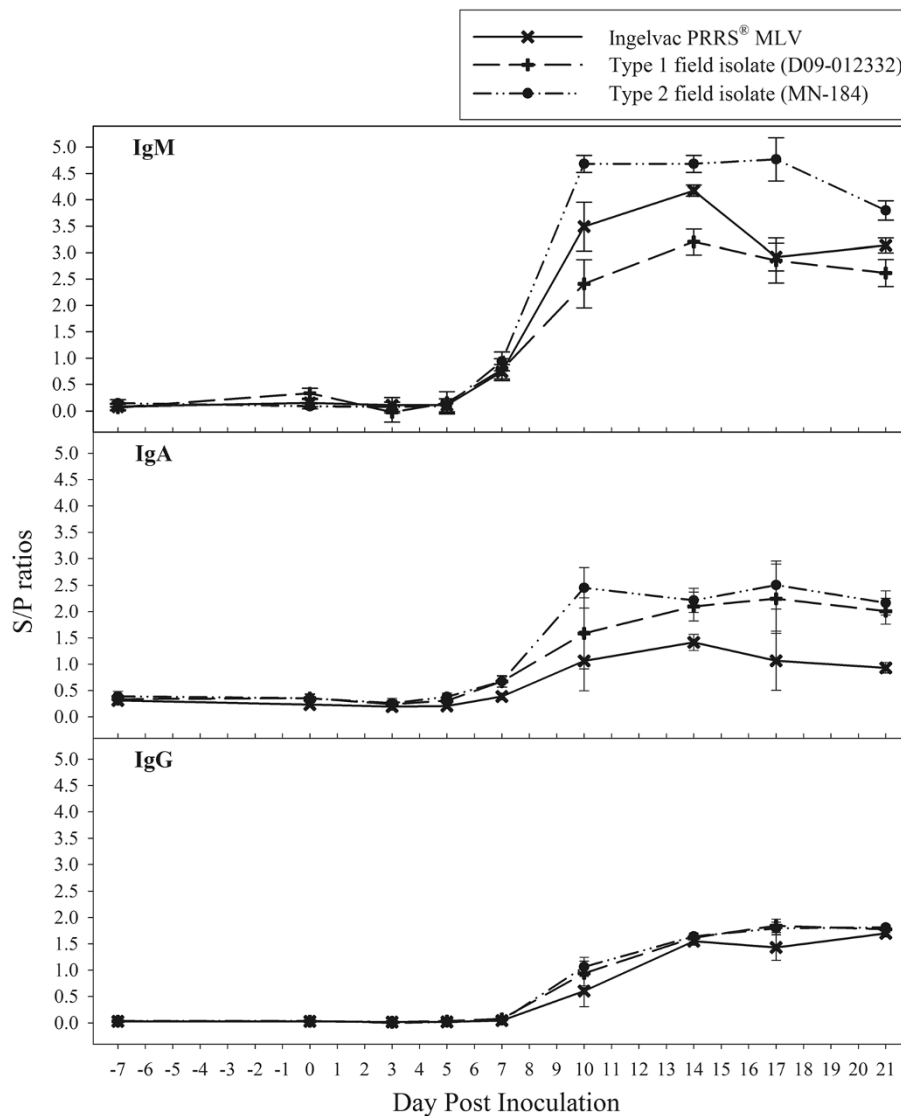


Figure 1 Kinetics of PRRSV antibody isotypes (IgM, IgA, and IgG) in serum based on responses in 72 boars inoculated with 3 different PRRSV isolates. Results are reported as mean sample-to-positive (S/P) ratios and standard errors.

and 17 were based on a subset of 12 animals randomly selected from the 72 boars. Based on pairwise comparisons, statistically significant levels of IgM were detected by DPI 7, peaked at DPI 14, and remained stable through DPI 21. In contrast, statistically significant levels of IgA and IgG were detected by DPI 10, after which they remained stable (IgA) or increased significantly (IgG) through DPI 21. Further analysis showed that IgM and IgA responses were associated with DPI (IgM: $p < 0.0001$, IgA: $p < 0.0001$), trial (IgM: $p < 0.0001$, IgA: $p < 0.0149$), and the interaction between trial and DPI (IgM: $p < 0.0001$, IgA: $p < 0.0001$). In contrast, DPI ($p < 0.0001$) was the only factor associated with the IgG response. Neither the age of the boar at the time of inoculation nor the quantity of oral fluid collected from each boar had a significant effect on IgM, IgA, or IgG.

PRRSV antibody isotype in oral fluid samples

The PRRSV antibody isotype (IgM, IgA, and IgG) responses in oral fluid samples are shown in Figure 2 and Table 1 for oral fluid samples collected on DPI -7, 0-14, 17, and 21. Estimates were based on ≥ 66 oral fluid samples at each sampling point. IgM S/P ratios were statistically significant on DPI 7, peaked at DPI 11, and declined thereafter. Levels of IgA and IgG were significant on DPI 8 and increased thereafter through the end of the experiment (DPI 21). Factors significantly associated with IgM and IgA S/P ratios included trial (IgM: $p < 0.0001$, IgA: $p = 0.0273$), DPI (IgM: $p < 0.0001$, IgA: $p < 0.0001$), oral fluid volume (IgM: $p = 0.0002$, IgA: $p < 0.0001$), and the interaction between trial and DPI (IgM: $p < 0.0001$, IgA: $p < 0.0001$). Factors associated with IgG included DPI ($p < 0.0001$) and

Table 1 Porcine reproductive and respiratory syndrome virus (PRRSV) antibody isotypes (IgM, IgA, and IgG) in oral fluid and serum samples collected from boars over day post inoculation (DPI)

DPI	Serum (mean S/P ratios)			Oral fluid (mean S/P ratios)				
	Samples	IgM	IgA	IgG	Samples	IgM	IgA	IgG
-7	72	0.10 ^d	0.35 ^b	0.03 ^d	70	0.02 ^g	0.15 ^g	0.01 ⁱ
0	72	0.19 ^d	0.31 ^b	0.03 ^d	68	0.02 ^g	0.08 ^g	0.01 ⁱ
1	-	-	-	-	69	0.04 ^g	0.12 ^g	0.01 ⁱ
2	-	-	-	-	68	0.04 ^g	0.13 ^g	0.02 ⁱ
3	12	0.05 ^d	0.23 ^b	0.01 ^d	70	0.07 ^g	0.16 ^g	0.02 ⁱ
4	-	-	-	-	68	0.13 ^g	0.24 ^g	0.01 ⁱ
5	12	0.12 ^d	0.30 ^b	0.02 ^d	67	0.05 ^g	0.11 ^g	0.01 ⁱ
6	-	-	-	-	66	0.09 ^g	0.12 ^g	0.01 ⁱ
7	72	0.82 ^c	0.58 ^b	0.05 ^d	69	0.30 ^f	0.23 ^g	0.03 ⁱ
8	-	-	-	-	66	1.28 ^e	0.41 ^f	0.17 ^h
9	-	-	-	-	66	2.32 ^c	0.95 ^e	0.59 ^g
10	12	3.56 ^b	1.70 ^a	0.87 ^c	70	2.72 ^b	1.45 ^{b, c}	0.99 ^f
11	-	-	-	-	70	2.95 ^a	1.75 ^a	1.41 ^e
12	-	-	-	-	70	2.70 ^b	1.59 ^{a, b}	1.78 ^d
13	-	-	-	-	69	2.64 ^b	1.64 ^a	2.05 ^c
14	72	4.02 ^a	1.91 ^a	1.60 ^b	68	2.32 ^c	1.56 ^{a, b}	2.28 ^b
17	12	3.51 ^b	1.94 ^a	1.69 ^{a, b}	69	1.53 ^d	1.25 ^{c, d}	2.63 ^a
21	72	3.18 ^b	1.70 ^a	1.76 ^a	68	1.02 ^e	1.14 ^{d, e}	2.60 ^a

^{a-i} Superscripts within columns indicate statistically significant differences among means (Tukey's Honestly Significant Differences test, $p < 0.05$).

the interaction of trial by DPI ($p < 0.0001$). Regardless of isotype, boar age at the time of inoculation had no significant effect on antibody response.

Comparison of PRRSV antibody responses in serum and oral fluid

A comparison of the qualitative antibody response in serum vs. oral fluid found no significant difference in the proportion of ELISA positive results on DPI 0 – 14, 17, and 21 (Table 2). No significant difference was detected in the number of positive results for each pair-wise sample combination (serum vs. oral fluid) by trial, DPI, or trial by DPI. On DPI 21, 100% of serum and oral fluid samples were positive with mean S/P ratios of 1.69 (95% Confidence Interval [CI]: 1.58, 1.79) and 2.60 (95% CI: 2.34, 2.86), respectively. An analysis of the quantitative antibody isotype responses in serum and oral fluid samples using test results from samples collected on DPI -7, 0 – 14, 17, 21 estimated Pearson's correlation coefficient as $r = 0.84$, 0.78 , and 0.90 for IgM, IgA, and IgG responses, respectively (Figure 3).

Comparison between viremia levels and antibody responses

All serum samples from DPI -7 and 0 ($n = 144$) were PRRSV qRT-PCR negative, whereas all serum samples from boars tested on DPI 3 ($n = 12$) and DPI 7 ($n = 72$)

were positive. PRRSV was detected in oral fluids from 7 of 69 boars at DPI 1, 52 of 68 boars at DPI 2, 66 of 70 at DPI 3, and all boars were PRRSV qRT-PCR positive at DPI 4. A comparison of matched samples from individual boars showed that oral fluid was equal to serum for the detection of PRRSV at DPI 7 and more likely to be positive than serum on DPI 14 and 21. These data are reported in detail elsewhere [11].

To evaluate the association between viremia and antibody responses, cumulative serum and oral fluid PRRSV qRT-PCR (\log_e geq/ μ l) and antibody isotype (IgM, IgA, IgG) responses over time were re-expressed as area under the curve (AUC) (MedCalc[®]) prior to performing the analyses. The mean qRT-PCR and antibody isotype AUCs for serum and oral fluid are given in Table 3. Statistical analysis (ANOVA) of main effects showed significant differences among trials ($p < 0.001$), sample type ($p < 0.001$), and quantitative responses (PRRSV, IgM, IgA, IgG; $p < 0.001$). With the exception of serum IgG, statistically significant differences in means were detected among trials both in the replication of PRRSVs and in antibody responses (Tukey's Honestly Significant Difference (HSD) test). However, at the individual boar level, correlation analysis found a weak association between PRRSV viremia AUC and serum IgM, IgA, or IgG AUCs ($r = 0.3762$, 0.2915 , and 0.0005). Likewise, the correlation was weak between PRRSV oral fluid AUC and

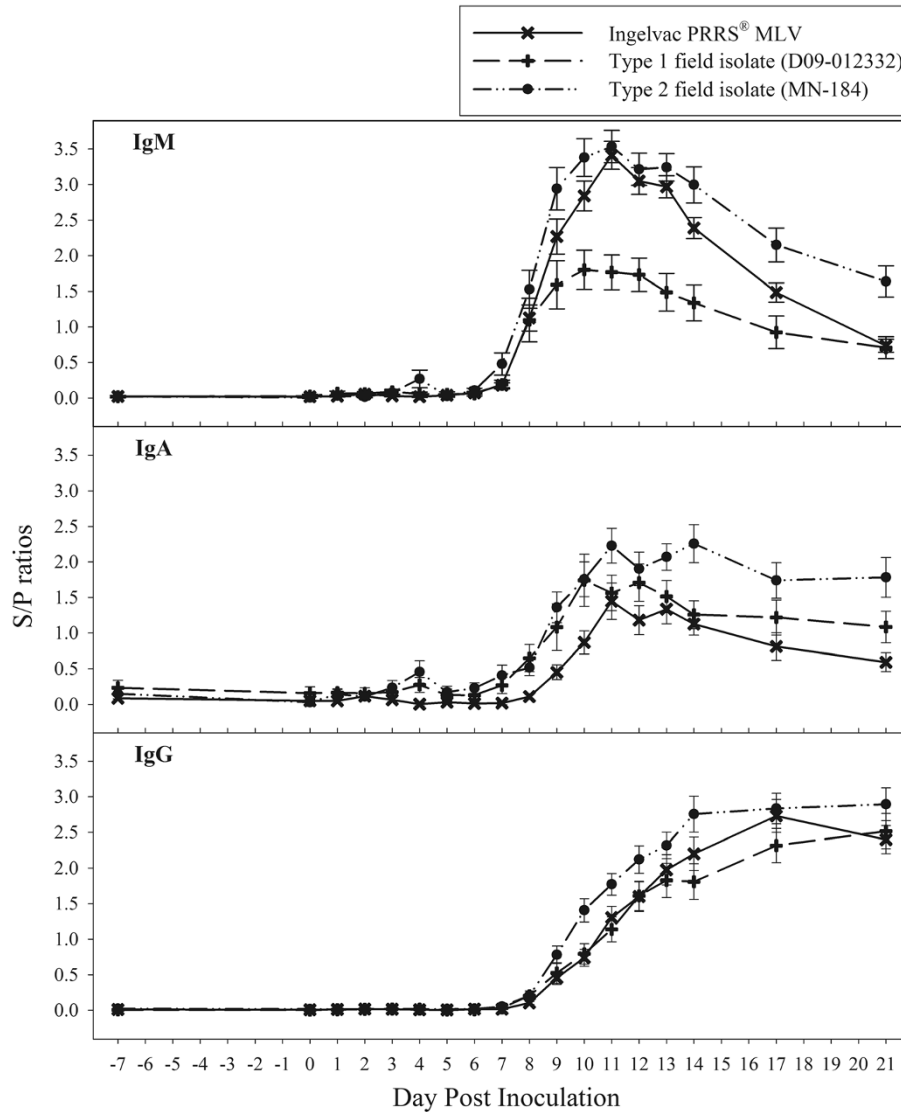


Figure 2 Kinetics of PRRSV antibody isotypes (IgM, IgA, and IgG) in oral fluid based on responses in 70 boars inoculated with 3 different PRRSV isolates. Results are reported as mean sample-to-positive (S/P) ratios and standard errors.

Table 2 Porcine reproductive and respiratory syndrome virus (PRRSV) serum^a and oral fluid^b ELISA qualitative results^c by day post inoculation (DPI)

Trial (virus isolate)	Sample	DPI 0 positive / tested	DPI 7 positive / tested	DPI 14 positive / tested	DPI 21 positive / tested
Trial 1: (Ingelvac [®] PRRS MLV)	Oral fluid	0 / 24	0 / 24	24 / 24	24 / 24
	Serum	0 / 24	0 / 24	24 / 24	24 / 24
Trial 2: (Type 1, D09-012332)	Oral fluid	0 / 22	0 / 22	17 / 21	22 / 22
	Serum	0 / 24	0 / 24	22 / 24	24 / 24
Trial 3: (Type 2, MN-184)	Oral fluid	0 / 24	0 / 23	24 / 24	22 / 22
	Serum	0 / 24	0 / 24	24 / 24	24 / 24
Total	Oral fluid	0 / 70	0 / 69	65 / 69	68 / 68
	Serum	0 / 72	0 / 72	70 / 72	72 / 72

^a PRRS X3 Ab Test (IDEXX Laboratories, Inc, Westbrook, Maine, USA) performed according to the manufacturer's instruction.

^b PRRS X3 Ab Test (IDEXX Laboratories, Inc, Westbrook, Maine, USA) modified to detect anti-PRRSV antibody in oral fluid specimens [5].

^c Samples with S/P ratio ≥ 0.4 were classified as positive.

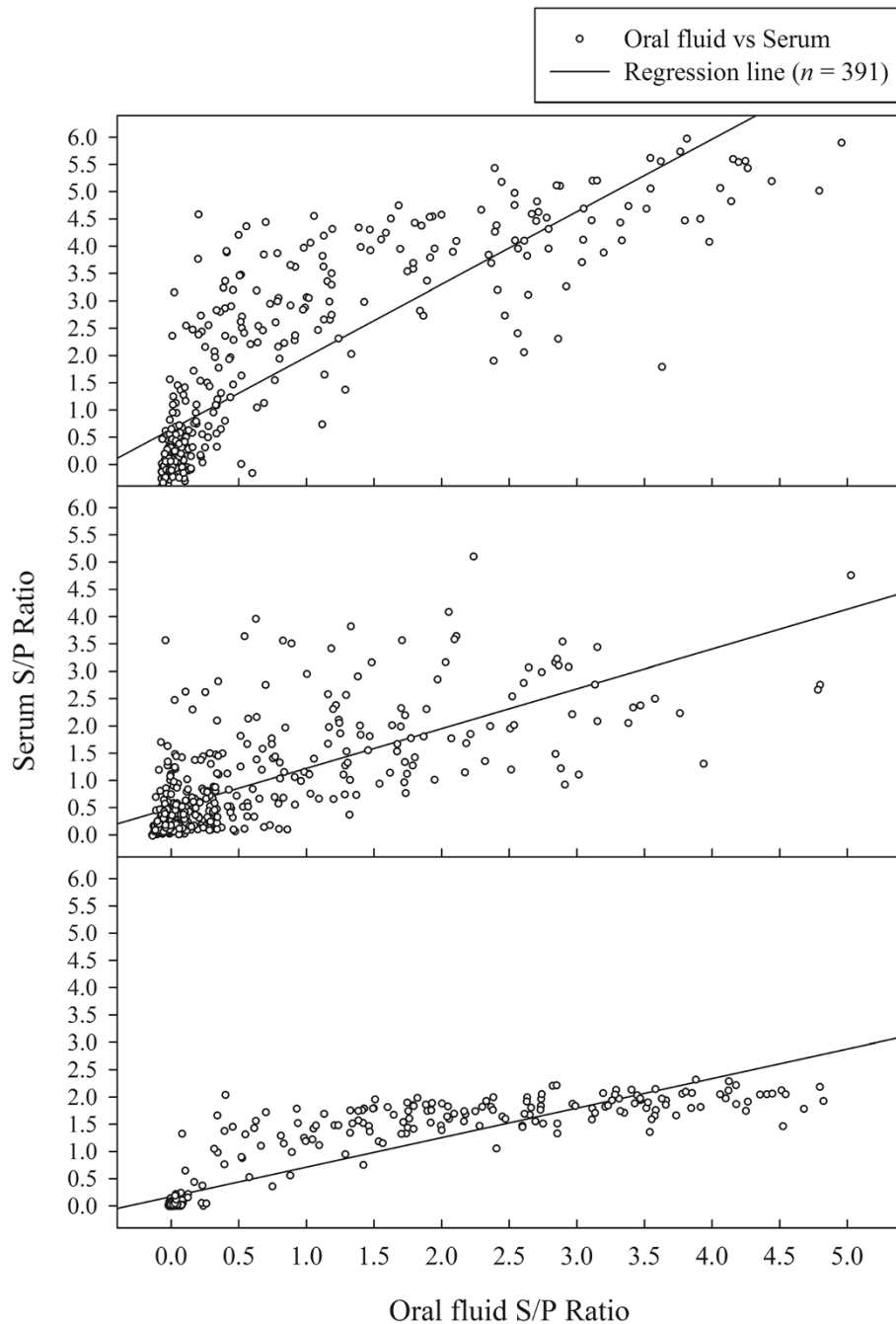


Figure 3 Correlation between serum and oral fluid PRRSV antibody isotypes (IgM, IgA, and IgG) based on results from individual boars.

oral fluid IgM, IgA, or IgG AUCs ($r = 0.3147$, 0.2671 , and 0.2137).

Discussion

By definition, oral fluid is a mixture of saliva, oral mucosal transudate, and gingival crevicular fluid recovered from the buccal cavity using an absorptive device [12]. Among a variety of other constituents, oral fluid contains

both locally-derived and systemic antibodies [13]. Thus, pathogen-specific IgM, IgA, and IgG for PRRSV [5], influenza A virus [7], and porcine circovirus type 2 [4] could be detected in oral fluid samples collected from groups of pigs (pens) under either experimental or field conditions. The largest proportion of locally produced antibody consists of dimeric secretory IgA (SIgA) produced by plasma cells in salivary glands and duct-associated lymphoid

Table 3 Comparison of cumulative quantitative reverse transcription polymerase chain reaction (qRT-PCR) and antibody responses (IgM, IgA, IgG) for 21 days following PRRSV inoculation¹

Trial (virus isolate)	Serum (mean AUC and 95% confidence intervals)				Oral fluid (mean AUC and 95% confidence intervals)			
	qRT-PCR	IgM	IgA	IgG	qRT-PCR	IgM	IgA	IgG
Trial 1: (Ingelvac [®] PRRS MLV)	21.6 ^b (18.8-24.5)	46.6 ^b (43.1-50.2)	18.5 ^b (14.6-22.6)	17.4 ^a (15.9-18.9)	21.0 ^b (17.0-25.1)	27.6 ^b (23.9-31.5)	12.3 ^b (7.7-17.0)	25.1 ^b (20.6-29.6)
Trial 2: (Type 1, D09-012332)	23.8 ^b (21.2-26.6)	39.5 ^b (31.1-48.0)	30.0 ^a (23.2-36.9)	18.3 ^a (16.5-20.2)	21.2 ^b (18.1-24.3)	17.6 ^c (12.0-23.2)	17.9 ^b (10.9-24.9)	21.9 ^b (16.7-27.2)
Trial 3: (Type 2, MN-184)	38.1 ^a (34.9-41.5)	53.7 ^a (48.1-59.3)	31.6 ^a (24.7-38.5)	18.4 ^a (17.1-19.85)	31.6 ^a (28.4-34.9)	35.4 ^a (28.6-42.3)	26.2 ^a (19.1-33.5)	30.1 ^a (25.3-34.9)

¹ Individual boar PRRSV (\log_e qeq/ μ l) and antibody (IgM, IgA, IgG) S/P responses over time were summarized as the area under the curve (AUC) prior to performing the statistical analysis.

^{abc} Superscripts within columns indicate statistically significant differences among means (Tukey's Honestly Significant Differences test, $p < 0.05$).

tissue (DALT) [14]. IgM and IgG are also produced in these tissues, but the majority of IgM and IgG in oral fluid is derived from serum via the gingival crevicular fluid [15].

In the present experiment, the collection of paired oral fluid and serum samples from 72 individually-housed boars inoculated with three different PRRSV isolates allowed for a more comprehensive evaluation of the onset and magnitude of serum and oral fluid antibody isotype responses, as well as the variation therein. Statistically significant differences in viral replication and antibody responses were observed among the three trials in both serum and oral fluid specimens (Table 3). Since each trial included only one virus isolate, statistically valid comparisons of the effects of virus isolates on PRRSV replication and antibody responses were not possible. Nevertheless, the statistically significant differences in virus replication and antibody response observed among trials was consistent with previous reports of virus isolate-dependent differences in the magnitude of replication in pigs [16] and corresponding differences in antibody response [16,17]. However, at the individual boar level, the correlation between virus replication and antibody response was relatively weak. Thus, what was true for the group of boars in a trial did not necessarily apply to an individual boar.

The purpose of this study was to describe and contrast the kinetics of PRRSV antibody in oral fluids and serum. PRRSV serum IgM, IgA, and IgG were first detected in samples collected on DPI 7, 10, and 10, respectively. These results were compatible with prior reports describing the detection of PRRSV serum IgM between DPI 5 and 7 [18-20], IgG between DPI 9 and 11 [19,21], and IgA at DPI 14 [19]. PRRSV oral fluid IgM, IgA, and IgG appeared concurrently with serum antibodies, but collection of daily oral fluid samples provided more precise estimates. That is, oral fluid IgM, IgA, and IgG were detected in samples collected between DPI 3 to 10, 7 to 10, and 8 to 14, respectively. There are no prior reports on PRRSV oral fluid antibody kinetics in individual

animals with which to compare these results. However, we previously reported the detection of PRRSV oral fluid IgM and IgG in pen-based oral fluid samples from experimentally inoculated animals on DPI 7 [5]. Thus, the PRRSV serum and oral fluid antibody responses observed in this study were in agreement with prior observations on PRRSV and our general understanding of the humoral immune response as reflected in these sample matrices [14,22]. Most significantly, this study provided a broader foundation for understanding, developing, and interpreting oral fluid antibody-based diagnostics in the context of the humoral immune response.

Conclusions

This study demonstrated that anti-PRRSV antibody isotypes can be detected in oral fluid specimens. These results were compatible with prior reports describing the detection of anti-PRRSV antibody in both serum and oral fluid. Detection of PRRSV antibody in oral fluids collected from individual boars could provide an effective approach for monitoring PRRSV infection in boar studs. Successful oral fluid collection and testing from individual boars suggests that approach could also be applied to population in swine production systems, i.e. pen-housed sows, farrowing crates, etc.

Methods

Experimental design

A total of 72 boars ranging from 6 months to 3.6 years of age under the ownership of PIC North America (Hendersonville, TN, USA) were used in this study. Housing, feed rations, animal care guidelines, and experimental protocols were approved and supervised by the PIC USA Health Assurance and Welfare department. In 3 trials of 24 boars each, animals were intramuscularly (i.m.) inoculated with either modified-live virus (MLV) PRRSV vaccine (Trial 1), a Type 1 field isolate (Trial 2), or a Type 2 field isolate (Trial 3). Serum and oral fluid samples were

collected from all boars beginning 7 days prior to inoculation and continuing through 21 days post inoculation (DPI). After the completion of Trial 3, samples were completely randomized and tested for PRRSV IgM, IgA, and IgG antibody isotypes. Descriptive and comparative statistical analyses were conducted to describe and compare PRRSV antibody responses in serum and oral fluid and evaluate differences among individual boars and between trials.

Animals and animal care

Boars were obtained from two Midwest USA breeding stock sources documented to be free of PRRSV infection. Culled boars ($n = 24$) ranged from one year to 3.6 years of age and select boars ($n = 48$) ranged from 5 to 6 months of age. The boars were housed in a commercial production facility equipped with nipple drinkers, concrete slatted flooring, curtains, and tunnel ventilation. Feeder space, water delivery, square footage per animal, sanitation, and ventilation parameters met or exceeded PIC North America health assurance and welfare requirements. Upon arrival, animals were housed individually in crates (Hog Slat, Inc., Newton Grove, NC USA) and fed a commercial corn/soy swine diet (Land O' Lakes[®] Farmland Feed, Roland, IA USA) at a rate of 4 pounds per animal per day for acclimation/training and 7 pounds per animal per day thereafter.

Porcine reproductive and respiratory syndrome viruses

In Trial 1, 24 boars were inoculated i.m. with 2 ml of a commercial MLV vaccine (Ingelvac[®] PRRS MLV, Boehringer Ingelheim Vetmedica, Inc., St. Joseph, MO 64506) rehydrated and administered according to the instructions provided by the manufacturer. In Trial 2, 24 boars were inoculated i.m. with 2 ml of a Type 1 PRRSV (isolate D09-012131) at an estimated concentration of $1 \times 10^{5.5}$ median tissue culture infectious dose (TCID₅₀) per ml. Isolate D09-012131 was isolated from serum samples submitted to the University of Minnesota Veterinary Diagnostic Laboratory (St. Paul, MN USA) in March 2009 as part of a routine monitoring program in a sow herd located in Illinois USA and propagated on pulmonary alveolar macrophage cells, as described elsewhere [11]. In Trial 3, 24 boars were IM inoculated with 2 ml of a Type 2 PRRSV isolate (MN-184, GenBank accession no. AY656992) at a concentration of $1 \times 10^{4.5}$ TCID₅₀ per ml. Isolate MN-184 (kindly provided by Dr. Scott Dee, University of Minnesota) was propagated on MARC-145 cells [23].

Sample collection

Oral fluid collection

Oral fluid samples were collected daily from individually-housed boars beginning 7 days prior to inoculation and continuing through 21 days post inoculation (DPI) using

a procedure described by Kittawornrat et al. [11]. In brief, oral fluid samples were collected by allowing boars to chew on 1.6 cm (5/8") cotton rope (Web Rigging Supply, Inc., Lake Barrington, IL USA). Prior to collection, ropes were soaked with a solution of sucrose and apple juice (unsweetened apple juice with 50% (v/v) sucrose) and then air-dried. To collect oral fluid samples, ropes were placed in "rope holders" fixed at the front of each pen for 20 minutes. Oral fluids were deposited as the boars chewed on the rope. To recover the oral fluid specimens, the bottom 15 cm (~6") of the rope (wet portion) was inserted into a 3.8 liter (one gallon) re-sealable plastic bag and severed from the dry portion of the rope. The bag with the wet rope inside was passed through a wringer (Dyna-Jet Products, Overland Park, KS USA), causing the fluid to pool in the bottom of the bag. Samples were then decanted into a 50 ml centrifuge tube and the volume recorded. Thereafter, samples were centrifuged at $1,000 \times g$ for 10 minutes at 4°C, aliquoted into 5 ml plastic tubes (Becton, Dickinson and Company, Bedford, MA USA), and stored at -80°C until assayed.

Serum collection

In each trial, serum samples were collected from all boars on DPI -7, 0, 7, 14, and 21. Additional serum samples were collected on DPI 3, 5, 10, 17 from a subset of boars ($n = 4$) randomly selected at the beginning of each trial. Blood was collected by jugular venipuncture using serum separation tubes (Corvac[®], Tyco Healthcare Group LP, Mansfield, MA USA). Samples were centrifuged at $1,000 \times g$ for 10 minutes and the serum was aliquoted into 5 ml plastic tubes (Becton, Dickinson and Company) and stored at -80°C until assayed.

PRRSV antibody ELISAs

Commercial PRRSV serum antibody ELISA All serum samples were assayed for PRRSV antibodies using a commercial indirect ELISA (PRRS X3 Ab Test, IDEXX Laboratories, Inc., Westbrook, ME USA) performed according to the manufacturer's instruction. As recommended by the manufacturer, a positive result was defined as a sample-to-positive (S/P) ratio ≥ 0.4 . Modifications to the commercial serum ELISA for the detection of antibody isotypes in serum and oral fluid are described below and listed in Table 4.

PRRSV antibody isotypes in serum The commercial indirect ELISA (PRRS X3 Ab Test) was modified to detect PRRSV-specific IgM, IgA, and IgG antibody isotypes in serum. In brief, serum samples were diluted 1:40 (5 μ l serum sample + 195 μ l kit diluent) for IgM and IgG and 1:5 (40 μ l serum sample + 160 μ l kit diluent) for IgA. 100 μ l of diluted serum was then transferred

Table 4 Summary of porcine reproductive and respiratory syndrome virus (PRRSV) serum and oral fluid antibody enzyme linked-immunosorbent assay (ELISA) condition^a

	IgM	IgA	IgG	Commercial ELISA
Serum ELISAs				
Sample dilution	1:40	1:5	1:40	1:40
Sample volume	100 µl	100 µl	100 µl	100 µl
Conjugate dilution	1:5,000 ^b	1:1,000 ^c	1:15,000 ^d	Provided with kit
Negative control	100 µl of pooled negative serum diluted 1:40	100 µl of pooled negative serum diluted 1:5	100 µl of kit negative control	100 µl of kit negative control
Positive control	100 µl of pooled serum from DPI 7 diluted 1:40	100 µl of pooled serum from DPI 21 diluted 1:5	100 µl of kit positive control	100 µl of kit positive control
Oral fluid ELISAs				
Sample dilution	1:2	1:2	1:2	
Sample volume	250 µl	250 µl	250 µl	
Conjugate dilution	1:3,800 ^b	1:2,000 ^c	1:2,400 ^d	
Negative control	250 µl of reference standard oral fluid ^e DPI 0 diluted 1:2	250 µl of reference standard oral fluid ^e DPI 0 diluted 1:2	100 µl of kit negative control diluted 1:30	
Positive control	250 µl of reference standard oral fluid ^e DPI 10 diluted 1:5	250 µl of reference standard oral fluid ^e DPI 91 diluted 1:2	100 µl of kit positive control diluted 1:30	

^a Oral fluid ELISA conditions represent modifications to a commercial PRRSV serum antibody ELISA protocol (IDEXX PRRS X3 Ab Test, IDEXX Laboratories, Inc., Westbrook, ME, USA).

^b Anti-pig IgM: HRPO conjugate (A100-100P, Bethyl Laboratories, Montgomery, TX, USA).

^c Anti-pig IgA: HRPO conjugate (A100-102P, Bethyl Laboratories, Montgomery, TX, USA).

^d Anti-pig IgG_{Fc}: HRPO conjugate (A100-104P, Bethyl Laboratories, Montgomery, TX, USA).

^e Reference standard oral fluid samples have been previously described [5].

to the PRRSV antigen-coated plates and incubated for 30 minutes at 22°C. After washing 3 times with 1X kit wash solution (400 µl), appropriately diluted horseradish peroxidase (HRPO)-conjugated anti-pig immunoglobulin (Ig) antibody (IgM (A100-100P), IgA (A100-102P), or IgG_{Fc} (A100-104P) (Bethyl Laboratories, Montgomery, TX USA) was added to each well and incubated for 30 minutes at 22°C. Thereafter, plates were washed three times with kit washing solution, after which 100 µl of tetramethylbenzidine (TMB) was added to each well and the plates incubated at 22°C for 15 minutes. At precisely 15 minutes, 100 µl of kit stop solution was added to each well. The plates were read at 650 nm using an ELISA plate reader (EL800 micro plate reader, Bio Tek[®] Instruments Inc., Winooski, VT) controlled by commercial software (Gen5[™] Bio Tek[®] Instruments Inc., Winooski, VT USA) and the reactions measured as optical density (OD).

PRRSV antibody isotypes in oral fluid Modification of the commercial PRRSV ELISA for the detection of PRRSV-specific IgM, IgA, and IgG antibody in swine oral fluid has previously been described [5]. In brief, oral fluid samples were diluted 1:2 (150 µl oral fluid sample + 150 µl kit diluent). 250 µl of diluted oral fluid was then transferred to PRRSV antigen-coated plates and incubated for 16 hours at 4°C. Thereafter, the plates were washed three times with 400 µl of 1X kit wash solution. To detect

the reaction, 100 µl of a solution containing appropriately diluted HRPO-conjugated anti-pig Ig (M, A, or G) was added to each well and the plates incubated for 30 minutes at 22°C. The procedure for determining the optimal dilution of secondary antibody is described in preparation of secondary antibody section. After washing three times, 100 µl of TMB was added to each well and the plates incubated at 22°C for 15 minutes. Finally, 100 µl of kit stop solution was added to each well. As described in preparation of secondary antibody section, the plates were read at 650 nm and the reactions measured as optical density (OD).

Preparation of secondary antibody

To assure assay repeatability, the concentration of anti-pig Ig (M, A, or G) was standardized using the positive control OD value listed in the manufacturer's Certificate of Analysis as the benchmark. The general procedure for calculating the conjugate dilution was as follows: 4 dilutions of anti-pig Ig (M, A, or G) were prepared in bottles wrapped in aluminum foil using diluent provided by manufacturer (IDEXX Laboratories, Inc., Westbrook, MA USA) and then stirred for 48 hours at 4°C. The reactivity of the 4 dilutions was determined using negative and positive kit controls. Specifically, kit negative control was dispensed into 48 wells (one-half plate) and kit positive control in each of the remaining 48 wells. Negative control OD values

were used to screen for non-specific reactions and positive control OD values were used to determine the equation of the line:

$$y = ax + c \quad (1)$$

where (y) is the anti-pig Ig (M, A, or G) OD response, (a) is the slope of the line, (x) is the dilution of anti-pig Ig (M, A, or G), and (c) is the intercept (Figure 4). Thereafter, the correct dilution of anti-pig Ig (M, A, or G) was calculated by substituting the mean positive control OD from the Certificate of Analysis for "y" in Equation 1 and solving for "x".

The general procedure for preparing the appropriate dilution of anti-pig Ig (M, A, or G) was modified for serum and oral fluid antibody ELISAs. In this study, all serum and oral fluid specimens were tested on a single lot of ELISA kits. Thus, the appropriate dilution of anti-pig Ig was calculated specifically for the PRRSV indirect ELISA (PRRS X3 Ab Test) kit lot (#99-40959-W721).

For the serum ELISA, the correct dilution of anti-pig IgG_{FC} was determined by titrating 4 dilutions of anti-pig IgG_{FC} (1:14,000, 1:14,500, 1:15,000, 1:15,500) against 100 µl of kit positive control and generating the equation of the line, as described above. The appropriate dilution of anti-pig IgM was determined by titrating 4 dilutions of secondary antibody (1:4,000, 1:4,500, 1:5,000, 1:5,500) against 100 µl of a positive control consisting of a 1:40 dilution of a pool of serum from 72 boars at DPI 7. For anti-pig IgA, 4 dilutions (1: 1,000, 1:2,500, 1:3,000, 1:3,500) were titrated against 100 µl of a positive control consisting of a 1:5 dilution of a pool of serum from 72 boars at DPI 21. Diluted

conjugate was simultaneously titrated against negative controls, i.e., kit negative control for anti-pig IgG_{FC} and a pool of serum from 72 boars at DPI 0 for anti-pig IgA (1:5) and anti-pig IgM (1:40).

The protocol for preparing the optimal secondary antibody dilution for the PRRSV oral fluid ELISA has been described elsewhere [5]. To prepare anti-pig IgG_{FC}, 4 conjugate dilutions (1:1,000, 1:1,500, 1:2,000, 1:2,500) were titrated against 100 µl of kit negative and positive controls diluted 1:30 (10 µl kit control + 290 µl kit sample diluent). Controls for anti-pig IgM and anti-pig IgA consisted of oral fluid samples from PRRSV-negative pigs vaccinated with 2 ml of PRRS modified live virus (MLV) vaccine (Ingelvac[®] PRRS MLV, Boehringer Ingelheim Vetmedica, Inc., St. Joseph, MO USA). The sample collected immediately prior to vaccination was used as negative control. Samples collected on 10 and 56 days post vaccination were used as positive controls for anti-pig IgM and anti-pig IgA conjugates, respectively. Four dilutions of anti-pig IgM (1: 3,000, 1:3,500, 1:4,000, 1:4,500) and anti-pig IgA (1:1,000, 1:1,500, 1:2,000, 1:2,500) were used to calculate appropriate dilution, as described previously.

Quantitative reverse transcription polymerase chain reaction (qRT-PCR)

Detailed qRT-PCR protocols for serum and oral fluids are reported elsewhere [11,24]. In brief, nucleic acid extraction from serum and oral fluid samples was performed using a commercial RNA extraction kit (Ambion[®] MagMax[™]-96 Viral RNA isolation kit, Applied Biosystems[™], Foster City, CA USA). Real-time PCR was performed with commercial reagent sets (TaqMAN[®] NA and EU PRRSV Reagents and TaqMAN[®] NA and EU PRRSV Controls, Applied Biosystems[™]) using the following cycling conditions: 1 cycle at 45°C for 10 minutes, 1 cycle at 95°C for 10 minutes, 40 cycles of: 97°C for 2 seconds, 60°C for 40 seconds. Eight 10-fold serially-diluted (10⁰ to 10⁷ copies/µl) plasmid-derived commercial standards (TaqMAN[®] NA and EU PRRSV RNA controls, Applied Biosystems[™]) were run on each PCR plate and their Ct values used to derive a standard curve. Samples were quantified as genome equivalents per µl (geq/µl) by fitting the sample Cts to the standard curve using the AB7500 Fast System SDS Software (Applied Biosystems[™]).

Statistical analysis

All optical density (OD) data were converted to sample-to-positive (S/P) ratios prior to statistical analysis using the following formula:

$$S/P = [\text{Sample A}(650) - \text{NC}] / (\text{PC} - \text{NC}) \quad (2)$$

where NC and PC represented the mean OD of the two negative control wells and two positive control wells,

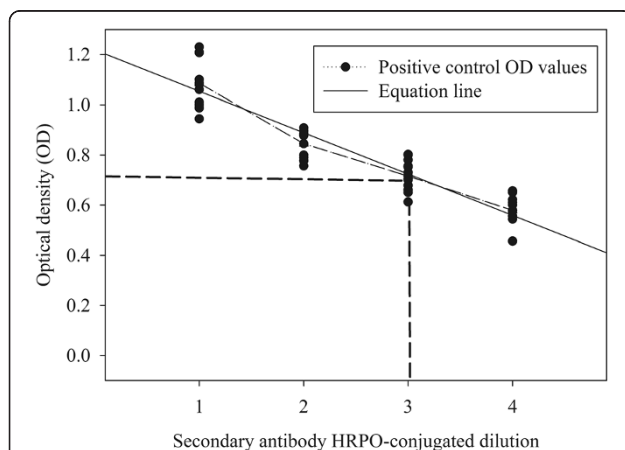


Figure 4 Calculation of the optimal dilution of anti-pig secondary antibody for PRRSV IgM, IgA, IgG ELISAs. The relationship between the positive control optical density (OD) and dilution of anti-pig secondary antibody was plotted as $y = ax + c$, where (y) is the secondary antibody OD response, (a) is the slope of the line, (x) is the dilution of secondary antibody, and (c) is the intercept. The appropriate dilution corresponds to the positive control value provided in the Certificate of Analysis.

respectively. Statistical analyses were performed using SAS[®] Version 9.2 (SAS[®] Institute Inc., Cary, NC USA) and MedCalc[®] 12.3.0.0 (MedCalc Software, Mariakerke Belgium).

Initially, descriptive and comparative analyses were conducted to describe the onset, magnitude, and duration of PRRSV antibody isotype S/P ratios in serum and oral fluid. Thereafter, S/P results were analyzed in a linear mixed model with repeated measures (SAS[®] PROC GLIMMIX) using trial (1, 2, 3), DPI, boar age (month), oral fluid volume (ml), and their pairwise interactions as fixed effects and “boar” as the subject of repeated measures. Tukey’s Honestly Significantly Difference (HSD) test was used to detect statistically significant differences between S/P ratios in each trial by DPI. Pearson’s correlation coefficient (SAS[®] PROC CORR) was used to evaluate the overall quantitative relationship between IgM, IgA, and IgG S/P ratios in serum and oral fluid samples. Defining an S/P ratio ≥ 0.4 as positive [5], McNemar’s test (SAS[®] PROC FREQ) for paired samples was used to determine whether the proportion IgG ELISA positive serum and oral fluid samples were significantly different by trial (1, 2, 3) and DPI.

The association between the level of PRRSV replication and the strength of the humoral response was evaluated in individual boars. The virus concentration (\log_e geq/ μ l) and antibody isotype S/P responses (IgM, IgA, IgG) in serum and oral fluid over the course of the experiment were re-expressed as the area under the curve (AUC; MedCalc[®]) and evaluated using

Pearson’s Correlation Coefficient (SAS[®] PROC CORR). In addition, the AUC data were evaluated for statistically significant differences among sample types (serum, oral fluid), trials (1, 2, 3), and quantitative responses (PRRSV, IgM, IgA, IgG) by analysis of variance (ANOVA). Sample type, trial, response, and their interaction were used as fixed effects in the model. Thereafter, Tukey’s Honestly Significant Differences (HSD) test was used to test for statistically significant differences among trial means.

Competing interests

Authors A. Rice and S. Lizano are employed by IDEXX Laboratories, Inc. The remaining author(s) declare no conflicting interests with respect to their authorship or the publication of this article.

Authors’ contributions

AK: data collection, antibody isotypes, manuscript preparation, and writing. ME: study conception, research design. YP: antibody isotypes, data collection. CO: antibody isotypes, data collection. KS: study conception, research design, and virus inoculation. AR: study conception. SL: study conception. CW: data analysis, study design. JZ: study design, manuscript preparation and writing. All authors read and approved the final manuscript.

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