Molecular phylogeny of the Pectinoidea (Bivalvia) indicates Propeamussiidae to be a non-monophyletic family with one clade sister to the scallops (Pectinidae)

G. Dalton Smedley  
_Iowa State University_, gsmedley@iastate.edu

Jorge A. Audino  
_University of São Paulo_

Courtney Grula  
_Iowa State University_

Anita Porath-Krause  
_Iowa State University_

Follow this and additional works at: [https://lib.dr.iastate.edu/eeob_ag_pubs](https://lib.dr.iastate.edu/eeob_ag_pubs)  
Part of the _Evolution Commons_, and the _Terrestrial and Aquatic Ecology Commons_

See next page for additional authors.  
The complete bibliographic information for this item can be found at [https://lib.dr.iastate.edu/eeob_ag_pubs/364](https://lib.dr.iastate.edu/eeob_ag_pubs/364). For information on how to cite this item, please visit [http://lib.dr.iastate.edu/howtocite.html](http://lib.dr.iastate.edu/howtocite.html).
Molecular phylogeny of the Pectinoidea (Bivalvia) indicates Propeamussiidae to be a non-monophyletic family with one clade sister to the scallops (Pectinidae)

Abstract
Scallops (Pectinidae) are one of the most diverse families of bivalves and have been a model system in evolutionary biology. However, in order to understand phenotypic evolution, the Pectinidae needs to be placed in a deeper phylogenetic framework within the superfamily Pectinoidea. We reconstructed a molecular phylogeny for 60 species from four of the five extant families within the Pectinoidea using a five gene dataset (12S, 16S, 18S, 28S rRNAs and histone H3). Our analyses give consistent support for the non-monophyly of the Propeamussiidae, with a subset of species as the sister group to the Pectinidae, the Propeamussiidae type species as sister to the Spondylidae, and the majority of propeamussiid taxa sister to the Spondylidae + Pr. dalli. This topology represents a previously undescribed relationship of pectinoidean families. Our results suggest a single origin for eyes within the superfamily and likely multiple instances of loss for these characters. However, it is now evident that reconstructing the evolutionary relationships of Pectinoidea will require a more comprehensive taxonomic sampling of the Propeamussiidae sensu lato.

Keywords
Pectinoidea, Pectinidae, Scallops, Propeamussiidae, Eyes

Disciplines
Ecology and Evolutionary Biology | Evolution | Terrestrial and Aquatic Ecology

Comments
This article is published as Smedley, G. Dalton, Jorge A. Audino, Courtney Grula, Anita Porath-Krause, Autum N. Pairett, Alvin Alejandrino, Latayshia Lacey et al. "Molecular phylogeny of the Pectinoidea (Bivalvia) indicates Propeamussiidae to be a non-monophyletic family with one clade sister to the scallops (Pectinidae)." Molecular phylogenetics and evolution 137 (2019): 293-299. doi: 10.1016/j.ympev.2019.05.006.

Rights
Works produced by employees of the U.S. Government as part of their official duties are not copyrighted within the U.S. The content of this document is not copyrighted.

Authors
G. Dalton Smedley, Jorge A. Audino, Courtney Grula, Anita Porath-Krause, Autum N. Pairett, Alvin Alejandrino, Latayshia Lacey, Felicity Masters, Peter F. Duncan, Ellen E. Strong, and Jeanne M. Serb

This article is available at Iowa State University Digital Repository: https://lib.dr.iastate.edu/eeob_ag_pubs/364
Molecular phylogeny of the Pectinoidea (Bivalvia) indicates Propeamussiidae to be a non-monophyletic family with one clade sister to the scallops (Pectinidae)

G. Dalton Smedleya, Jorge A. Audinob, Courtney Grulaa,1, Anita Porath-Krausea,2, Autum N. Pairetta, Alvin Alejandrinoa,3, Latayshia Laceyd, Felicity Mastersc, Peter F. Duncanc, Ellen E. Strongd, Jeanne M. Serba,*

⁎ Corresponding author at: Department of Evolution, Ecology, and Organismal Biology, Iowa State University, 2200 Osborn Dr., Room 251 Bessey Hall, Iowa State University, Ames, IA 50011, USA.

E-mail addresses: gsmedley@iastate.edu (G.D. Smedley), jorgeaudino@ib.usp.br (J.A. Audino), courtney.grula@ndsu.edu (C. Grula), aпорathkh@umn.edu (A. Porath-Krause), apairett@iastate.edu (A.N. Pairett), saleza@whittier.edu (A. Alejandrino), felicity.masters@research.usc.edu.au (F. Masters), pduncan@usc.edu.au (P.F. Duncan), StrongE@si.edu (E.E. Strong), serb@iastate.edu (J.M. Serb).

1 Present address: Department of Biological Sciences, North Dakota State University, Fargo, North Dakota, USA.

2 Present address: Department of Ecology, Evolution, and Behavior, 140 Gortner Lab, 1479 Gortner Ave, University of Minnesota, Saint Paul, MN, USA.

3 Present address: Department of Biology, Whittier College, 306 Science and Learning Center, Whittier, CA, USA.

ARTICLE INFO

Keywords: Pectinoidea Pectinidae Scallops Propeamussiidae Eyes

ABSTRACT

Scallops (Pectinidae) are one of the most diverse families of bivalves and have been a model system in evolutionary biology. However, in order to understand phenotypic evolution, the Pectinidae needs to be placed in a deeper phylogenetic framework within the superfamily Pectinoidea. We reconstructed a molecular phylogeny for 60 species from four of the five extant families within the Pectinoidea using a five gene dataset (12S, 16S, 18S, 28S rRNAs and histone H3). Our analyses give consistent support for the non-monophyly of the Propeamussiidae, with a subset of species as the sister group to the Pectinidae, the Propeamussiidae type species as sister to the Spondylidae, and the majority of propeamussiid taxa sister to the Spondylidae + Pr. dalli. This topology represents a previously undescribed relationship of pectinoidean families. Our results suggest a single origin for eyes within the superfamily and likely multiple instances of loss for these characters. However, it is now evident that reconstructing the evolutionary relationships of Pectinoidea will require a more comprehensive taxonomic sampling of the Propeamussiidae sensu lato.

1. Introduction

Scallops Pectinidae Rafinesque, 1815 are one of the most ecologically and morphologically diverse families in the class Bivalvia. With over 250 extant species currently considered valid, they are distributed across polar, temperate, and tropical marine ecosystems of shallow sublittoral reefs, sandy bays, sea grass beds and coarse substrates of the continental shelves, with a smaller number of species restricted to deeper water (Serb, 2016). Pectinidae is an ideal model to study the evolution of complex traits due to the number and biological diversity of extant species, the link between shell morphology and habitat use (Stanley, 1970), and their high preservability in the paleontological record (Valentine et al., 2006). Researchers have investigated the evolution of traits such as shell shape (Serb et al., 2011, 2017; Sherratt et al., 2016; Stanley, 1970), behavior (Alejandrino et al., 2011), swimming mechanics (Guderley and Tremblay, 2013; Hayami, 1991; Millward and Whyte, 1992; Tremblay et al., 2015), and phototransduction (Faggionato and Serb, 2017; Gomez et al., 2011; Kingston et al., 2015; Porath-Krause et al., 2016; Serb et al., 2013). One compelling set of phenotypes is the complex sensory systems, including eyes, found in this family (Audino et al., 2015a, 2015b, 2015c; Land, 1965; Speiser et al., 2011, 2016; Speiser and Johnsen, 2008). Most
work has concentrated on the eyes of scallops, which were first described in 1791 (Poli, 1791). Subsequent research focused on the anatomy and optics of these eyes to understand how the eyes capture light and focus images (Land, 1965; Palmer et al., 2017; Speiser et al., 2016; Speiser and Wilkens, 2016). Recent molecular approaches have provided insights into the evolution of gene families involved in scallop photoreception (Gomez et al., 2011; Kojima et al., 1997; Pairett and Serb, 2013; Piatigorsky et al., 2000; Porath-Krause et al., 2016; Serb et al., 2013). However, in order to understand the origin and evolution of these and other traits, the family Pectinidae needs to be placed in a deeper phylogenetic framework within the superfamily Pectinoidea.

The relationship of the Pectinidae to the other families in the Pectinoidea has been highly contentious due to high levels of homoplasy in shell characters (Dijkstra and Maestrati, 2012; Hertlein, 1969) and alternative interpretations of the fossil record (Waller, 2006, 1991, 1978) (Fig. 1). As a result, three families (Propeamussiidae, Spondylidae, Entoliidae) singly or in combination have been proposed to be the sister taxon to the Pectinidae by different authors at different times. The prevailing view has been that the Propeamussidae Abbott, 1954, or glass scallops (~200 species), represent the closest relatives of the Pectinidae. Propeamussioids possess very thin, often translucent shells and inhabit the marine epipelagic (80 m) to the abyssal (4000 m) zones. They appear to be a lineage of relict species that survived severe environmental changes at the end of the Cretaceous by inhabiting deep and/or cold-water refugia (Waller, 1991) where most modern propeamussiids and the oldest extant lineage of Pectinidae (Camptonectinidae: Delectopetcen) are still found. Additionally, propeamussiids and some pectinid lineages have a similar shell shape. These data suggest a possible sister relationship between the two families, which has been supported by other studies which include molecular data for their phylogenetic analyses (Bieler et al., 2014, Fig. 30; Matsumoto and Hayami, 2000) (Fig. 1A). Recently, one lineage of micro glass scallops (1.5-6 mm as adults) was elevated to its own family, the Cyclochlamydidae. Samples used in this study were obtained from colleagues and museum collections (see supplementary Table S1 and Acknowledgments). The majority of Indo-Pacific specimens included in this study were obtained during expeditions organized by the MNHN and Pro-Natura International as part of the Our Planet Reviewed program, and by the MNHN and the Institut de Recherche pour le Développement as part of the Tropical Deep-Sea Benthos program. Species identifications of the Indo-Pacific specimens were determined by Henk H. Dijkstra at the Naturalis Biodiversity Center (Netherlands). All tissues were preserved in ethanol and shell voucher specimens are available from museum collections listed in supplementary Table S1.

2. Materials and methods

2.1. Specimens and samples

We assembled 60 taxa from four of the five extant families in the superfamily Pectinoidea plus five species of Limidae to serve as the outgroup. We sampled 18 species from the Propeamussiidae, 37 species of Pectinidae, four species of Spondylidae, and a single extant species of Entoliidae (supplementary Table S1). Due to the challenges of acquiring samples, we were unable to include taxa from the newly described family Cyclochlamydidae. Samples used in this study were obtained from colleagues and museum collections (see supplementary Table S1 and Acknowledgments). The majority of Indo-Pacific specimens included in this study were obtained during expeditions organized by the MNHN and Pro-Natura International as part of the Our Planet Reviewed program, and by the MNHN and the Institut de Recherche pour le Développement as part of the Tropical Deep-Sea Benthos program. Species identifications of the Indo-Pacific specimens were determined by Henk H. Dijkstra at the Naturalis Biodiversity Center (Netherlands). All tissues were preserved in ethanol and shell voucher specimens are available from museum collections listed in supplementary Table S1.

2.2. Molecular laboratory methods

Total genomic DNA (gDNA) was extracted from either mantle or adductor tissues following the manufacturer’s protocol of the Qiagen DNeasy Blood and Tissue kit. A portion of the nuclear gene 18S ribosomal RNA (~700 bp) was amplified using the 18S a2.0 forward (5′-ATGGTGCAAGCTGGAA-3′) and 18S b reverse (5′-GGTACCTCTGGCAGGTTCAC-3′) primers (Giribet et al., 1996; Whiting et al., 1997). PCR reactions were carried out in 25 µl total volume reactions containing 12.5 µl 2x MyTaq Red Mix (Bioline), 1 µl of 10 µM 18S rRNA forward and reverse primers (18s a2.0 and 18s b, respectively), 9.5 µl double distilled water, and 1 µl of template. Reactions underwent one round of PCR consisting of an initial denaturation step (2 min at 95 °C) followed by 30 cycles of chain denaturation (15 s at 95 °C), primer annealing (15 s at 50 °C), and elongation (10 to 60 s at 72°C). Roughly 5 µl of the amplification products were visualized on a 2% agarose gel using a 1 kb size standard. Samples with the expected band size (~700 bp) were sent to Iowa State University DNA Facility for Sanger sequencing using Applied Biosystems 3730 × 1. In total, 18S rRNA sequences for 60 taxa (16 Propeamussiidae species, 35 Pectinidae species, three Spondylidae species, one Entoliidae species, and five Limidae species) were successfully generated.

The 18S rRNA sequences were added to a multigene dataset consisting of two mitochondrial genes (12S and 16S rRNAs) and two nuclear genes (28S rRNA and histone H3) from previously published work from our lab (Alejandrino et al., 2011) (supplementary Table S1). To complete the dataset, we generated DNA sequences for 28S rDNA and histone H3 of Pectinella aequorita (Entoliidae) using the methods in
Alejandrino et al. (2011).

2.3. Phylogenetic analyses

DNA sequences for each gene portion were aligned separately in MAFFT v7.222 (Katoh and Standley, 2013) using the automatic algorithm to select the best alignment method and remaining settings/options set as default. Ambiguously aligned nucleotides due to large insertion-deletions (indels) in 12S, 16S, and 28S rRNA genes were removed using settings for a less stringent selection on the Gblock server (Castresana, 2000; Dereeper et al., 2008; Talavera and Castresana, 2007). Individual gene alignments were concatenated in Geneious v4.7.6 (Kearse et al., 2012) to produce a final dataset of five gene regions: 12S rRNA (1–315 bp), 16S rRNA (316–674 bp), 18S rRNA (675–1161 bp), 28S rRNA (1162–1937 bp), and histone H3 (1938–2276 bp). Mitochondrial-only (12S and 16S rRNAs) and nuclear-only (18S rRNA, 28S rRNA and histone H3) datasets were also produced.

Phylogenetic analyses were carried out under maximum likelihood (ML: (Felsenstein, 1981)) and Bayesian inference (BI: (Mau et al., 1999)). Nucleotide substitution model was determined using PartitionFinder2 (Lanfear et al., 2016). For this analysis, the datablock was defined by gene, as above, with branch lengths unlinked. All evolution models and schemes were investigated using Akaike Information Criterion with sample size correction (AICc) metric. ML analyses were conducted using RAxML-HPC v8.2.9 on XSEDE (Stamatakis, 2014) as implemented on the CIPRES Scientific Gateway v3.3 (Miller et al., 2010). Branch support was determined with 500 bootstrap iterations for best-scoring ML tree. All other parameters were set at the program’s default. BI analyses were conducted using MrBayes v3.2.6 (Ronquist et al., 2012) as implemented on the CIPRES Scientific Gateway v3.3. We ran three independent analyses, each with eight Markov chain Monte Carlo (MCMC) chains sampling every 100 generations and the temperature for heated chains set at 0.15. The MCMC analysis was set to run for 50 million generations or until a standard deviation of split frequency value of 0.01 was reached signifying convergence following the stoprule after 4.2 million generations. The post-run analyses were set with a 50% burn-in and all other parameters not mentioned above were left at the program’s default. We then visually inspected the combined trace files to confirm acceptable mixing and highESS (effective sampling size) across all parameters (≥ 300) in Tracer v1.6 (Rambaut et al., 2018). Post-burn-in trees were used to construct the 50% majority rule consensus tree and to estimate posterior probabilities.

We used the Approximately Unbiased (AU) test (Shimodaira, 2001) to compare our results to six alternative phylogenetic hypotheses. These alternative topologies were generated via ML in RAXML to constrain either (1) a monophyletic Propeamussiidae or (2) a clade of Propeamussiidae that excluded Parvamussium ina. In addition, four hypotheses from previous studies (Fig. 1) were compared. Site-wise likelihoods were calculated in RAXML for the unconstrained and constrained ML topologies and analyzed in CONSEL (Shimodaira and Hasegawa, 2001) using default parameters for p-values.

Divergence time estimation was conducted using RevBayes version 1.0.9 under the Fossilized Birth-Death model (Hohna et al., 2016). A relaxed molecular clock model was defined assuming an uncorrelated exponential model on branch rates. Posterior probabilities were sampled by Markov chain Monte Carlo process (MCMC) for 500,000 iterations. Maximum clade credibility tree, with a burn-in of 10%, was generated after pruning the five fossil taxa used to calibrate internal nodes. Fossil ages were incorporated based on available data in Waller (2006) and in the Paleobiology Database (https://paleobdb.org/). Priors for fossil ages were drawn from uniform distributions and the root (Pectinoidea + Limidae) was constrained between 485.4 and 419.2 MYA (million year ago). The age of Pectinidae was constrained around 251.3–247.2 MYA based on the fossil of Prachlamys spp., also considering the fossil record of Argopecten spp. (15.99–2.61 MYA), an extant genera. The Spondylidae was constrained around 171.6–168.3 while Entoliidae was calibrated based on the fossil of Pectinella spp. (251.3–247.2 MYA). Finally, the Limidae was also constrained between 330.9 and 323.2 MYA, based on Paleolima spp.

3. Results

A total of 111 sequences were generated in this study and 196 sequences were obtained from previous work (Sherritt et al., 2016) for 60 species across four families of Pectinoidea with five species of Limidae serving as the outgroup. The lengths of each gene region after alignment were: 12S rRNA: 315 bp; 16S rRNA: 359 bp; 18S rRNA: 487 bp; 28S rRNA: 776 bp; histone H3: 339 bp. DNA sequences were deposited in GenBank (NCBI accession numbers MH MH463998-MH464109; Table S1). Our concatenated five-gene dataset had a total aligned length of 2276 bp. The molecular dataset was complete for 54 of the 65 taxa, while the remaining 11 taxa lacked at least one gene. Incomplete gene sets occurred in some species from all four families of Pectinoidea, but there was no pattern based on taxonomic membership (Supplementary Table 1). PartitionFinder 2 suggested a four partition scheme. A GTR + G evolution model was suggested for 125 and 16S partitions and a GTR + I + G evolution model for 28S and 18S + H3 partitions. However, after 200 million generations, the MrBayes analyses still had not reached convergence suggesting the PartitionFinder scheme too complicated given the dataset, requiring us to use a less complicated substitution model. A general time reversible (GTR) model with gamma-distributed rates across nucleotide sites was applied to ML and both BI analyses using the gene partitions described above.

ML and BI analyses of the concatenated five gene dataset reconstructed the same five lineages of pectinoidean taxa and produced similar topologies (Fig. 2 for ML; Fig S1 for BI phylogram). The only difference between the two topologies was that the Bayesian analysis was unable to resolve the relationships among the five pectinoidean clades. Interestingly, the relationships among these clades in the ML topology did not match any of the proposed phylogenetic hypotheses for Pectinoidea (Fig. 1). The single representative of Entoliidae (Pectinella acquisor) was recovered as sister to the remaining pectinoideans in the ML tree with high support (100% BS). The Propeamussiidae is not monophyletic, with the majority of the species (n = 13) forming a clade with low support (64% BS, 78 PP). The type species, Propeamussium dalli was not a member of this clade, but rather the sister group (55% BS, 72 PP) to a well-supported monophyletic Spondylidae (100% BS; 100 PP). A third propeamussiid lineage of three species was a moderately supported clade that was the sister group to the Pectinidae (64% BS, 86 PP), and a fourth was represented by Parvamussium ina nested within the Pectinidae (66% BS, 89 PP). Thus, the Pectinidae as currently conceived is paraphyletic in our analyses, and the Propeamussiidae polyphyletic. Non-monophyly of the Propeamussiidae was also supported in ML and BI analyses of the mitochondrial-only and nuclear-only datasets (Figs. S2–S5).

Using the best tree from each ML analysis, AU tests were performed to statistically compare our results against competing hypotheses that constrain the Propeamussiidae as monophyletic and that constrain the Propeamussiidae as monophyletic to the exclusion of Parvamussium ina. Additionally, we compared our results with four alternative sister groups for the Pectinidae described in previous studies (Fig. 1). The AU test significantly rejected (p-values < 0.01) the hypotheses with a monophyletic Propeamussiidae + Pectinidae (Fig. 1A), Spondylidae + Pectinidae (Fig. 1B), and Entoliidae + Pectinidae (Fig. 1C) (Table 1).

We estimated divergence dates among extant taxa using five fossil calibration points with horizontal bars representing the 95% highest posterior density (HPD) intervals for each node (Fig. 3). The resulting topology recovered similar relationships to topologies derived from the RAXML (Fig. 2) and MrBayes analyses (Fig. S1) with two exceptions.
Fig. 2. Maximum likelihood phylogeny of pectinoidean families (lnL = \(-25647.23\)) based on combined 12S, 16S, 18S, 28S and histone H3 sequences. Taxa are color coded by family. Numbers above the branches indicate bootstrap support; numbers below branches are Bayesian posterior probabilities. A dash (\(\_\)) indicates no support for that node.
First, the time-calibrated phylogeny recovered the Entoliidae taxon as sister to largest Propeamussiidae clade with an inferred divergence time of approximately 300.3 MYA (Late Carboniferous). Second, the Pectinidae is monophyletic in the time-calibrated phylogeny as Parva-

**Table 1**

Summary of AU tests of alternativepectinoidean topologies.

<table>
<thead>
<tr>
<th>Topology</th>
<th>AU test p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Unconstrained ML</td>
<td>0.599</td>
</tr>
<tr>
<td>Monophyletic Propeamussiidae</td>
<td>0.323</td>
</tr>
<tr>
<td>Monophyletic Propeamussiidae, excluding Pa. ina</td>
<td>0.574</td>
</tr>
<tr>
<td>Fig. 1A Hypothesis</td>
<td>0.011</td>
</tr>
<tr>
<td>Fig. 1B Hypothesis</td>
<td>0.014</td>
</tr>
<tr>
<td>Fig. 1C Hypothesis</td>
<td>0.005</td>
</tr>
<tr>
<td>Fig. 1D Hypothesis</td>
<td>0.069</td>
</tr>
</tbody>
</table>

*RAXML constraint analyses and corresponding p-values of AU tests implemented in CONSEL. Significantly different topologies are in bold.

4. Discussion

Four different sister group relationships to the scallops have been hypothesized based on morphological evidence spanning the paleo- and neontological record or from molecular data. A traditional interpretation of shell similarity between the Propeamussiidae and some scallop...
taxa led some to conclude a sister group relationship between the two families (Fig. 1A). However, morphological comparison of fossil and Recent taxa and re-interpretation of first occurrences in the fossil record have been the basis of three other possible topologies. Waller (1978) proposed Spondylidae + Entoliidae (= Syncyclonemidae) to be the sister taxon of the Pectinidae based on a single synapomorphy of lip morphology, but noted that these taxa have many primitive features and resemble the fossil precursors to the Pectinidae more than the extant members (Fig. 1D). Subsequently, Waller (1991) presented a revised hypothesis with the Entoliidae alone as the sister to the Pectinidae (Fig. 1C). Most recently, fossil evidence from the Mesozoic appears to bridge morphological gaps among pectinoidean lineages (Waller, 2006). This and recognition of a “pectiniform” in an early stage of spondylid growth led Waller (2006) to propose the Spondylidae as the sister lineage to the Pectinidae (Fig. 1B). Two of these hypotheses (Fig. 1C, D) place the Propeamussiidae as sister to all other Pectinoidea. Interestingly, molecular phylogenetics has largely supported a fifth relationship, with the Propeamussiidae + Spondylidae as the sister group to the Pectinidae. Our estimated phylogenies show both a propeamussiid clade sister to the Pectinidae as well as a second propeamussiid lineage that shares a common ancestor with a monophyletic Spondylidae (Figs. 2 and 3). Thus, our data support the traditional hypothesis, in part, but highlights two important future directions. First, the non-monophyly of the Propeamussiidae suggests that the characteristics that have been used as synapomorphies for the family should be re-examined. Second, if the relationship between Spondylidae and Propeamussium dalli (the type species) holds, a taxonomic revision of the Propeamussiidae will be necessary.

Few published time-calibrated phylogenies have included the Pectinoidea, and those that do have been inferred from a small subset of pectinoidean taxa (e.g., Bieler et al. 2014). In contrast, our estimation of divergence times for the Pectinoidea is based on a larger taxonomic sampling that includes four of the five families and fossil taxa from three of these families (Entoliidae, Pectinidae, and Spondylidae). Through this sampling strategy, we were able to independently estimate age of the superfamily. Interestingly, our time-calibrated phylogeny supports a somewhat earlier origin of the Pectinoidea (Late Devonian, 395 MYA) than currently accepted date of the Early Carboniferous period (358.9 MYA) when †Pernepcticoidina is regarded as the stem group of the superfamily (see Waller 2006). Future inclusion of fossil taxa in phylogenetically informed macroevolutionary analyses will be critical for interpreting patterns of diversification and extinction for the group.

Understanding relationships among the families of Pectinoidea could give an interesting context to the evolution of eyes within the superfamily. Eyes occur ventrally and often serially repeated on both left and right mantle lobes, located at the end of short stalks on the middle fold (Dakin, 1910). Scallops possess many single chambered eyes with a mirror-like reflector lining the back of the eye which focuses light back onto a double-retina system in the middle of the eye (Land, 1965; Palmer et al., 2017). Pectinidae and Spondylidae are known to have this unique eye structure, while Propeamussiidae were thought to lack eyes (Waller, 1972); however, the absence of eyes in propeamussiids may reflect their distribution in dysphotic (200–1000 m) or aphotic (> 1000 m) depths (Waller, 2006, but see Morton and Thurston, 1989). There has been some debate regarding the presence or absence of eyes in extinct entoliids. Eyes may be present in the extant genus Pectinella ([Waller, 2006] images of the eyes were not illustrated), but with only two extant species, fluid-preserved specimens are rare (e.g., no specimens in the largest US collection USNM, co-authored EE Strong) and we have been unable to secure a specimen for examination. If eyes are present in the Entoliidae (Waller, 2006) and our phylogenetic hypothesis is corroborated with future analyses, its placement as the sister taxon to the remaining Pectinoidea suggests a single origin of eyes in the common ancestor of the superfamily. However, patterns of eye loss in the Propeamussiidae sensu lato need to be examined from both historical and habitat perspectives.

5. Conclusion

The results of these current analyses suggest a novel topology for relationships within the superfamly Pectinoidea. Our results tentatively indicate the Propeamussiidae may be polyphyletic, but the AU test results do not reject all alternative hypotheses in which the family is constrained to be monophyletic. The inclusion of molecular data for a species of Entoliidae for the first time provides the first test of its phylogenetic placement as the sister to all other Pectinoidea. Our phylogenetic hypothesis also impacts the interpretation of trait histories in the superfamily with implications to phenotypic evolution. For instance, our data tentatively supports the hypothesis for a single origin of eyes in the superfamily. Future work should focus on bolstering support for this scenario through the examination of a more comprehensive molecular dataset. However, if the relationships recovered here hold, a taxonomic revision of the Propeamussiidae is warranted.

Acknowledgements

The majority of Indo-Pacific specimens were obtained during expeditions (PI Philippe Bouchet) organized by the MNHN and Pro-Natura International as part of the Our Planet Reviewed program, and by the MNHN and the Institut de Recherche pour le Développement as part of the Tropical Deep-Sea Benthos program. Ship time was programmed on R.V. Alis by the French Oceanographic Fleet, on R.V. Vizconde de Eza by the Instituto Español de Oceanografía, on F.V. DA-BFAR by the Philippines Bureau of Fisheries and Aquatic Resources, and on chartered local fishing boats through funding from the Total Foundation and Prince Albert II of Monaco Foundation. The loans of molecular samples from MNHN were arranged by Nicolas Puillandre and Barbara Buge. We are very grateful for the assistance and loans provided by the staff of museums and research institutions, especially G. Paulay and J. Slapcinsky [UF]; S. Morrison and C. Whisson [WAM]; P. Greenhall and C. Walter [NMNH]; A. Baldinger [MCZ]; M. Siddall and S. Lodhi [AMNH]; R. Bieler and J. Gerber [FMNH]; R. Kawamoto [BPBM]; E. Kools [CAS]. Members of the D.C. Adams, T.A. Heath, and N. Valenzuela labs provided valuable comments on an earlier version of this manuscript. The authors are grateful to two anonymous reviewers for helpful input. Authors declare no conflict of interest.

Funding

Financial support was provided by the United States of America National Science Foundation [NSF DEB-1118884 to JMS]. Funding for specimen collections was provided by the Total Foundation, Prince Albert II of Monaco Foundation, Stavros Niarchos Foundation, Richard Lounsbery Foundation, the French Ministry of Foreign Affairs, and the Philippines Bureau of Fisheries and Aquatic Resources. The funding bodies did not play a role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Appendix A. Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.ympev.2019.05.006.

References