

2015

# Performance of an Infrared Photoacoustic Single Gas Analyzer in Measuring Ammonia from Poultry Houses

Hong Li  
*University of Delaware*

Chen Zhang  
*University of Delaware*

Hongwei Xin  
*Iowa State University, hxin@iastate.edu*

Follow this and additional works at: [http://lib.dr.iastate.edu/abe\\_eng\\_pubs](http://lib.dr.iastate.edu/abe_eng_pubs)

 Part of the [Agriculture Commons](#), [Bioresource and Agricultural Engineering Commons](#), and the [Poultry or Avian Science Commons](#)

The complete bibliographic information for this item can be found at [http://lib.dr.iastate.edu/abe\\_eng\\_pubs/651](http://lib.dr.iastate.edu/abe_eng_pubs/651). For information on how to cite this item, please visit <http://lib.dr.iastate.edu/howtocite.html>.

---

This Article is brought to you for free and open access by the Agricultural and Biosystems Engineering at Iowa State University Digital Repository. It has been accepted for inclusion in Agricultural and Biosystems Engineering Publications by an authorized administrator of Iowa State University Digital Repository. For more information, please contact [digirep@iastate.edu](mailto:digirep@iastate.edu).

---

# Performance of an Infrared Photoacoustic Single Gas Analyzer in Measuring Ammonia from Poultry Houses

## Abstract

A single-gas photoacoustic analyzer, Chillgard RT, was evaluated for its performance of measuring ammonia ( $\text{NH}_3$ ) concentrations in poultry production under laboratory and field conditions, using a multi-gas photoacoustic analyzer, INNOVA 1412, as a reference method. Calibration gases were used to evaluate the cross-interferences of carbon dioxide ( $\text{CO}_2$ ), nitrous oxide ( $\text{N}_2\text{O}$ ), methane ( $\text{CH}_4$ ), and water vapor on  $\text{NH}_3$  measurements by Chillgard RT. The response times of Chillgard RT and INNOVA were measured in the laboratory using four vessels containing broiler litter. Side-by-side field comparisons between the Chillgard RT and INNOVA were conducted on a laying-hen farm and a broiler farm over two five-month periods. A strong linear relationship existed between the Chillgard RT and INNOVA  $\text{NH}_3$  readings. The  $\text{NH}_3$  measurement by the Chillgard RT showed no effect by  $\text{CO}_2$ ,  $\text{N}_2\text{O}$ ,  $\text{CH}_4$ , or water vapor under typical operational conditions in poultry operations. The Chillgard RT demonstrated a faster response to  $\text{NH}_3$  than the INNOVA. Following 120-s measurements, the Chillgard RT and INNOVA achieved 99.6% and 96.7% of the expected values, respectively, when the INNOVA was configured at a 30-s sampling interval. The INNOVA response time was positively affected by water vapor level in the air samples. The Chillgard RT overestimated  $\text{NH}_3$  concentrations by 3.24%, 10%, and 22.9% for laying-hen houses, stored boiler litter, and broiler houses, respectively, as compared to the INNOVA. Thus, performance of the Chillgard RT should be carefully validated before used in field  $\text{NH}_3$  measurements.

## Keywords

Ammonia, Analyzer, Gas Measurement, Photoacoustic, Poultry

## Disciplines

Agriculture | Bioresource and Agricultural Engineering | Poultry or Avian Science

## Comments

This article is from *Applied Engineering in Agriculture* 31 (2015): 471–477, doi:[10.13031/aea.31.10826](https://doi.org/10.13031/aea.31.10826).  
Posted with permission.

## PERFORMANCE OF AN INFRARED PHOTOACOUSTIC SINGLE GAS ANALYZER IN MEASURING AMMONIA FROM POULTRY HOUSES

H. Li, C. Zhang, H. Xin

**ABSTRACT.** *A single-gas photoacoustic analyzer, Chillgard RT, was evaluated for its performance of measuring ammonia (NH<sub>3</sub>) concentrations in poultry production under laboratory and field conditions, using a multi-gas photoacoustic analyzer, INNOVA 1412, as a reference method. Calibration gases were used to evaluate the cross-interferences of carbon dioxide (CO<sub>2</sub>), nitrous oxide (N<sub>2</sub>O), methane (CH<sub>4</sub>), and water vapor on NH<sub>3</sub> measurements by Chillgard RT. The response times of Chillgard RT and INNOVA were measured in the laboratory using four vessels containing broiler litter. Side-by-side field comparisons between the Chillgard RT and INNOVA were conducted on a laying-hen farm and a broiler farm over two five-month periods. A strong linear relationship existed between the Chillgard RT and INNOVA NH<sub>3</sub> readings. The NH<sub>3</sub> measurement by the Chillgard RT showed no effect by CO<sub>2</sub>, N<sub>2</sub>O, CH<sub>4</sub>, or water vapor under typical operational conditions in poultry operations. The Chillgard RT demonstrated a faster response to NH<sub>3</sub> than the INNOVA. Following 120-s measurements, the Chillgard RT and INNOVA achieved 99.6% and 96.7% of the expected values, respectively, when the INNOVA was configured at a 30-s sampling interval. The INNOVA response time was positively affected by water vapor level in the air samples. The Chillgard RT overestimated NH<sub>3</sub> concentrations by 3.24%, 10%, and 22.9% for laying-hen houses, stored boiler litter, and broiler houses, respectively, as compared to the INNOVA. Thus, performance of the Chillgard RT should be carefully validated before used in field NH<sub>3</sub> measurements.*

**Keywords.** *Ammonia, Analyzer, Gas Measurement, Photoacoustic, Poultry.*

High levels of ammonia (NH<sub>3</sub>) in animal houses could adversely affect the health of animals and farm workers. Aerial NH<sub>3</sub> is considered a precursor in fine particulate matter (PM) formation; and its deposition can affect ecological nitrogen balance and cause eutrophication (Asman et al., 1998). Measurement of NH<sub>3</sub> concentration is essential for animal feeding operations (AFOs) management and emission determination and mitigation. Various types of analytical instruments based on different detection principles have been evaluated and used to measure NH<sub>3</sub> concentration in AFOs. Colorimetric detection tube (CDT) and passive samplers (PS) are inexpensive but only suitable for semi-

quantitative, time-weighted-average measurements with limited sampling frequency and points. Electrochemical (EC) sensor and chemical-cassette (CC) tape based colorimetric analyzer provide economical solutions to continuous measurement due to their relatively low cost and long life span (Liang et al., 2004). Fourier transform infrared (FTIR) and chemiluminescence (CL) based analyzers offer best accuracy and sensitivity but are expensive and normally require a clean and controlled environment to operate. Other infrared-based technologies, such as photoacoustic spectrometer (PAS), near-infrared (NIR), open-path laser (OPL), have been widely used for both point and area source measurements.

Determining NH<sub>3</sub> emissions from AFOs requires continuous or semi-continuous measurements of NH<sub>3</sub> concentration with good sensitivity and wide dynamic measurable ranges. For area source and downwind fence line monitoring, the typical NH<sub>3</sub> concentration is lower than 1 ppm. FTIR, CL, PAS, and OPL techniques are appropriate due to their capability of detecting sub-ppb level NH<sub>3</sub>. Harper et al. (2010) used OPL to measure downwind NH<sub>3</sub> concentrations, combined with measured micrometeorological data, and to quantify emissions from broiler houses in the San Joaquin Valley of California with backward Lagrangian stochastic (bLS) model. For monitoring emissions from point sources (e.g., animal house air exhaust) with relatively high NH<sub>3</sub> concentrations,

---

Submitted for review in June 2014 as manuscript number 10826; approved as a Technical Note for publication by the Plant, Animal, & Facility Systems Community of ASABE in December 2015.

Mention of product or company names is for presentation clarity and does not imply endorsement by the authors or their affiliations, nor exclusion of other suitable products.

The authors are **Hong Li, ASABE Member**, Assistant Professor, Department of Animal and Food Sciences, University of Delaware, Newark, Delaware; **Chen Zhang**, Graduate Student, Department of Civil and Environmental Engineering, University of Delaware, Newark, Delaware; and **Hongwei Xin, ASABE Fellow**, Distinguished Professor, Iowa Egg Council Endowed Professor, Department of Agricultural and Biosystems Engineering, Iowa State University, Director of Egg Industry Center, Ames, Iowa. **Corresponding author:** Hong Li, University of Delaware, Newark, DE; phone: 302-831-1652; e-mail: hli@udel.edu.

PAS, CL, NIR, EC, and CC may be selected because of their wide detection range (0 to ~100 ppm or higher). It is challenging to find a sensor or analyzer that is suitable for poultry operations and can withstand harsh environmental conditions including high concentrations of corrosive gases (e.g.,  $\text{NH}_3$  and  $\text{H}_2\text{S}$ ), particulates and microorganisms, as well as large variability in temperature and relative humidity. PAS based multi-gas analyzers have been widely used for laying hen, broiler, and turkey emission studies over the past decade and demonstrates excellent detection limit, accuracy, repeatability, and response time (van der Peet-Schwering et al., 1999; Xin et al., 2009; Li and Xin, 2010; Li et al., 2011; Li et al., 2012; Lin et al., 2012a,b). PAS has been recognized by U.S. Environmental Protection Agency (EPA) and used by the National Air Emission Measurement Study (NAEMS) project for  $\text{NH}_3$  measurements in poultry, swine, and dairy operations (Ni and Heber, 2008). Our laboratory comparison study (unpublished data) had demonstrated the equivalency of the PSA multi-gas (including  $\text{NH}_3$ ) analyzer used in the current paper to an EPA-approved CL analyzer for measuring  $\text{NH}_3$  generated from poultry manure. Chillgard RT (Mine Safety Appliances Company, Pittsburgh, Pa.) is a single gas monitor and operates on the PAS principle, allowing a continuous measurement of  $\text{NH}_3$  concentration. This instrument has been used in several agricultural  $\text{NH}_3$  emissions studies in United States for its advantages over other  $\text{NH}_3$  analyzers, including stability, low cost, and minimal maintenance (Li et al., 2008a). The main drawbacks and concerns about the validity and quality of data obtained with Chillgard RT include its high detection limit (1 ppm) and susceptibility to potential interference by moisture content and other gaseous constituents in the sample air from poultry operations.

The objective of this study was to evaluate the operating performance of a single-gas Chillgard RT with regard to measurement uncertainty, repeatability, stability, and interference of moisture when measuring aerial  $\text{NH}_3$  from laying-hen and broiler facilities under laboratory and field conditions.

## MATERIALS AND METHODS

### PHOTOACOUSTIC GAS ANALYZER

Photoacoustic spectroscopy measures the effects of light absorption by solids, liquids, and gases by means of acoustic detection (Malkin and Cahen, 1979). For gas detection, it converts light absorbance (at wavelengths characteristic of an analyte) to acoustic signals, and in general delivers better sensitivity than conventional infrared (IR) gas spectrometry. Figure 1 illustrates the schematic of a photoacoustic gas analyzer. The IR light from a tungsten lamp is first modulated by a mechanical chopper and then passed through a narrow-band optical filter to remove all wavelengths except for the “measuring wavelength” characteristic of a target gas. In the measurement chamber, the target gas molecules become energized upon light absorption, and dissipate the absorbed energy in the form of heat. The chopper-modulated pulsed

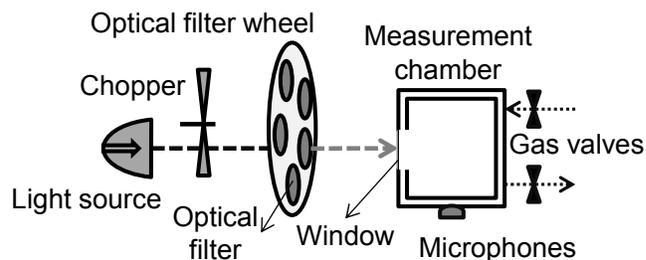


Figure 1. Schematic of a photoacoustic gas analyzer's operation.

light results in periodic heat expansion and contraction of the sample air, creating sound. The acoustic signal is recorded by microphones and converted to an electrical signal correlated to gas concentration.

The single-gas Chillgard RT analyzer was originally designed for monitoring  $\text{NH}_3$  concentrations typical of 1 to 1000 ppm with a sensitivity of 1 ppm and an accuracy of  $\pm 2$  ppm in refrigeration systems (Chillgard RT Manual, 2013). An internal vacuum pump draws air samples in at a flow rate varying from 0.75 to 1 LPM, depending on the size (e.g., length and diameter) of air sampling tubing and inlet. Its display reading updates every 7 s and can be recorded on a computer via analog input (0 to 10 VDC) or RS-232 serial connection.

An INNOVA 1412 multi-gas analyzer (LumaSense Technologies, Inc., Santa Clara, Calif.) served as a reference method and was compared with Chillgard RT under different laboratory and field conditions. Two INNOVA analyzers were equipped with five optical filters for measuring  $\text{NH}_3$ , carbon dioxide ( $\text{CO}_2$ ), nitrous oxide ( $\text{N}_2\text{O}$ ), methane ( $\text{CH}_4$ ), and water vapor, respectively. The sampling pump of the INNOVA drew air samples of up to 1.8 LPM to the measurement chamber where the samples were analyzed for different gases by different optical filters. The display readings of the INNOVA updated every 30 s and were stored in the internal memory. The data were exported to a computer via RS-232 or RS-485 connection. The instrument had an  $\text{NH}_3$  detection limit of 0.44 ppm with 1-s sampling and integration time (INNOVA gas detection limits, 2011).

The Chillgard RT and INNOVA analyzers were challenged weekly and calibrated, as needed, with zero, 23 (for laying hens), or 53 ppm (for broilers)  $\text{NH}_3$  (All zero or span gases were balanced with ultra-high-purity  $\text{N}_2$  unless otherwise noted), 3000 ppm  $\text{CO}_2$ , 5.02 ppm  $\text{N}_2\text{O}$ , and 105 ppm  $\text{CH}_4$  gas calibration standards (Certified plus or EPA grade, Matheson Tri-gas, Swedesboro, N.J.). The concentrations of the calibration standards were selected based on the actual gaseous concentrations in poultry operations and the availability of the standards.

### LABORATORY EVALUATION WITH STANDARD GASES

A laboratory evaluation system was built to assess the Chillgard RT for interference of  $\text{CH}_4$ ,  $\text{CO}_2$ ,  $\text{N}_2\text{O}$ , and water vapor on  $\text{NH}_3$  measurement (fig. 2). Mass flow controllers (GFC-17S, Aalborg, Orangeburg, N.Y.) were used to dilute gas calibration standards (53 ppm  $\text{NH}_3$ , 3000 ppm  $\text{CO}_2$ , 5.02 ppm  $\text{N}_2\text{O}$ , and 100 ppm  $\text{CH}_4$ ) to various challenge concentrations (table 1). A total of 12 concentration

**Table 1. Combinations of different gas concentrations for laboratory evaluation of Chillgard RT.**

ID	NH <sub>3</sub> (ppm)	CO <sub>2</sub> (ppm)	N <sub>2</sub> O (ppm)	CH <sub>4</sub> (ppm)	Dew point (°C)	Chillgard (ppm) <sup>[a]</sup>	INNOVA (ppm) <sup>[a]</sup>			
							NH <sub>3</sub>	CO <sub>2</sub>	N <sub>2</sub> O	CH <sub>4</sub>
1	53	0	0	0	-	53 (0)	52.8 (0.19)	4.61 (4.75)	0.03 (0.01)	0.63 (0.21)
2	0	3000	0	0	-	0 (0)	0.401 (0.22)	3035 (10.5)	0.07 (0.02)	1.84 (0.36)
3	0	0	5.02	0	-	0 (0)	0.264 (0.07)	8.96 (7.99)	5.04 (0.01)	1.19 (0.45)
4	0	0	0	100	-	0 (0)	0.342 (0.07)	8.04 (4.68)	0.03 (0.02)	101.6 (0.33)
5	0	0	0	0	19.8	0 (0)	0.206 (0.15)	8.28 (5.93)	0.03 (0.02)	0.71 (0.61)
6	26.5	1500	0	0	-	27 (0)	26.6 (0.15)	1518 (7.5)	0.06 (0.03)	2.26 (0.65)
7	26.5	0	2.5	0	-	27 (0)	26.4 (0.19)	7.33 (5.24)	2.50 (0.01)	0.92 (0.73)
8	26.5	0	0	50	-	27 (0)	26.8 (0.14)	4.79 (6.58)	0.02 (0.02)	50.38 (0.68)
9	26.5	0	0	0	13.5	27 (0)	26.5 (0.09)	0.05 (2.54)	0.05 (0.03)	-0.14 (3.31)
10	18	1000	0	0	11.1	18 (0)	17.7 (0.15)	1013 (4.0)	0.08 (0.04)	3.03 (1.49)
11	18	0	1.7	0	11.1	18 (0)	17.4 (0.27)	-4.33 (4.54)	1.71 (0.01)	-1.74 (2.03)
12	18	0	0	34	11.1	18 (0)	17.6 (0.14)	1.15 (6.98)	0.25 (0.03)	35.7 (0.55)

<sup>[a]</sup> Chillgard and INNOVA concentrations: mean (standard deviation).

combinations were tested, at an environmental temperature of 20°C and a total gas flow rate of 3 LPM. Calibration gases were diluted and mixed in a polytetrafluoroethylene (PTFE) manifold (mixing manifold) before proceeding to a second manifold (sampling manifold) where gases were sampled for analysis. Each concentration combination was tested for 5 min and the NH<sub>3</sub> readings of both Chillgard RT and INNOVA at the end of the 5-min period were compared. PTFE tubing's (4 mm o.d. × 3 mm i.d. for INNOVA and 6.35 mm o.d. × 3.18 mm i.d. for Chillgard RT) were used and the tubing length between the sampling manifold and each analyzer was 1 m.

**LABORATORY EVALUATION SYSTEM: USING BROILER LITTER**

One Chillgard RT and one INNOVA were selected to monitor NH<sub>3</sub> gas concentrations from four 19-L plastic vessels containing broiler litter (fig. 3). Both air inlet and outlet were located on the air-tight lid of the vessel. Samples of the exhaust air from each vessel were sequentially taken using an air sampling pump (Model BTC-IIS, Parker Hannifin, Hollis, N.H.) at 5-min intervals. This sampling sequence yielded a measurement cycle of 25 min for the entire system (including 5 min for the ambient air). The successive sampling was accomplished through controlled operation of four solenoid valves (Model 456654, Burkert, Irvine, Calif.). An inline Teflon-membrane filter (4.7 cm dia., 5 µm pore dia.) was mounted upstream of each solenoid valve. The vessels were operated under negative pressure and were placed in an environment-controlled 20°C room. Analog outputs from the Chillgard RT and digital outputs from the INNOVA were

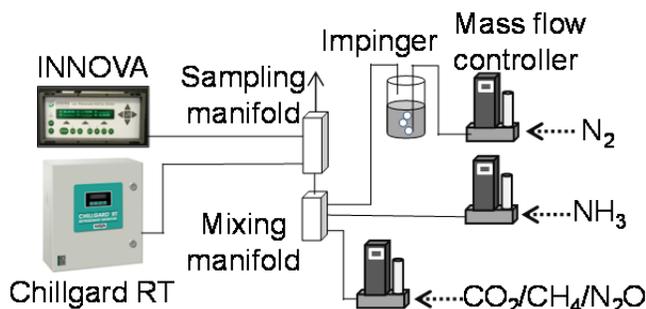


Figure 2. Schematic of laboratory system setup for gaseous interference evaluation.

logged on a computer at 1-s intervals using a data acquisition (DAQ) module (Model USB-2416, Measurement Computing Corporation, Norton, Mass.) and a LabVIEW (National Instrument Corp., Austin, Tex.) program. The program also averaged all the raw measurement data over 60-s intervals and recorded them in a data file. Fluorinated ethylene propylene (FEP; 6.35 mm o.d. × 3.97 mm i.d.) tubing was used except for the tubing (PTFE) between the sampling manifold and gas analyzers.

**FIELD EVALUATION: LAYING HENS**

One Chillgard RT and One INNOVA were installed in a mobile air emissions monitoring unit (MAEMU) on site to continuously collect data on NH<sub>3</sub>, CO<sub>2</sub>, CH<sub>4</sub>, and N<sub>2</sub>O concentrations and emissions from three laying-hen houses on a commercial farm in Iowa. The three identical laying-hen houses had high-rise style, with manure storage in the lower level and no supplemental heating. Air samples were drawn from two composite sampling points in the manure storage of each house, as well as from an air inlet located on the ceiling of one of the houses to provide the ambient background data. Each sampling point was equipped with dust filters to keep large particulates from plugging or contaminating sample line, servo valves and delicate gas analyzers. A positive-pressure gas sampling system (PP-GSS) was used in the MAEMU and continuously pumped air samples from each sampling point using a dedicated air pump (fig. 4). The sample air was bypassed when not analyzed. Air samples from each location were collected sequentially over 2-min period via the controlled operation

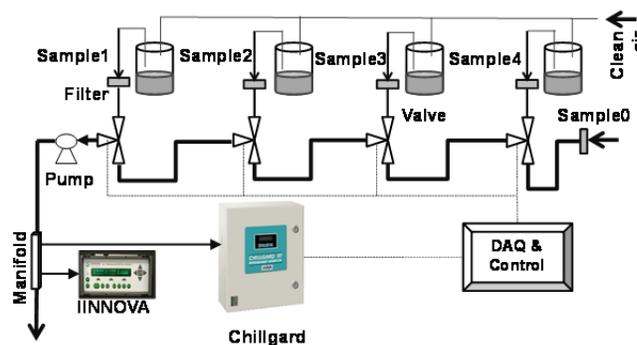


Figure 3. Schematic of the laboratory system setup for the Chillgard RT evaluation. The system measured NH<sub>3</sub> concentrations of air samples from four broiler litter vessels.

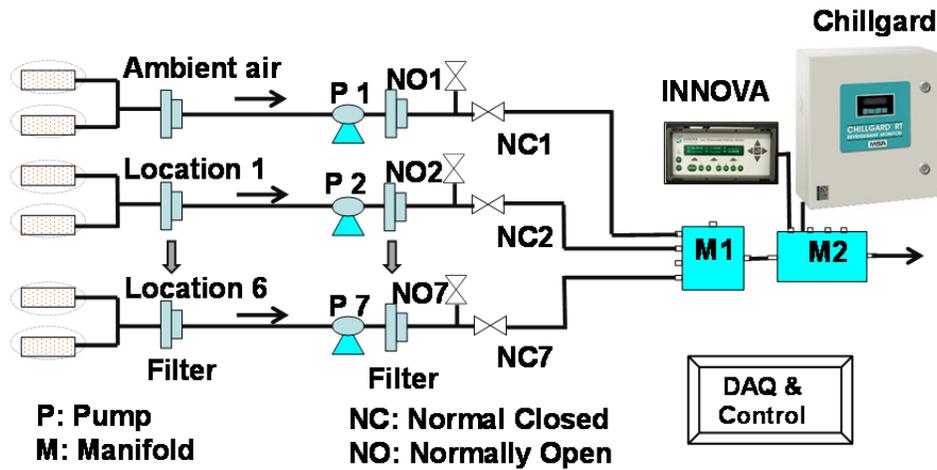


Figure 4. Schematic of the field system setup on a laying-hen farm for the Chillgard RT evaluation. The system measured  $\text{NH}_3$  concentration at six composite sampling points and an air inlet in three high-rise laying-hen houses.

of the servo valves of the PP-GSS. Every 2 hours, air samples from the ambient (background) location were drawn and analyzed for 8 min. FEP (9.53 mm o.d.  $\times$  6.35 mm i.d.) tubing was used except for the tubing (PTFE) between the sampling manifold and gas analyzers.

#### FIELD EVALUATION: BROILERS

Two identical broiler houses on a commercial farm in Delaware were monitored for air emissions with the INNOVA and Chillgard RT. Built-up litter was used and only top-caked litter was removed after each 42-day grow-out during this monitoring study. Liquid propane radiant heaters were used during brooding periods and cold seasons when housing temperature dropped below setting temperatures. Air sampling equipment, gas analyzers, and data acquisition and control system were housed in a temperature-controlled cabinet in one of the houses' control room. Air samples were drawn from two sampling points in each broiler house. A negative-pressure gas sampling system (NP-GSS) was used and it collected air samples

from each sampling point using two pumps, one for gas analyzers and the other for bypass air (fig. 5). Air samples from each location were collected sequentially over 3-min period via the controlled operation of the servo valves of the NP-GSS. FEP (6.35 mm o.d.  $\times$  3.97 mm i.d.) tubing was used except for the tubing (PTFE) between the sampling manifold and each analyzer.

A compact Fieldpoint device (CFP-2020, National Instrument Corp., Austin, Tex.) programmed with LabVIEW was used for data acquisition and system control in both laying hen and broiler field evaluations. Temperatures in animal houses, MAEMU, and equipment cabinets were monitored, along with the dew point of air samples. Heating tapes and thermostats were used to heat the air sampling tubing to  $>30^\circ\text{C}$ , preventing potential moisture condensation. The analog outputs of the Chillgard RT and INNOVA were recorded at 1-s intervals and then averaged over 60-s intervals.

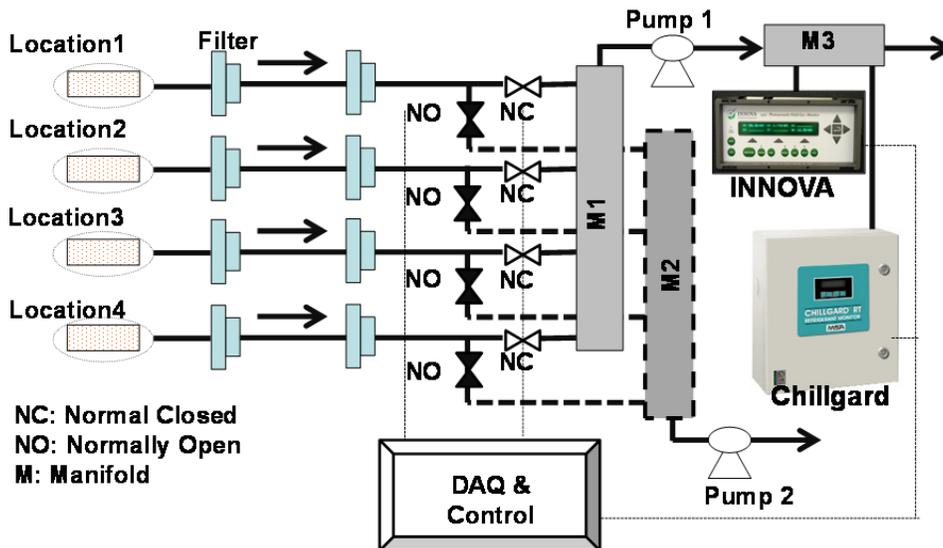


Figure 5. Schematic of the field system setup on a broiler farm for the Chillgard RT evaluation. The system measured  $\text{NH}_3$  concentration at four air sampling points in two broiler houses.

## DATA ANALYSIS

Analog outputs of the Chillgard RT were converted to  $\text{NH}_3$  concentrations to match the display readings. Statistical analyses of the Chillgard RT and INNOVA measurement results were performed with analysis of variance (ANOVA) using standard least squares of JMP (SAS Institute, Cary, N.C.). Significant differences for all comparisons were based on  $P < 0.05$ .

## RESULTS AND DISCUSSION

### CROSS-INTERFERENCE OF $\text{CH}_4$ , $\text{CO}_2$ , $\text{N}_2\text{O}$ , AND WATER VAPOR

The cross-interferences of  $\text{CH}_4$ ,  $\text{CO}_2$ ,  $\text{N}_2\text{O}$ , and water vapor on  $\text{NH}_3$  measurement are well recognized by the INNOVA manufacturer. An algorithm, embedded in the instrument's internal memory, automatically corrects  $\text{NH}_3$  readings based on the concentrations of other gaseous constituents. Comparatively, no such correction is available to the Chillgard RT; and it is stated in the manual that Chillgard RT is cross-sensitive to  $\text{CH}_4$ . However, our experiments showed that, within the test concentration ranges ( $\text{NH}_3$ : 0 to 50 ppm;  $\text{CO}_2$ : 0 to 3000 ppm;  $\text{N}_2\text{O}$ : 0 to 5 ppm,  $\text{CH}_4$ : 0 to 100 ppm), none of the four gases had a significant cross-interference on  $\text{NH}_3$  measurement ( $P > 0.14$ ). The drift of  $\text{NH}_3$  readings caused by  $\text{CH}_4$  was estimated to be 1 ppm  $\text{NH}_3$  per 5,000 ppm  $\text{CH}_4$ . In poultry houses,  $\text{CH}_4$  concentrations are well below this level. Thus, no cross-interference of  $\text{CH}_4$  was noted given the resolution of  $\text{NH}_3$  measurement (1 ppm) by the Chillgard RT.

### RESPONSE TIMES OF CHILLGARD RT AND INNOVA

Air samples from broiler-litter vessels and ambient air were sequentially measured for 5 min each and the last reading during the 5-min analysis was used as the final stabilized reading. The 30-s instantaneous readings by the Chillgard RT and INNOVA over a 45-min period are plotted to illustrate the time for them to respond to step changes in concentration (response time) (fig. 6a). The dynamic concentrations at 30-s intervals were compared to the final stabilized reading and the percentage responses were calculated and presented in figure 6b. For all tested  $\text{NH}_3$  concentrations, Chillgard RT showed a quicker response than INNOVA. It took 120 s for the Chillgard RT to reach a  $>95\%$  expected value when samples were switched from ambient air to  $\text{NH}_3$ -laden airs; whereas it the INNOVA 180 s. Flushing both instruments with  $\text{NH}_3$ -laden air prior to the measurement, the average 95% response time (T95) of the Chillgard and INNOVA decreased to 90 and 120 s, respectively. Water vapour could affect the response time of the INNOVA as its T95 decreased with the increasing dew-point temperature of air samples ( $P < 0.01$ ). Upon a sudden change in  $\text{NH}_3$  concentration, the Chillgard RT and INNOVA reached an average of 99.6% and 96.7% full responses, respectively, after 120 s (fig. 7). The INNOVA's response time can be shortened by reducing the numbers of optical filters used. For instance, for single-gas measurement, the sampling interval of

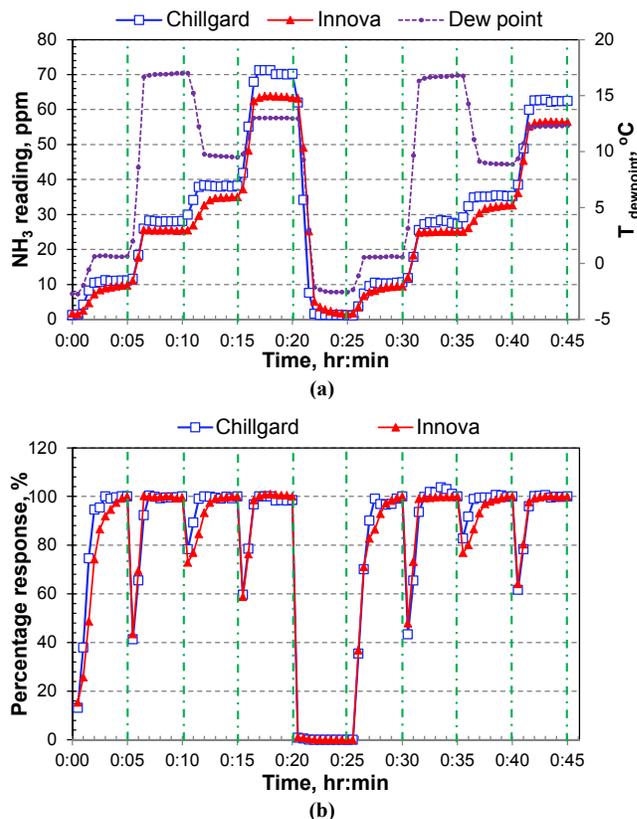


Figure 6. Response times of the Chillgard RT and INNOVA for air samples with different ammonia concentrations (a: dynamic ammonia concentration; b: percentage of dynamic concentration to stabilized concentration during each 5-min sampling). Vertical dash lines indicate changes in air samples.

INNOVA (between two readings) could be set to 20 s and the response of INNOVA at 120 s would increase to 98.6%.

### CHILLGARD RT VS. INNOVA WITH BROILER LITTER

The Chillgard RT measured higher  $\text{NH}_3$  concentrations than the INNOVA when monitoring  $\text{NH}_3$  gas emissions from broiler litter (fig 6a). To further examine such differences, we compared the  $\text{NH}_3$  readings of the Chillgard RT and INNOVA at multiple response times,

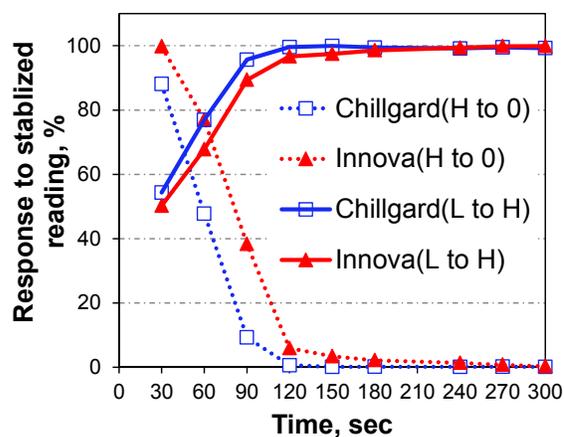


Figure 7. Average response times of the Chillgard RT and INNOVA. (H to 0: concentration change from high to zero; L to H: concentration change from low to high).

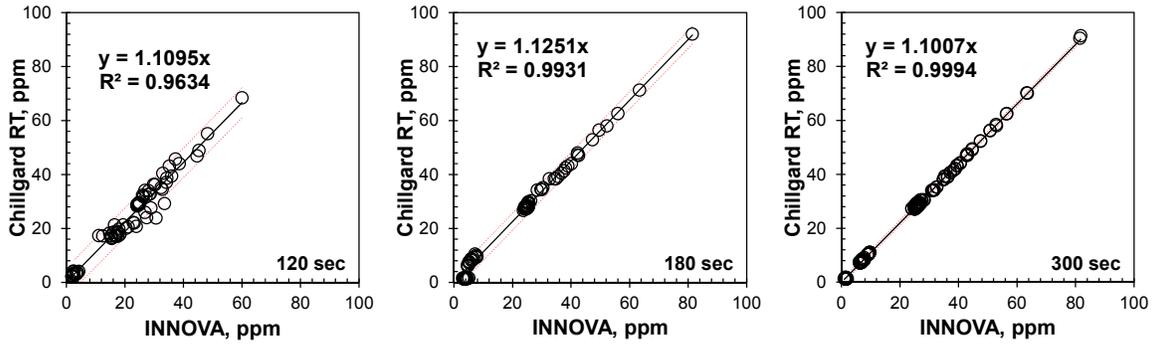


Figure 8. Relationship of ammonia readings with the Chillgard RT vs. INNOVA at 120, 180, and 300 s. The dashed lines below and above the regression lines represent 95% confidence intervals of the observations.

including 120, 180, and 300 s (fig. 8). All paired comparisons yielded an excellent linearity ( $R^2 > 0.96$ ). The  $\text{NH}_3$  readings given by the Chillgard RT were 11.0%, 12.5%, and 10.0% higher than those derived from the INNOVA at 120-, 180-, and 300-s response time, respectively. The P-values of the paired t-tests were all  $< 0.0001$ , indicating significant differences. The difference may have resulted from the co-existence of other gas constituents (e.g., hydrocarbons) with  $\text{NH}_3$  in the air sample, which might share similar light-adsorption wavelengths to  $\text{NH}_3$ . Trabue et al. (2010) reported the top 25 speciated non-methane hydrocarbons (NMHCs) and their concentration levels in a commercial broiler house. The NMHC species and concentrations differed with and without birds inside the house. Their concentrations were related to litter conditions and ventilation rate, and showed a similar time-series trend to  $\text{NH}_3$  concentration (Burns et al., 2007; Li et al., 2008b). The cross-sensitivity of the Chillgard RT and INNOVA to most NMHC compounds was specified in the manual by the manufacturer except for *o*-xylene, propane, and pentane. On the other hand, the interference of NHMCs on  $\text{NH}_3$  measurement (i.e.,  $\text{NH}_3$  detection selectivity) is affected by the band (e.g., width of transmitting wavelengths) of the optical filter used. Different filters employed by the Chillgard RT and INNOVA, thus, would result in their different susceptibility to the interference by NHMCs and accordingly different  $\text{NH}_3$  readings.

#### CHILLGARD RT VS. INNOVA IN LAYING-HEN AND BROILER HOUSES

Figure 9a compares  $\text{NH}_3$  concentrations measured by the Chillgard RT and INNOVA in the high-rise laying-hen manure storage, with readings acquired at the end of every 2-min sampling period. Laying hen  $\text{NH}_3$  concentrations derived from the Chillgard RT were 3.2% higher than those from the INNOVA; however, in the broiler houses, the Chillgard RT readings were 22.9% higher (fig. 11b). The gas composition of the exhaust air from laying-hen and broiler houses could be significantly different. For example, liquid propane is typically burned for supplemental heating in broiler houses during winter while no supplemental heating is provided in laying-hen houses. The compositions of laying-hen and broiler feces can be different due to different feed formulation and nutrient content, which may result in different volatile organic compounds (in both composition and concentrations) when the feces is degraded by microorganisms. The results of this study suggest that the Chillgard RT should be evaluated by a reference method, such as chemiluminescence, FTIR, or INNOVA for cross-interferences from uncharacterized volatile organic compounds. The Chillgard RT may be good for mitigation studies that focus on relative changes in  $\text{NH}_3$  concentration. But for emission monitoring (e.g., emission inventory development), the measurement bias by the Chillgard RT should be fully checked and be properly corrected prior to use.

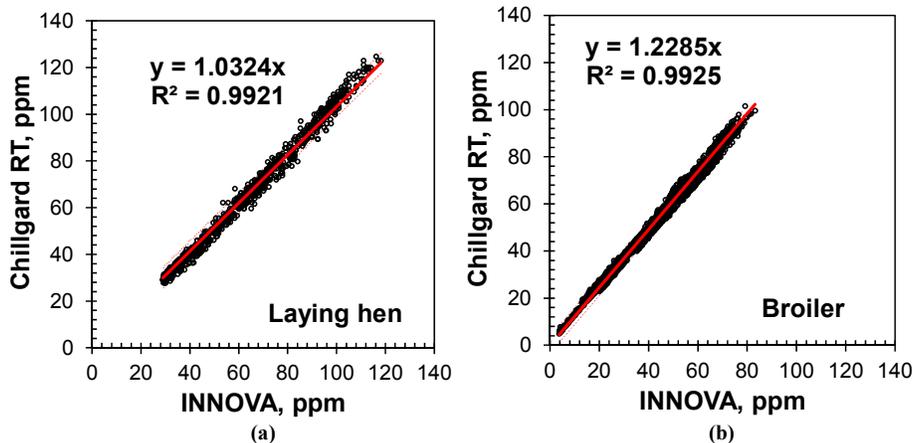


Figure 9. Comparison of ammonia readings measured by the Chillgard RT vs. INNOVA during two field monitoring events (a: laying hens with 1395 points; b: broilers with 2602 points).

## CONCLUSIONS

A commercially available single photoacoustic analyzer, Chillgard RT, was evaluated and compared with a multi-gas photoacoustic analyzer, INNOVA 1412, for real-time continuous monitoring of NH<sub>3</sub> in poultry operations. The following conclusions can be drawn:

1. The NH<sub>3</sub> readings by the Chillgard RT were not significantly affected by CO<sub>2</sub>, N<sub>2</sub>O, CH<sub>4</sub>, or water vapor under typical operational conditions in poultry houses.
2. The Chillgard RT has a quicker response than the INNOVA. The Chillgard RT and INNOVA achieved an average of 99.6% and 96.7% expected values, respectively, after 120 s measurement when the INNOVA was configured at a 30-s sampling interval.
3. There was a strong linear relationship between Chillgard RT and INNOVA under all the experimental conditions. The Chillgard RT overestimated NH<sub>3</sub> concentrations by an average of 3.24%, 10%, and 22.9% for measurements with the laying-hen houses, stored boiler litter, and broiler houses, respectively.
4. Performance of the Chillgard RT should be evaluated using certain reference NH<sub>3</sub> analyzers, such as chemiluminescence, FTIR, and INNOVA before used for monitoring concentrations and emissions in a new environment.

## ACKNOWLEDGMENTS

Financial support of the study was provided in part by the USDA NRCS Conservation Innovation Grant Program (Awards NRCS 69-3A75-12-244 and 69-3A75-7-91), an Agriculture and Food Research Initiative competitive grant (2013-67021-21128) from the USDA National Institute of Food and Agriculture, U.S. Poultry & Egg Association, and University of Delaware. We thank the producers for allowing access to their flocks.

## REFERENCES

- Asman, W. A. H., Cellier, P., Genermont, S., Hutchins, N. J., & Sommer, S. G. (1998). Ammonia emission research: From emission factors to process descriptions. *EUROTRAC Newsletter*, 20, 2-10.
- Burns, R. T., Xin, H., Gates, R. S., Li, H., Overhults, D. G., Moody, L., & Earnest, J. W. (2007). Ammonia emissions from broiler houses in the Southeastern United States. *Proc. Intl. Symp. Air Quality and Waste Management for Agriculture*. St. Joseph, Mich.: ASABE.
- Harper, L. A., Flesch, T. K., & Wilson, J. D. (2010). Ammonia emissions from broiler production in the San Joaquin Valley. *Poultry Sci.*, 89(9), 1802-1814. <http://dx.doi.org/10.3382/ps.2010-00718>.
- INNOVA gas detection limits. (2011). LumaSense Technologies, Inc. Retrieved from [http://www.lumasenseinc.com/uploads/Solutions/pdf/Technical\\_Literature/English/Lumasense-gas-detection-limits\\_Wall-Chart.pdf](http://www.lumasenseinc.com/uploads/Solutions/pdf/Technical_Literature/English/Lumasense-gas-detection-limits_Wall-Chart.pdf).
- Li, H., & Xin, H. (2010). Lab-scale assessment of gaseous emissions from laying-hen manure storage as affected by physical and environmental factors. *Trans. ASABE*, 53(2), 593-604. Retrieved from <http://dx.doi.org/10.13031/2013.29574>.
- Li, H., Xin, H., Burns, R. T., & Liang, Y. (2008a). Reduction of ammonia emission from stored poultry manure using additives: Zeolite, Al+Clear, Ferix-3 and PLT. *J. Appl. Poultry Res.*, 17, 421-431. <http://dx.doi.org/10.3382/japr.2007-00076>.
- Li, H., Burns, R. T., Xin, H., Gates, R. S., Trabue, S., Overhults, D. G., Moody, L., & Earnest, J. W. (2008b). Hydrogen sulfide and nonmethane hydrocarbon emissions from broiler houses in the southeastern United States. ASABE Paper No. 084417. St. Joseph, Mich.: ASABE.
- Li, H., Xin, H., Burns, R. T., Jacobson, L. D., Noll, S., Hoff, S. J., Harmon, J. D., Koziel, A., & Hetchler, B. P. (2011). Air emissions from tom and hen turkey houses in the U.S. Midwest. *Trans. ASABE*, 54(1), 305-314. <http://dx.doi.org/10.13031/2013.36258>.
- Li, H., Xin, H., Burns, R. T., Roberts, S. A., Li, S., Kliebenstein, J., & Bregendahl, K. (2012). Reducing ammonia emissions from laying-hen houses through dietary manipulation. *J. Air & Waste Mgmt. Assoc.*, 62(2), 160-169. <http://dx.doi.org/10.1080/10473289.2011.638414>.
- Liang, Y., Xin, H., Hoff, S. J., Richard, T. L., & Kerr, B. J. (2004). Performance of single point monitor in measuring ammonia and hydrogen sulfide gases. *Appl. Eng. Agric.*, 20(6), 863-872. <http://dx.doi.org/10.13031/2013.17712>.
- Lin, X. -J., Cortus, E. L., Zhang, R., Jiang, S., & Heber, A. J. (2012a). Ammonia, hydrogen sulfide, carbon dioxide and particulate matter emissions from California high-rise layer houses. *Atmospheric Environ.*, 46, 81-91. <http://dx.doi.org/10.1016/j.atmosenv.2011.10.021>.
- Lin, X.-J., Cortus, E. L., Zhang, R., Jiang, S., & Heber, A. J. (2012b). Air emission from broiler houses in California. *Trans. ASABE*, 55(5), 1895-1908. <http://dx.doi.org/10.13031/2013.42377>.
- Malkin, S., & Cahen, D. (1979). Photoacoustic spectroscopy and radiant energy conversion: Theory of the effect with special emphasis on photosynthesis. *Photochem. Photobiol.*, 29, 803-813.
- Chillgard RT Manual. (2013). Mine safety appliances company. Retrieved from <http://s7d9.scene7.com/is/content/minesafetyappliances/Chillgard%20RT%20Refrigerant%20Monitor%20Instruction%20Manual%20-%20EN>.
- Ni, J., & Heber, A. J. (2008). Sampling and measurement of ammonia at animal facilities. *Adv. Agron.*, 98, 201-269. [http://dx.doi.org/10.1016/S0065-2113\(08\)00204-6](http://dx.doi.org/10.1016/S0065-2113(08)00204-6).
- Trabue, S. L., Scoggin, K. D., Li, H., Burns, R. T., Xin, H., & Hatfield, J. L. (2010). Speciation of volatile organic compounds from poultry production. *Atmospheric Environ.*, 44(29), 3538-3546. <http://dx.doi.org/10.1016/j.atmosenv.2010.06.009>.
- van der Peet-Schwering, C. M. C., Aarnink, A. J. A., Rom, H. B., & Dourmad, J. Y. (1999). Ammonia emissions from pig houses in The Netherlands, Denmark and France. *Livestock Prod. Sci.*, 58, 265-269. [http://dx.doi.org/10.1016/S0301-6226\(99\)00017-2](http://dx.doi.org/10.1016/S0301-6226(99)00017-2).
- Xin, H., Li, H., Burns, R. T., Gates, R. S., Overhults, D. G., & Earnest, J. W. (2009). Use of CO<sub>2</sub> concentration difference or CO<sub>2</sub> balance to assess ventilation rate of broiler houses. *Trans. ASABE*, 52(4), 1353-1361. <http://dx.doi.org/10.13031/2013.27787>.

**Author**

First Name or initial	Middle Name or initial	Surname	Role	E-mail (and phone for contact author)	Contact author?
Hong		Li	ASABE Member	hli@udel.edu	Yes

**Affiliation**

Organization	Address	Country	URL
University of Delaware	046 Townsend Hall, Newark, Delaware, 19716	USA	

**Author**

First Name or initial	Middle Name or initial	Surname	Role	E-mail (and phone for contact author)	Contact author?
Hongwei		Xin	ASABE Fellow, Distinguished Professor	hxin@iastate.edu	No

**Affiliation**

Organization	Address	Country	URL
Iowa State University	3204 NSRIC, Ames, Iowa, 50011-3310	USA	

**Author**

First Name or initial	Middle Name or initial	Surname	Role	E-mail (and phone for contact author)	Contact author?
Chen		Zhang	Non-member	czh@udel.edu	No

**Affiliation**

Organization	Address	Country	URL
University of Delaware	111 Worrilow Hall, Newark, DE 19716	USA	

**Information about the Journal where this article will be published**

Full Title of the Journal	DOI example only 10.13031/aea.29.10149
Applied Engineering in Agriculture	

**Pub Info**

Copyright or not	Year	Volume, Issue	Manuscript ID
yes	2015	31(1)	PAFS10826