Methicillin-sensitive and Methicillin-resistant Staphylococcus aureus in pigs: (co-) colonization dynamics and clonal diversity

Fetsch, A.1, Roesler, U. 2, Kraushaar, B. 1, Friese, A. 2

Abstract

Methicillin-susceptible Staphylococcus (S.) aureus (MSSA) and methicillin-resistant S. aureus (MRSA) are colonizers of skin and mucosa. In humans, MSSA and MRSA compete for colonization space in the anterior nares and one clone can be found rather than differing types of MSSA and MRSA. We investigated the colonization dynamics and clonality of both, MSSA and MRSA in pigs over a longer time period. Eighteen sows were nasally sampled three times every ten weeks. Additionally, environmental samples were taken. Samples were investigated for MSSA and MRSA, respectively. Spa-typing was done with up to five MRSA and MSSA isolates found per sample and time point; selected isolates were further investigated by microarray. 38.9 % of sows were infrequently MSSA/MRSA co-colonized and 16.7 % were permanent carriers of MSSA. The majority of sows showed a changing colonization status. CC398 and CC9 associated spa-types were exclusively found among MRSA and MSSA, respectively. In 44.4 % of sows at least two different types of MSSA were present at the same time and sample. Strains of the same clonal lineage showed a high genetic identity despite their origin. MSSA of different spa-types but 100 % identical microarray profiles were found in sows and their environment. Our results show that pigs may be colonized with MSSA and MRSA at the same time, i.e. that MSSA/MRSA do not exclude each other in the anterior nares of pigs. Highly identical clones are present in sows and their environment, but pigs can be colonized with different clones at the same time.

Introduction

Methicillin-susceptible Staphylococcus (S.) aureus (MSSA) and methicillin-resistant S. aureus (MRSA) are colonizers of skin and mucosa and may cause severe infections both, in humans and animals. In the last decade, MRSA were frequently detected in healthy farm animals. Especially MRSA of the clonal complex (CC) 398 are widespread in the pig population in Europe (Graveland et al., 2011) and are also able to cause serious diseases in animals and humans (van der Mee-Marquet et al., 2014). Livestock professionals (farmers, veterinarians etc.) have a higher risk to get colonized with these so called livestock-associated (LA-) MRSA (van Cleef et al., 2011). In this context MSSA carriage in pig farmers as a possible protective effect against the colonization with MRSA was postulated (van Cleef et al., 2014). A multiple strain colonization of S. aureus has already been shown in humans (Votintseva et al., 2014) and in pigs (Espinosa-Gongora et al., 2014; Vandendriessche et al., 2014). We aimed to investigate the colonization dynamics of both, MSSA and MRSA in sows over a longer time period (30 weeks). Hence, the clonal diversity of MSSA and MRSA found was investigated as little is known about the clonality of MSSA, neither in pigs nor their environment while it is well described that the majority of MRSA found in pigs belong to CC398 (EFSA, 2009). Special emphasize was also given to the animals’ environment as this might have an important impact as a potential source for colonization (Broens et al., 2012).
Material and Methods

Sampling

Eighteen sows and their environment were investigated. The animals were sampled three times at intervals of about ten weeks. The sows were housed in four groups (A, B, C, and D) within one barn with possible direct contact between group A and B as well as between group C and D. One anterior nostril of every pig was sampled using a dry cotton swab (Sanstedt AG & Co. KG, Nürnberg, Germany). Additionally, in every group environmental samples from walls, toys, brushes, watering places, and troughs were collected at the 2nd and 3rd sampling time by swabbing five different locations each using the same cotton swab moistened with 1.5 ml phosphate buffered saline (PBS), together with one boot swab sample and a pooled dust sample from the whole barn.

Laboratory analyses

Isolation

Each swab was incubated in 10 ml Mueller Hinton broth with 6.5 % NaCl (MHB+) overnight at 37°C. One loop of the MHB+ was then streaked onto the S. aureus selective agar (BBL™ CHROMagar™ Staph aureus, Becton and Dickinson, USA) and incubated under aerobic conditions for 24 hours at 37°C. Another 1 ml of the MHB+ was added to 9 ml tryptone soy broth including 75 mg/l aztreonam and 3.5 mg/l cefoxitin (TSB+) to grow MRSA aerobically at 37°C for 17 hours. After that, a loop-full of TSB+ was streaked onto selective MRSA agar (CHROMagarMRSA™, MAST Diagnostica GmbH, Reinfeild, Germany) and incubated at the same conditions.

Identification and characterization of MSSA and MRSA isolates

Up to five MRSA and MSSA isolates per sample were further characterized. All susceptible isolates were phenotypically examined. Subsequently DNA of the isolates was extracted using the DNeasy Blood and Tissue kit (Qiagen, Hilden, Germany). Isolates were confirmed as MSSA and MRSA, resp., by means of an in-house multiplex-Real-Time PCR (adapted to (Fosheim et al., 2011; Kilic et al., 2010)). Spa- (Shopsin et al., 1999) and in case of MRSA, SCCmec- (Zhang et al., 2005) typing were also conducted.

To investigate the clonal identity of strains isolated from the animals and the environment, a subset of 62 MSSA and MRSA isolates was selected and investigated by means of a highly discriminatory microarray analysis (Identibac S. aureus Genotyping, Alere Technologies GmbH, Jena, Germany) according to the manufacturer’s instructions.

Results

Overall, 38.9 % of pigs were infrequently MSSA/MRSA co-colonized; 16.7 % were permanent car-riers of MSSA while the majority of pigs (66.7 %) showed a changing colonization status (Figure 1).

Discussion

From the sows of our study, 38.9 % were co-colonized with MSSA and MRSA at the same time and sample, resp. at least once during the study period. These findings do not support the hypothesis that MSSA and MRSA compete for colonization space. Others too concluded that MRSA seem to add to the burden of S. aureus rather than replacing it (Vandendriessche et al., 2014), also in humans (Cespedes et al., 2005). Spa-typing and microarray analyses showed a clear discrimination between MSSA and MRSA originating from the sows of our study. On the other hand, majority of MRSA found in pigs mainly belonged to CC9 associated spa-types. Other authors confirm these findings but did also detect CC398 associated spa-types (Hasman et al., 2010; Vandendriessche et al., 2014). On the other hand, the majority of MRSA found in pigs belong to CC398 associated spa-types (EFSA, 2009; Smith and Pearson, 2011), also in the present study. Interestingly, none of the study pigs harbored MRSA of more than one differing spa-type at the same time and over the study period, resp. In contrast, two different spa-types of MSSA were present at the same time and sample in the majority of pigs. A multiple-strain colonization in human nasal carriers of S. aureus has been shown recently (Votintseva et al., 2014). These findings call for a critical review of the number of isolates that need to be analyzed, particularly in case of in-depth epidemiological investigations.
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Identification and characterization of MSSA and MRSA isolates

To further characterize the MSSA and MRSA isolates, all susceptible isolates were phenotypically examined. Subsequently, DNA of the isolates was extracted using the DNeasy Blood and Tissue kit (Qiagen, Hilden, Germany). Isolates were confirmed as MSSA and MRSA, respectively, by means of an in-house multiplex-Real-Time PCR (adapted to (Fosheim et al., 2011; Kilic et al., 2010)). Spa- (Shopsin et al., 1999) and in case of MRSA, SCCmec- (Zhang et al., 2005) typing were also conducted.

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Molecular analysis of MSSA and MRSA isolated within our study revealed that strains detected both, at animal level and in the environment were highly identical or, in the latter case, did even show 100% homogeneity in the microarray cluster analysis. This may allow for the conclusion that clones are exchanged between pigs and their environment.

Conclusion

Pigs may be colonized with MSSA and MRSA at the same time, i.e. MSSA/MRSA do not exclude each other in the anterior nares of pigs;

Pigs can be colonized with different clones at the same time;

Highly identical MSSA and MRSA clones are present in pigs and their environment.

Acknowledgements

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A manuscript about this study (Fetsch et al.) has been submitted to the Journal Veterinary Microbiology.

References


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