Behavioral Response of Weaned Pigs during Gas Euthanasia with CO2, CO2 with Butorphanol, or Nitrous Oxide

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Behavioral Response of Weaned Pigs during Gas Euthanasia with CO2, CO2 with Butorphanol, or Nitrous Oxide

Abstract
The swine industry is often forced to euthanize pigs in the first few weeks of life due to injuries, hernias, or unthriftiness. The majority of pigs are euthanized using carbon dioxide (CO2) gas asphyxiation but concerns as to the humaneness of CO2 are increasing. This study compared the euthanasia of weaned pigs using N2O (N2O; n = 9) or CO2 (n = 9), at 50% and 25% min\(^{-1}\) exchange rate, respectively. In addition, we administered an analgesic prior to euthanasia with CO2 (CO2B) exposure as a third treatment (n = 9) to elucidate behaviors indicative of pain. Pigs in the CO2 and N2O treatments lost posture at similar times (latency of 145.0 ± 17.3 and 162.6 ± 7.0 s respectively, \(p > 0.10\)), while the CO2B treatment pigs lost posture the soonest (101.2 ± 4.7 s, \(p < 0.01\)). The pigs in the CO2B treatment made more escape attempts than the CO2 or N2O pigs (16.4 ± 4.2, 4.7 ± 1.6, 0.3 ± 0.2, respectively; \(p < 0.0004\)). However, pigs in N2O squealed more often than either the CO2 or CO2B pigs (9.0 ± 1.6, 2.8 ± 1.2, 1.3 ± 0.6, respectively, \(p < 0.001\)). Given the similar time to loss of posture and shorter time displaying open mouth breathing, N2O may cause less stress to pigs; however, the greater number of squeals performed by these pigs suggests the opposite. It was not apparent that any behavior measured was indicative of pain. In conclusion, N2O applied at a 50% min\(^{-1}\) flow rate can be an alternative to CO2 for pig euthanasia.

Keywords
gas flow rate, sus scrofa, swine, euthanasia, welfare

Disciplines
Agriculture | Bioresource and Agricultural Engineering

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Behavioral Response of Weaned Pigs during Gas Euthanasia with CO₂, CO₂ with Butorphanol, or Nitrous Oxide

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Simple Summary: Pig farmers are forced to euthanize a significant number of pigs due to injuries, hernias, or unthriftiness. The majority of pigs are euthanized using carbon dioxide gas asphyxiation. However, the humaneness of carbon dioxide is being increasingly questioned. An alternative is the use of nitrous oxide gas. We conducted this study to compare the euthanasia of young pigs using nitrous oxide or carbon dioxide. In addition, we tested the administration of a pain relief drug prior to carbon dioxide exposure to determine if we could eliminate behaviors indicative of pain. Pigs became unable to control their muscle movement, breathed heavily, and lost posture at the same time regardless of treatment. Pigs exposed to both gases showed heavy breathing and open-mouth breathing prior to losing posture. However, pigs exposed to carbon dioxide made more escape attempts but fewer squeals than pigs exposed to nitrous oxide. Administration of pain relief prior to exposure to carbon dioxide did not alter behaviors indicative of pain. The findings are inconclusive as to whether using nitrous oxide is significantly better than using carbon dioxide, but the results show that its use is just as effective, and possibly more humane.

Abstract: The swine industry is often forced to euthanize pigs in the first few weeks of life due to injuries, hernias, or unthriftiness. The majority of pigs are euthanized using carbon dioxide (CO₂) gas asphyxiation but concerns as to the humaneness of CO₂ are increasing. This study compared the euthanasia of weaned pigs using N₂O (N₂O; n = 9) or CO₂ (n = 9), at 50% and 25% min⁻¹ exchange rate, respectively. In addition, we administered an analgesic prior to euthanasia with CO₂ (CO₂B) exposure as a third treatment (n = 9) to elucidate behaviors indicative of pain. Pigs in the CO₂ and N₂O treatments lost posture at similar times (latency of 145.0 ± 17.3 and 162.6 ± 7.0 s respectively, p > 0.10), while the CO₂B treatment pigs lost posture the soonest (101.2 ± 4.7 s, p < 0.01). The pigs in the CO₂B treatment made more escape attempts than the CO₂ or N₂O pigs (16.4 ± 4.2, 4.7 ± 1.6, 0.3 ± 0.2, respectively; p < 0.0004). However, pigs in N₂O squealed more often than either the CO₂ or CO₂B pigs (9.0 ± 1.6, 2.8 ± 1.2, 1.3 ± 0.6, respectively, p < 0.001). Given the similar time to loss of posture and shorter time displaying open mouth breathing, N₂O may cause less stress to pigs; however, the greater number of squeals performed by these pigs suggests the opposite. It was not apparent that any behavior measured was indicative of pain. In conclusion, N₂O applied at a 50% min⁻¹ flow rate can be an alternative to CO₂ for pig euthanasia.

Keywords: gas flow rate; sus scrofa; swine; euthanasia; welfare
1. Introduction

Pork is the most consumed livestock meat in the world, with 1.4 billion pigs slaughtered each year [1]. Unfortunately, each year approximately 7.65 million newborn pigs in the US need to be euthanized due to various problems, including injury and starvation (calculated from production and loss data [2]). In addition, a significant number of neonatal pigs have to be euthanized prior to weaning due to hernias, lameness, and lack of thriftiness. Thus, ensuring a humane death for these animals represents a significant opportunity to safeguard animal welfare.

Any method of euthanasia should minimize pain and distress, induce rapid loss of consciousness, and a quick death [3,4]. The most common methods of euthanasia are blunt force trauma for neonatal pigs and CO$_2$ for neonates and older pigs. There are aesthetic concerns with blunt force trauma, and if done incorrectly, the pig can suffer. Carbon dioxide is widely used in the swine industry with automated gas chambers designed for on-farm use, typically using a fill flow rate of 25% of the chamber volume per minute (half-life 2.46). However, CO$_2$ is aversive to pigs [5,6] and is a welfare concern [7]. Research in our laboratory [6] found that pigs exposed to CO$_2$ squealed and flailed when concentrations of CO$_2$ reached about 20%, which was interpreted as an aversive response. The recommendation for the use of CO$_2$ is largely based on the speed at which it induces unconsciousness, as well as being economically affordable, widely available, and relatively safe to use for operators, despite being aversive to pigs. Therefore, the use of CO$_2$ remains a significant welfare concern for producers, scientists, and the public.

Nitrous oxide (N$_2$O) is one of the most common agents used in anesthetic practice for humans [8]. It is commonly used in human dental practices and referred to as ‘laughing gas’ due to its analgesic, sedative, and anxiolytic properties [9]. In mice, N$_2$O mixed with CO$_2$ decreased the time to loss of consciousness compared to CO$_2$ alone [10]. In pigs, Rault et al. [11] showed that N$_2$O was less aversive than CO$_2$ and was capable of inducing anesthesia [6]. In our previous research [12] to determine if a two-step process of using N$_2$O followed by CO$_2$ would be more effective than N$_2$O alone, one group of pigs was exposed to N$_2$O at a flow rate of 25% replacement [3] for 6 min before being exposed to CO$_2$ and the other group was directly exposed to CO$_2$. All pigs in the N$_2$O treatment lost posture (a sign of the onset of loss of consciousness) prior to entering the CO$_2$. However, because the pigs in the N$_2$O group were able to spend more time upright, they also had more time to squeal and attempt to escape; therefore, it could not be concluded whether the two-step method was more humane than CO$_2$. The authors speculated that increasing the flow rate for N$_2$O would cause pigs to lose posture sooner and those behaviors potentially indicative of aversiveness would be reduced.

It is generally believed that behaviors, such as heavy breathing, open-mouth breathing, squeals, and escape attempts, are signs of varying degrees of distress [6,13,14]. Importantly, it would be helpful to determine if these behaviors are also indicative of pain. In order to value one method of euthanasia over another, both pain and distress need to be assessed and alleviated. For instance, pigs exposed to CO$_2$ lose posture quicker than those exposed to N$_2$O, using similar flow rates for each gas [11]. However, if the pigs that were exposed to CO$_2$ experience pain and distress before losing posture and those exposed to N$_2$O do not experience pain or distress, then N$_2$O would be considered a more humane method.

Therefore, we sought to determine if increasing the flow rate for N$_2$O would decrease the time to loss of posture to be equivalent to CO$_2$. Further, we conducted the following study to determine if behaviors indicative of pain could be elucidated by using an analgesic prior to euthanizing pigs with CO$_2$. We hypothesized that: (1) A greater flow rate of N$_2$O would decrease the latency to loss of posture and therefore also reduce squeals and escape attempts, and (2) pigs administered butorphanol and exposed to CO$_2$ would squeal less and perform fewer escape attempts. Because both treatments deprive the pig of oxygen, we expected that heavy breathing, open-mouth breathing, and gaping to occur in both treatments.
2. Materials and Methods

2.1. Animals and Housing

All research was approved by the Purdue University Animal Care and Use Committee (FA1801001687). The pigs were the progeny of a commercial crossbred line born to sows from a local producer. The research was conducted at the local farm. Weaning aged pigs, approximately 21 days of age, which were destined to be euthanized, were used for the project. This farm produced weaned pigs for sale; thus, any pig that was deemed not to meet the quality standards for sale had to be euthanized. Reasons for euthanasia included: Injuries, prolapse, hernia, lame, and lightweight. The majority (72%) were males who exhibited scrotal hernias. For each treatment, 2 pigs were euthanized together in 9 repetitions; thus, 54 pigs in total were euthanized (2 pigs \( \times 9 \) repetitions \( \times 3 \) treatments).

The euthanasia gas chamber was the same as that used in our previous research [12]. Briefly, the chamber was 61 \( \times \) 38 \( \times \) 46 cm (Euthanex\textsuperscript{\textregistered} Ag Pro\textsuperscript{TM}, NutriQuest Inc., Mason City, IA, USA) that was modified with acrylic viewing windows to the front and back sides. Battery-operated lights were secured to the top interior of the box to provide more visibility for video recording. Gas was delivered to one side of the box after passing through a mass flow controller (GFC47, Aalborg Instruments & Controls, Inc., Orangeburg, NY, USA). Gas exited the tank through ducting connected to an exhaust fan (FR100, Fantech, Lenexa, KS, USA), which kept the chamber under negative pressure as confirmed via a manometer and served to flush the chamber between treatments. Gas was delivered at a 25% replacement rate per min for the CO\(_2\) treatments, as recommended per the American Veterinary Medical Association (AVMA) [3], and at a 50% replacement rate for the N\(_2\)O treatment. The effect on the O\(_2\) concentration at these replacement rates meant that O\(_2\) was depleted by half (from 20% to 10%) after 2.77 and 1.39 min for the 25% and 50% replacement rates, respectively.

2.2. Procedures

Pigs were subjected to 1 of 3 euthanasia treatments: CO\(_2\) after receiving 0.085 mL saline i.m. (CO\(_2\)), CO\(_2\) after receiving a dose calculated at 0.2 mg/kg BW for a 4.25 kg piglet, i.m. (0.085 mL dose) of butorphanol (CO\(_2\)-B), or N\(_2\)O (N\(_2\)O, no injection received). Butorphanol is an analgesic commonly used in veterinary medicine. Butorphanol is a synthetic partial agonist-antagonist analgesic, acting on kappa- and mu-opioid receptors [15], with a potency 7 times greater than morphine. It reaches its maximum analgesic effect within 20 to 30 min. The treatment with butorphanol was included to determine if specific aversive behaviors (‘pain behaviors’) could be verified that have been reported in pigs when they are euthanized with CO\(_2\). Many of the behaviors reported as aversive/painful are confounded with the body’s natural response to obtain air. In contrast, a fourth treatment using butorphanol was not included for the N\(_2\)O treatment because no such aversion has been reported in humans when it is used in the dental industry and pigs also do not find it aversive [6].

Farm staff collected the pigs directly from the sow and delivered them in carts in groups of 6 to 8, until a new group was needed. The pigs were kept in a cart in a group and more pigs were brought in such that two pigs always had the company of at least two other pigs. Pigs in the CO\(_2\)-B treatment received butorphanol 30 min prior to treatment. Pigs were euthanized in pairs. When possible, a male and female were placed into the chamber, but because there were more males, often, two males entered the chamber. Our previous experiment [12] used groups of 4, but because these pigs were larger, euthanizing in pairs provided more space to stand and walk in the chamber, which would not have been possible if 4 pigs were used. Two cameras (KPC-N502NUB, KT&C, Fairfield, NJ, USA) were positioned on two sides of the acrylic glass windows into the chamber and video was recorded using video management software (GeoVision Network Video Recorder GV-NVR, Taipei, Taiwan). The video was recorded to quantify their behavior (Table 1) and was later analyzed with a software program (Observer XT 11, Noldus, Wageningen, The Netherlands). Behaviors indicative of activity, loss of consciousness, and distress were recorded (Table 1) [12]. When that rhythmic gaping ceased the gas was turned off. After no further movement was detected, the pigs were then taken out.
of the chamber, and a lack of heartbeat confirmed that each pig was dead. The pig’s body weight and sex were then recorded.

Table 1. Behavioral states and events recorded during euthanasia. Interruptions shorter than 3 s were considered the same bout of behavior. Behavioral recording started when the gas was turned on.

<table>
<thead>
<tr>
<th>Category</th>
<th>Behavior</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Duration</td>
<td>Stand</td>
<td>Up on 4 legs</td>
</tr>
<tr>
<td></td>
<td>Lying</td>
<td>Lying down with side or sternum contact with the floor</td>
</tr>
<tr>
<td></td>
<td>Locomotion</td>
<td>Any movement more than 2 steps, walk or run</td>
</tr>
<tr>
<td></td>
<td>Inactive</td>
<td>Immobile, not doing any particular behavior</td>
</tr>
<tr>
<td>Latency</td>
<td>Ataxic</td>
<td>Lack of muscles coordination in basic movements, loss of balance on one or more feet</td>
</tr>
<tr>
<td></td>
<td>Loss of Posture</td>
<td>Lying on ground and does not get back up</td>
</tr>
<tr>
<td></td>
<td>Heaving Breathing</td>
<td>Forceful and quick repetition of flank movements, mouth closed</td>
</tr>
<tr>
<td></td>
<td>Open Mouth Breathing</td>
<td>Jaw held open with mouth open</td>
</tr>
<tr>
<td></td>
<td>Gaping</td>
<td>Deep forceful breath, rhythmic movements of the chest with mouth open</td>
</tr>
<tr>
<td></td>
<td>Last Movement</td>
<td>Clinically dead, stops gaping, end of experiment time</td>
</tr>
<tr>
<td></td>
<td>Paddle Bout</td>
<td>Rhythmic movements of one or more legs while lying</td>
</tr>
<tr>
<td></td>
<td>Panic/Convulsing Bout</td>
<td>Erratic, uncontrolled movements including flips, flops, thrashing before loss of posture</td>
</tr>
<tr>
<td>Events</td>
<td>Righting Response *</td>
<td>Unsuccessful effort to right up on 4 legs</td>
</tr>
<tr>
<td></td>
<td>Escape Attempt *</td>
<td>Rear on its hind legs, jump, or scratch with front legs against the walls or the floor</td>
</tr>
<tr>
<td></td>
<td>Squeal *</td>
<td>High-pitched vocalization; extended sound of high amplitude and frequency</td>
</tr>
<tr>
<td></td>
<td>Grunt *</td>
<td>Low-pitched vocalization; sound of low to medium amplitude.</td>
</tr>
</tbody>
</table>

* Behaviors recorded as events due to their brief nature, rather than as states.

2.3. Data Processing

The duration (the sum of time when expressing the behavior), latency (the difference between the start time and the time the event started), and event data from the behavioral software were totaled for each pig. Data for the two pigs were then averaged for each repetition prior to analyses. Vocalizations were divided by the total time, which ended as defined by the cessation of gaping. No pigs were observed to panic for any of the treatments; thus, no data were analyzed. In six instances, pigs did not paddle, and this occurred in either of the CO\textsubscript{2} treatments; thus, the sample size was not always \( n = 9 \).

2.4. Statistical Analysis

Data were analyzed using a general linear mixed model (Proc Glimmix, SAS version 9.4., SAS Institute Inc., Cary, NC, USA) and was checked for normality and homogeneity of variance prior to analysis. The treatment of euthanasia gas was included as a fixed effect with repetition as a random effect. Means of significant effects were separated with a Tukey’s adjustment. Log transformations were performed on data not meeting the statistical assumptions: Number of paddle bouts, latency to last movement, total time duration, and number of escape attempts. Influential outliers were detected with a Cook’s D test. One outlier was removed from the number of squeals. Data are presented as arithmetic means and standard error of means (means ± SE).

3. Results

The average body weight of pigs \( (p = 0.3306) \) and the average duration of gas exposure \( (p = 0.8223) \) among the three treatments did not differ (Table 2).
3.1. Duration and Latencies of Behaviors

Pigs spent a similar amount of time lying, being inactive, and in locomotion in all treatments ($p > 0.05$; Table 3). Pigs in the CO$_2$B treatment spent the shortest time standing, with pigs in CO$_2$ being intermediate and pigs in N$_2$O being the longest ($p < 0.001$; Table 3). While the total duration of heavy breathing was shortest in the CO$_2$B treatment ($p < 0.03$), the duration of open mouth breathing was shortest in the N$_2$O treatment.

Latency to heavy breathing ($p = 0.0018$) and gaping ($p < 0.001$) were shorter for CO$_2$ and CO$_2$B pigs compared to N$_2$O pigs. Latency to open-mouth breathing ($p < 0.001$) and paddling ($p < 0.001$) time was the shortest in the CO$_2$B treatment, was intermediate in the CO$_2$ treatment, and was longest in the N$_2$O treatment (Table 3). Latency to ataxia ($p = 0.0104$) and loss of posture ($p < 0.0001$) were similar for CO$_2$ and N$_2$O pigs but shorter for CO$_2$B pigs (Table 3). Figure 1 provides a graphical depiction of the sequence of behaviors, showing the latencies and durations for each behavior for each treatment.

![Figure 1](image_url)

Figure 1. The sequence of behaviors (plotted from data in Table 3), showing latencies and durations, for pigs in each treatment: CO$_2$, CO$_2$ with butorphanol, and N$_2$O. Using the means of each latency, a timeline was created to highlight the difference in the order in which the behaviors occurred. HB = heavy breathing, OMB = open-mouth breathing, LOP = loss of posture, LM = last movement. Postural behaviors standing and lying were mutually exclusive; and the activity behaviors paddling, inactivity, and locomotion were mutually exclusive.

### Table 2. Data collected (mean ± SE) on individual pigs for the three treatments.

<table>
<thead>
<tr>
<th>Variable $^1$</th>
<th>Carbon Dioxide</th>
<th>Carbon Dioxide with Butorphanol</th>
<th>Nitrous Oxide</th>
<th>$p$-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Avg. Weight (kg)</td>
<td>4.47 ± 0.38</td>
<td>4.63 ± 0.42</td>
<td>3.97 ± 0.29</td>
<td>0.3306</td>
</tr>
<tr>
<td>Total duration (sec)</td>
<td>559.44 ± 24.47</td>
<td>544.11 ± 23.81</td>
<td>556.44 ± 21.31</td>
<td>0.8223</td>
</tr>
</tbody>
</table>

$^1$ kg = kilogram, sec = second.
Table 3. Behavioral state results (mean ± SE) for multiple pairs of pigs subjected to the three treatments. Duration refers to the total elapsed time from onset to the cessation of the indicated variable. For the latency category, the numbers in parentheses indicate how many animals out of 18 in each treatment performed that specific behavior listed under column 2, “Variable”.

<table>
<thead>
<tr>
<th>Category</th>
<th>Variable(s) 1</th>
<th>Carbon Dioxide</th>
<th>Carbon Dioxide with Butorphanol</th>
<th>Nitrous Oxide</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Standing</td>
<td>123.20 ± 10.83&lt;sup&gt;b&lt;/sup&gt;</td>
<td>84.35 ± 3.35&lt;sup&gt;c&lt;/sup&gt;</td>
<td>144.55 ± 5.75&lt;sup&gt;a&lt;/sup&gt;</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td></td>
<td>Lying</td>
<td>432.28 ± 26.72</td>
<td>446.97 ± 26.34</td>
<td>407.69 ± 23.21</td>
<td>0.4508</td>
</tr>
<tr>
<td></td>
<td>Paddling</td>
<td>31.09 ± 7.51&lt;sup&gt;ab&lt;/sup&gt;</td>
<td>15.91 ± 2.92&lt;sup&gt;b&lt;/sup&gt;</td>
<td>43.35 ± 9.83&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.0294</td>
</tr>
<tr>
<td></td>
<td>Inactivity</td>
<td>423.58 ± 39.97</td>
<td>475.01 ± 26.06</td>
<td>487.80 ± 22.98</td>
<td>0.9253</td>
</tr>
<tr>
<td></td>
<td>Locomotion</td>
<td>81.62 ± 25.26</td>
<td>55.56 ± 5.37</td>
<td>63.06 ± 8.06</td>
<td>0.4564</td>
</tr>
<tr>
<td></td>
<td>Heavy Breathing</td>
<td>26.07 ± 10.63&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.50 ± 1.73&lt;sup&gt;b&lt;/sup&gt;</td>
<td>34.41 ± 9.77&lt;sup&gt;a&lt;/sup&gt;</td>
<td>&lt;0.03</td>
</tr>
<tr>
<td></td>
<td>Open Mouth Breathing</td>
<td>49.47 ± 7.26&lt;sup&gt;a&lt;/sup&gt;</td>
<td>59.35 ± 10.34&lt;sup&gt;a&lt;/sup&gt;</td>
<td>19.30 ± 8.61&lt;sup&gt;b&lt;/sup&gt;</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td></td>
<td>Heavy Breathing</td>
<td>68.37 ± 11.07&lt;sup&gt;b&lt;/sup&gt;</td>
<td>45.96 ± 11.96&lt;sup&gt;b&lt;/sup&gt;</td>
<td>164.62 ± 7.97&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.0018</td>
</tr>
<tr>
<td></td>
<td>Open Mouth Breathing</td>
<td>85.64 ± 11.58&lt;sup&gt;b&lt;/sup&gt;</td>
<td>49.69 ± 5.66&lt;sup&gt;c&lt;/sup&gt;</td>
<td>149.15 ± 19.02&lt;sup&gt;a&lt;/sup&gt;</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Latency</td>
<td>Gaping</td>
<td>145.50 ± 12.84&lt;sup&gt;b&lt;/sup&gt;</td>
<td>124.97 ± 7.56&lt;sup&gt;b&lt;/sup&gt;</td>
<td>272.50 ± 9.07&lt;sup&gt;a&lt;/sup&gt;</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td></td>
<td>Ataxia</td>
<td>103.87 ± 11.46&lt;sup&gt;a&lt;/sup&gt;</td>
<td>66.41 ± 4.24&lt;sup&gt;b&lt;/sup&gt;</td>
<td>108.99 ± 6.57&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.0104</td>
</tr>
<tr>
<td></td>
<td>Loss of Posture</td>
<td>144.99 ± 17.27&lt;sup&gt;a&lt;/sup&gt;</td>
<td>101.19 ± 4.67&lt;sup&gt;b&lt;/sup&gt;</td>
<td>162.64 ± 6.99&lt;sup&gt;a&lt;/sup&gt;</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td></td>
<td>Paddling</td>
<td>172.49 ±12.78&lt;sup&gt;b&lt;/sup&gt;</td>
<td>132.27 ± 5.96&lt;sup&gt;c&lt;/sup&gt;</td>
<td>204.84 ± 6.22&lt;sup&gt;a&lt;/sup&gt;</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

<sup>1</sup> s = second; <sup>a,b,c</sup> Means within a row with unlike superscript letters differed (p < 0.05).

3.2. Behavioral Events

The rate of righting attempts and grunts did not differ significantly between treatments (p > 0.05; Table 4). Pigs performed more escape attempts in the CO₂ B treatment than pigs in the CO₂ or N₂O treatments (p = 0.0004). Pigs in the N₂O treatment performed more paddle bouts (p = 0.0002) and squeals (p = 0.0016) than those in the CO₂ or CO₂ B treatments (Table 4).

Table 4. Behavioral event results (mean ± SE) on individual pigs for the three treatments. The numbers in parentheses indicate how many animals out of 18 in each treatment performed that specific behavior listed under column 2, “Variable”.

<table>
<thead>
<tr>
<th>Variable (Frequency) 1</th>
<th>Carbon Dioxide</th>
<th>Carbon Dioxide with Butorphanol</th>
<th>Nitrous Oxide</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Escape Attempts (14)</td>
<td>4.67 ± 1.55&lt;sup&gt;a&lt;/sup&gt;</td>
<td>16.44 ± 4.23&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.3333 ± 0.24&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.0004</td>
</tr>
<tr>
<td>Righting Attempts (16)</td>
<td>2.78 ± 0.60</td>
<td>1.11 ± 0.45</td>
<td>2.33 ± 0.75</td>
<td>0.2212</td>
</tr>
<tr>
<td>Paddle Bouts (16)</td>
<td>6.11 ± 1.09&lt;sup&gt;b&lt;/sup&gt;</td>
<td>2.11 ± 0.39&lt;sup&gt;b&lt;/sup&gt;</td>
<td>10.56 ± 1.46&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.0002</td>
</tr>
<tr>
<td>Squeals/(min) (18)</td>
<td>2.80 ± 1.17&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.28 ± 0.55&lt;sup&gt;b&lt;/sup&gt;</td>
<td>9.02 ± 1.62&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.0016</td>
</tr>
<tr>
<td>Grunts/(min) (18)</td>
<td>16.46 ± 2.04&lt;sup&gt;b&lt;/sup&gt;</td>
<td>12.29 ± 2.03&lt;sup&gt;b&lt;/sup&gt;</td>
<td>20.97 ± 5.22&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.2015</td>
</tr>
</tbody>
</table>

<sup>1</sup> min = minute; <sup>a,b</sup> Means within a row with unlike superscripts differ (p < 0.05).
4. Discussion

The results partially support our first hypothesis that using N\textsubscript{2}O at a greater flow rate than our previous study [12] achieved a comparable time to loss of posture as CO\textsubscript{2}, and it reduced escape attempts by the pigs. Nonetheless, pigs exposed to N\textsubscript{2}O performed a higher number of squeals per minute than pigs exposed to CO\textsubscript{2}. However, the results do not support our second hypothesis that giving butorphanol to the pigs prior to exposure of CO\textsubscript{2} would help us to elucidate behaviors indicative of pain because pigs given butorphanol actually made more escape attempts and the same number of squeals as CO\textsubscript{2} pigs.

The onset of loss of posture and ataxia indicates that the animal starts to lose consciousness [5,16]. However, Smith et al. [12] noted that animals could make righting attempts after the loss of posture showing that they are not unconscious; thus, it is not clear for how long after losing posture the animal can feel pain or distress. For a euthanasia method to be more humane, death should occur as soon as possible with minimal pain and distress. Therefore, shorter latency to ataxia and loss of posture is considered as an indicator of the humaneness of the method. Nevertheless, if the animal is not feeling pain or distress, then the euthanasia method would be considered humane even if the latency to loss of posture was longer. Thus, the alleviation of pain and distress is more important than the length of the euthanasia process [17].

The latency to loss of posture and ataxia did not differ between pigs in the CO\textsubscript{2} and N\textsubscript{2}O treatments. In our previous study [12], we used N\textsubscript{2}O with a 25% replacement rate and pigs had a longer latency to loss of posture. By increasing the flow rate to 50%, we were able to induce loss of posture as quickly as when CO\textsubscript{2} is used at a 25% replacement rate. These data support our hypothesis that an increased flow rate of N\textsubscript{2}O could induce unconsciousness as quickly as CO\textsubscript{2} at a 25% replacement rate.

Breathlessness is considered a very aversive experience, especially when exposed to hypercapnia [13,18]. Therefore, we consider heavy breathing and open-mouth breathing to be distressful behaviors, which are associated with compromised welfare [5,13]. Open-mouth breathing occurs just before the loss of posture when pigs were euthanized with CO\textsubscript{2} [14,19,20]. The latency to heavy breathing and open-mouth breathing started earlier in pigs exposed to CO\textsubscript{2} compared to N\textsubscript{2}O. Similarly, pigs exposed to CO\textsubscript{2} experienced this respiratory distress (HB and OMB) for a longer duration of time than pigs exposed to Ar [20]. In our study, we had similar results in terms of latency to heavy breathing between pigs of CO\textsubscript{2} and CO\textsubscript{2}B treatments; pigs in CO\textsubscript{2} had a shorter latency to respiratory distress compared to pigs in N\textsubscript{2}O. However, while there was no difference in the duration of heavy breathing between pigs in CO\textsubscript{2} and N\textsubscript{2}O, pigs in N\textsubscript{2}O had a shorter duration of open-mouth breathing compared to pigs in CO\textsubscript{2}. Because heavy breathing and open-mouth breathing are indicative of distress [13], limiting the duration of these behaviors increases the humaneness of the procedure. Thus, even though latency to heavy breathing and open-mouth breathing was shorter in the CO\textsubscript{2} treatments, N\textsubscript{2}O pigs spent less time open-mouth breathing. We contend that heavy breathing is less stressful than open-mouth breathing because the latter is an exaggerated form of the former and always follows it on the time sequence, thus it is more intense and distressful. The observed results are likely due to CO\textsubscript{2}’s main action by hypercapnia, stimulating the acid-sensing ion channels in the medulla to cause an increase in the breathing rate and depth [21]. In contrast, N\textsubscript{2}O administration does not stimulate these receptors but rather acts by hypoxia, and gases that cause these effects through hypoxia rather than hypercapnia appear to minimize distress in pigs [6].

Escape attempts are considered evidence of stress or aversion in pigs during euthanasia [14,22]. In our study, pigs in the CO\textsubscript{2} treatment had 14 times more escape attempts than pigs in the N\textsubscript{2}O treatment. On the other hand, pigs in N\textsubscript{2}O had 3.2 times more squeals than pigs in the CO\textsubscript{2} treatment. This result is similar to experiment 2 in our previous research [12]; but in contrast to experiment 1 in the same study, we found no difference in squealing. The difference in methods between the two experiments was that experiment 1 euthanized only one pig at a time whereas in the experiment 2 groups of four to six pigs were euthanized together. We also found that pigs exposed to N\textsubscript{2}O in different combinations with other gases, or by itself did not squeal (6,11) upon exposure. Each study differed
slightly, thus variables, such as the number of pigs and gas mixtures, may influence whether pigs squeal or not. Da Silva Cordeiro et al. [23] noted that animals in pain emit longer vocalizations and that it is critical to determine the duration, frequency, and amplitude to accurately determine if the vocalization is due to pain or another stressor. Consequently, we cannot be certain if the pig squeals recorded in this study were due to pain or distress (isolation from their dam and siblings) or if indeed N₂O caused more pain than CO₂. Given that N₂O is commonly used in humans for its analgesic property with no known painful response, it seems unlikely that this is a stress response, but this finding is intriguing.

There was no difference between the CO₂ treatments and the N₂O treatment in the number of righting attempts. This observation in conjunction with the fact that CO₂ pigs and N₂O pigs lost posture at similar latencies suggests that they commenced unconsciousness at similar times. Pigs in both the CO₂ and N₂O treatment made approximately two righting attempts after losing posture. Thus, they quickly lost the ability to attempt to regain a standing posture.

Paddling is uncoordinated clonic convulsions that start after the onset of loss of consciousness and when the central nervous system has lost control of the brain stem and spinal cord [24]. Similarly, gaping is deep, rhythmic, and forceful breathing movements of the jaw and paired movements of the chest, which indicates respiratory arrest and is also coordinated by the brain stem [25]. Both behaviors (gaping and paddling) are indications that the brain is becoming ‘brain dead’ [25]. Latency to gaping and paddling was shorter in pigs of the CO₂ treatments than those in the N₂O treatment. Thus, although N₂O pigs and CO₂ pigs lost posture at similar times, it seems that the CO₂ pigs became brain dead more quickly. In our previous work, we employed a lower flow rate of N₂O, and CO₂ pigs also had a quicker latency to paddling and gaping behaviors compared to N₂O pigs [12]. Therefore, the quicker flow rate could shorten the latency to the onset of loss of consciousness but not to brain death. Possibly, a new protocol using a two-step procedure in which N₂O is administered at a 50% replacement rate and then CO₂ is delivered could be efficacious.

While pigs in the CO₂ treatments showed HB, OMB, and then ataxia in consecutive order, pigs in the N₂O treatment showed ataxia first, and then showed HB and OMB consecutively. This is an important indication that the N₂O pigs are starting to lose central nervous system control prior to experiencing distress due to air hunger, possibly making the sensation of HB and OMB less distressful, although further research is needed to confirm this hypothesis.

Butorphanol was used prior to exposure to CO₂ to determine if we could identify behaviors indicative of pain, with possible candidates to include squeals and escape attempts. If painful behaviors could be identified they would be useful in future experiments when comparing alternative gas euthanasia methods. Compared to the CO₂ pigs, latency to ataxia, loss of posture, and paddling were shorter for CO₂B pigs. These results are likely due to direct suppressive effects on the central nervous system and less likely informative about the pain pigs may have experienced. Thus, butorphanol had a positive effect on shortening the time to loss of consciousness and brain death, but it remains unfeasible for use as an on-farm euthanasia method because it is not approved for use in swine and would require the presence of a veterinarian. There was no difference in latency to HB between pigs in CO₂ and CO₂B treatments, but latency to OMB was shorter for CO₂B pigs. More importantly, even though there was no difference in the duration of OMB between the CO₂ and CO₂B treatment, pigs in the CO₂B treatment had a shorter duration of HB compared to the CO₂ treatment. Butorphanol is known to have side effects, which can include respiratory depression, but this did not seem to be the case in this study since pigs in both treatments had the same duration of open-mouth breathing. The shorter duration of HB is likely due to butorphanol depressing the central nervous system while the pigs progress to loss of posture and paddling more quickly.

The number of righting attempts and squeals between the CO₂ treatments did not differ. This implies that either these behaviors are not associated with pain or that the dosage of the butorphanol was insufficient to decrease behaviors that may be indicative of pain, such as squeals. Alternatively, it could be that the stress of breathlessness is so severe that it over-rides the sensation of pain. It is unlikely that the dose was not sufficient though, as this is the dose that is recommended for swine and
proven to be a successful analgesic [26]. Similarly, the other possibility could be that the drug had not reached its full analgesic potential, but again, proven pharmacologic data suggest this would not be the problem either, with the maximal effect starting around 30 min. Interestingly, the pigs in the CO₂B treatment made more escape attempts. It is unclear why this occurred, but possibly, if the pigs in CO₂B were not suffering as much from HB and OMB, then they could focus on escaping which typically occurs when pigs are separated from their dam at weaning and put into a novel environment. The sequences of HB, OMB, ataxia, loss of posture, gaping, and paddling were similar for pigs in the CO₂ and CO₂B treatment, although typically, the CO₂B pigs entered these stages sooner than CO₂ pigs, indicating that the drug was affecting the central nervous system. Unfortunately, the compilation of these results, comparing using CO₂ with or without butorphanol, did not allow us to clearly identify behaviors indicative of pain when pigs are exposed to CO₂.

5. Conclusions

Weaned pigs euthanized with CO₂ or N₂O gases lost posture and became ataxic after a similar length of time, suggesting a similar efficacy. Pigs exposed to N₂O displayed less OMB, which is a behavior indicative of distress but also squealed more often, which suggests greater distress or pain. Pigs administered the butorphanol analgesic prior to exposure of CO₂ made more escape attempts, a similar number of squeals, and less heavy breathing but similar OMB. Whether HB relates to pain or merely to respiratory depression should be further explored.

Overall, these results show that N₂O can be as effective as CO₂, and may be more humane although further research is needed to dissociate whether behaviors, such as squeals, HB, and OMB, indicate pain or distress.

Author Contributions: D.C.L.J., J.-L.R. and R.G. conceived and designed the experiment, R.G. contributed engineering expertise and materials; and D.C.L.J. performed the experiment and analyzed the data. D.C.L.J. and E.Ç. wrote the first draft of the manuscript. All authors have read and agreed to the published version of the manuscript.

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References


