

2010

1,25-Dihydroxyvitamin D3 Enhances Bovine Mammary Epithelial Innate Immune Responses

Corwin Nelson
Iowa State University

Donald C. Beitz
Iowa State University

Timothy A. Reinhardt
United States Department of Agriculture

John Lippolis
United States Department of Agriculture

Recommended Citation

Nelson, Corwin; Beitz, Donald C.; Reinhardt, Timothy A.; and Lippolis, John (2010) "1,25-Dihydroxyvitamin D3 Enhances Bovine Mammary Epithelial Innate Immune Responses," *Animal Industry Report*: AS 656, ASL R2488.
Available at: https://lib.dr.iastate.edu/ans_air/vol656/iss1/9

This Animal Health is brought to you for free and open access by the Animal Science Research Reports at Iowa State University Digital Repository. It has been accepted for inclusion in Animal Industry Report by an authorized editor of Iowa State University Digital Repository. For more information, please contact digirep@iastate.edu.

1,25-Dihydroxyvitamin D₃ Enhances Bovine Mammary Epithelial Innate Immune Responses

A.S. Leaflet R2488

Corwin Nelson, graduate research assistant, Biochemistry, Biophysics, and Molecular Biology;
Donald Beitz, distinguished professor of animal science, Iowa State University;
Tim Reinhardt, animal scientist;
John Lippolis, molecular biologist,
USDA National Animal Disease Center

Summary and Implications

Bovine mammary epithelial cells that were treated with bacterial lipopolysaccharide and 1,25-dihydroxyvitamin D₃ showed an increase in the expression of the genes for inducible nitric oxide synthase (iNOS) and S100 calcium binding protein A12 (S100 A12). iNOS and S100 A12 are part of the innate immune response and expressed in the mammary gland during mastitis. Production of 1,25-dihydroxyvitamin D₃ in the mammary gland during mastitis, then, may be an important component of the innate immune response.

Introduction

The active form of vitamin D₃ is 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃). 1,25(OH)₂D₃ binds and activates the vitamin D receptor (VDR), a nuclear hormone receptor that regulates the expression of genes that contain a vitamin D response element in their promoter. 1,25(OH)₂D₃ is produced from 25-hydroxyvitamin D₃ (25(OH)D₃), the predominant circulating form of vitamin D₃, by the enzyme 1 α -hydroxylase (1 α -OHase). We have recently shown that bovine monocytes express 1 α -OHase in response to toll-like receptor (TLR) recognition of bacteria. Production of 1,25(OH)₂D₃ by 1 α -OHase in activated bovine monocytes increased production of nitric oxide and expression of the chemokine RANTES in monocytes. We also have observed that 1 α -OHase is expressed in inflamed mammary tissue during mastitis. Mammary epithelial cells are involved in protecting the mammary gland from bacterial infection and production of 1,25(OH)₂D₃ by 1 α -OHase in the mammary gland may affect their innate immune function. The objective of this experiment was to determine the effects of 1,25(OH)₂D₃ and lipopolysaccharide (LPS) on iNOS, RANTES, and S100 calcium binding protein A12 (S100 A12) gene expression in bovine mammary epithelial cells (MEC).

Materials and Methods

MEC were derived from mammary tissue biopsies of three Holstein cows in mid-lactation. The cells were

cultured in RPMI 1640 plus 10% fetal bovine serum and mammary epithelial growth supplement (Invitrogen). Prior to treatment, MEC were transferred to 96-well tissue culture treated plates and cultured until they were confluent. MEC were treated with 0 or 1 μ g/mL LPS along with 0, 0.1, 1 or 10 nM 1,25(OH)₂D₃ for 16 hours. After treatment, total RNA was isolated from MEC and reverse transcribed to cDNA. Quantitative real-time PCR using the 2^{- Δ ACT} method was used to measure relative abundance of 24-hydroxylase (24-OHase), iNOS, RANTES and S100 A12. Ribosomal protein S9 (RPS9) was used as the reference gene.

Results and Discussion

Treatment of MEC with 1,25(OH)₂D₃ caused an increase in 24-OHase gene expression in a dose dependent manner in both the presence and absence of LPS (Figure 1A). The 24-OHase enzyme adds a hydroxyl group to the 24 position of vitamin D metabolites, which inactivates 1,25(OH)₂D₃. In bovine monocytes, induction of 24-OHase expression by 1,25(OH)₂D₃ was repressed by LPS treatment, so regulation of 24-OHase expression differs between monocytes and MEC.

In MEC treated with LPS, both iNOS and S100 A12 gene expression increased with 1,25(OH)₂D₃ dose (Figure 1B and C). Increased expression of iNOS resulted in increased nitric oxide production by MEC (not shown). Nitric oxide may increase antimicrobial activity and blood flow in the mammary gland during mastitis, but it also may lead to MEC apoptosis. The function of S100 A12 is not completely understood, but it has been shown to have chemotactic and antimicrobial properties.

RANTES expression in MEC did not increase with 1,25(OH)₂D₃ treatment (Figure 1D). Treatment with LPS, however, caused an increase in MEC RANTES expression. In contrast, RANTES expression was up-regulated by 1,25(OH)₂D₃ treatment in LPS-stimulated bovine monocytes, but was not affected by LPS treatment alone.

In conclusion, 1,25(OH)₂D₃ acts in bovine MEC to enhance expression of iNOS and S100 A12 that is induced by TLR recognition of LPS. Therefore, production of 1,25(OH)₂D₃ by 1 α -OHase in the mammary gland during mastitis may affect the host defense capabilities of MEC. Production of 1,25(OH)₂D₃ by 1 α -OHase is dependent on availability of 25(OH)D₃. The circulating concentration of 25(OH)D₃ is dependent on acquisition of vitamin D₃ in the diet or in skin exposed to sunlight. So, dietary intake of vitamin D₃ and exposure to sun may affect incidence and severity of mastitis in cattle.

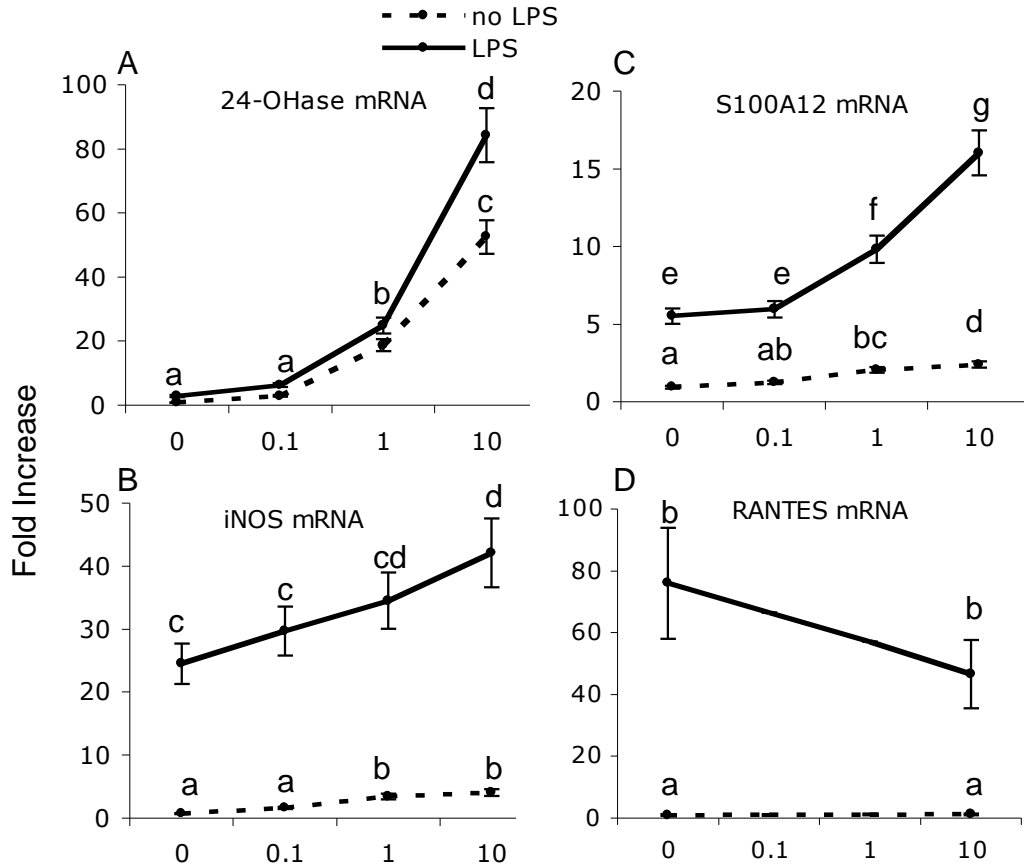


Figure 1. Mammary epithelial cell cultures were treated with 0, 0.1, 1, or 10 nM 1,25-dihydroxyvitamin D₃ for 16 hours. The amount of 24-hydroxylase (24-OHase; **A**), inducible nitric oxide synthase (iNOS; **B**), S100 calcium binding protein A12 (S100 A12; **C**), and RANTES (**D**) mRNA in MEC was measured by quantitative PCR. The expression of each gene was normalized to ribosomal protein S9 expression. The fold increase is relative to the control treatment. Results are the average of three separate experiments done in duplicate. Cells were from different cows for each experiment. Error bars represent standard error. Means with different letters are different ($P < 0.05$).